

# 知母皂苷元对痴呆模型大鼠的影响

Extracellular senile plaques composed of amyloid  $\beta$ -peptide (A $\beta$ ), intracellular neurofibrillary tangles, and extensive drastic neuronal and synaptic degeneration constitute the major pathological hallmarks of Alzheimer's disease (AD). Increasing experimental as well as genetic evidence supports a causal role of A $\beta$  in the pathogenesis of AD, and the senile plaques contai- ning A $\beta$  as the core protein are often closely associated with neuronal degeneration [1]. A number of studies have shown that high concentrations of A $\beta$  are toxic and may cause damage of the biomacromolecules. A $\beta$  is capable of inducing neuronal apoptosis[2] and inhibiting cellular redox activity[3], which are believed to be dependent on the peptide assembly state such as its solution or aggregation[4]. With recent findings sugges- ting links between AD, deposition of A $\beta$  and oxidative stress[5], much attention is given currently to antioxidant research. The free-radical theory of aging[6] suggests that cumulative oxidative damage at the cellular and tissue level arises as a consequence of normal aerobic metabolism [7].

In traditional oriental medicine, quite a few herbal drugs have been used for centuries for treating disorders associated with aging, causing generally less side effects and consequently ensuring safe application. Ginkgo biloba, whose existence can be date back to the time of the dinosaurs, is one of the oldest tree still thriving on the planet [8]. In European countries, the extract of Ginkgo biloba is prescribed for the treatment of peripheral and cerebral blood insufficiency. Experimental evidence suggests that this extract inhibits blood coagulation[9], attenuates age-related deterioration of cognitive fun- ctions in rats[10], and improves degenerative dementias of the Alzheimer's and multi-infarct types [11] [12]. Existing pharmacological data and clinical trials have demon- strated Ginkgo biloba extract EGb 761 to be a potent antioxidant agent and free radical scavenger with neuroprotective effects [13] [14] [15][16]. Anemarrhena asphodeloides Bge, a traditional Chinese herbal drug, is derived from the dry rhizoma of the plant Anemarrhena aspho- deloides, a species of Liliaceae. Anemarrhena saponins and timosaponins are the main active components of this drug. Sapogenins from Anemarrhean asphodeloides was reported to improve the learning and memory abilities with also effect to increase the levels of brain- derived neurotrophic factor (BDNF) in aged rats[17]. In addition, our previous study showed that timosaponins could increase cellular level of nicotinic acetylcholine receptors in rat brains in a dose-dependent manner [18]. Timosaponin also inhibits N-formyl-methionyl-leucyl- phenylalanine (fMLP)-induced generation of superoxide in leukocytes[19] in vitro and lower blood glucose con- centration[20][21]. Recent study suggests that timosaponin is capable of some estrogen-like actions<sup>[22]</sup> to serve as an important protective agent against AD[23].

The A $\beta$  peptide fragment A $\beta$ (25-35) has been shown to be directly toxic to neurons and able to increase the vulnerability of neurons to other insults[24]. According to Varadarajan et al

[25], this truncated peptide is more rapidly toxic and causes more oxidative damage than A $\beta$  (1-42). Additionally, Giovannelli et al reported the onset of impairment of object recognition in the first two weeks after A $\beta$  (25-35) injection at the nucleus basalis in rats, whereas A $\beta$  (1-40) impaired the performance till two months after the injection[26]. This study was designed to determine whether timosaponins (timosaponin B-II, timosaponin E1 and timosaponin B) may antagonize the toxicity of A $\beta$ (25-35) in a rat model of learning and memory dysfunction induced by microinjection of A $\beta$  (25-35) into the lateral cerebral ventricles.

### MATERIALS AND METHODS

# Rats

Totally 60 adult male Sprague-Dawley rats with body weight ranging from 200 to 250 g were purchased from the Experimental Animal Center, Southern Medical University (certificate No. 2003A073). The rats were kept in cages (5 in each cage) with the temperature controlled at 24  $\pm 2$  °C, relative humidity of (60 $\pm$ 10)%, and in 12-hour light-dark cycles. Free access to food in the form of dry pellets and water was allowed. The animal experiments were performed according to the internationally accepted ethical guidelines.

Drugs

The timosaponins were extracted from Anemarrhena aspholeloides Bunge (purchased from Hebei Province, China) in our laboratory. The timosaponins, with the main components of timosaponin B-II, timosaponin E1 and timosaponin B, were dissolved in sterile distilled water at different concen- trations. Ginkgo biloba extract EGB761 serving as a positive therapeutic control drug was purchased from Cerenin, Dr. Willmar Schwabe Co., (Germany), also suspended with sterile distilled water. A $\beta$  (25-35) (Sigma Chemical Co, USA) was dissolved in sterile distilled water, and stored at -20 °C. To obtain the neurotoxic form of A $\beta$ (25-35), the peptide solution was incubated at 37 °C for 7 days [26]. Superoxide dismutase (SOD), malondialdehyde (MDA), total antioxidation capacity and protein content test kits were purchased from Nanjing Jiancheng Bioengineering Institute (Nanjing, China).

Induction of learning and memory dysfunction

The rats were anesthetized with 10% chloral hydrate (0.35g/kg) and fixed in a stereotaxic apparatus for microinjection of A $\beta$  (25-35) into the lateral cerebral ventricle. The bregma and the skull surface served as the stereotaxic zero points [27][28]. The dura mater was surgically exposed, and the needle was lowered into the right lateral ventricle [posterior (P) 1.3mm, lateral (L) 1.3 mm, ventral (V) 4.0 mm][29], where 1  $\mu$ l A $\beta$  (25-35) (10 mmol/L) or sterile distilled water was slowly infused within 5 min, and the syringe was not withdrawn until 5 min after the injection. The therapeutic drug administration was performed 24 h after the surgery, once a day for 14 consecutive days.

Animal grouping

The rats were randomly divided into 6 groups to receive intracerebral ventricular injection (i.c.v.) and intragastric administration (i.g.) of distilled water (control group), i.c.v. of A $\beta$  (25-35) with i.g. of distilled water (dementia model group), i.c.v. of A $\beta$  (25-35) and i.g. of EGB761 (EGB761 group), or i.c.v. of A $\beta$  (25-35) and i.g. of three doses (50,100 and 200 mg/kg, designed according to the results of our previous study) of timosaponins (treatment groups).

#### Morris water maze test

The rats were trained in Morris water maze test for 7 consecutive days from days 8 to 14

following A $\beta$  (25-35) injection. The water maze apparatus consisted of a circular pool filled with water 30 cm in depth, and a circular platform supported by a base resting on the bottom of the pool, 2 cm below the surface of the water. The water was mixed with milk for the coloring and the rat heads were stained black. The platform resided in the center of the northwest quadrant. At the start of the test, the rats were made to face the wall, and when they managed to locate the platform, they were allowed to remain on it for 15 s. If the rats failed to locate the platform within 2 min, they were removed from the water and placed on the platform for 15 s. Data collection was automated by an on-line video tracking device designed to track the object with the highest contrast, which rendered only the black heads of the rats visible against the milk-white background. The escape latency (time to locate the platform) and swimming path were recorded.

Assay of SOD, total antioxidation capacity and MDA

Two weeks after distilled water and A $\beta$  (25-35) injection and corresponding treatment, the rats were killed by decapitation and the brain tissue was taken to prepare 10% tissue homogenate by addition of 0.86% cold saline (pH 7.4) and stored at -30  $\degree$  for later examination. Total antioxidation capacity and MDA assays were performed according to the instructions of the test kits and the tissue homogenate was diluted to 2% for SOD test. MDA was quantified by thiobarbituric (TBA) method to measure the absorbance at 532 nm (expressed as nmol/mg.protein). In SOD assay, xanthine oxidase was used as a superoxide generator to reduce nitroblue tetrazolium (NBT), which was inhibited by SOD present in the solution. This inhibition of NBT reduction into blue-colored formozan in the presence of phenazine metha sulphate (PMS) and NADH was measured at 560 nm. One unit of enzyme activity was defined as the amount of enzyme that inhibited the rate of reaction by 50% in 1 min under the defined assay conditions and the result expressed as units (U) of SOD activity/mg.protein. The antioxidant defense system consisted of enzymatic and non-enzymatic antioxidants, which were able to reduce  $Fe^{3+}$  to  $Fe^{2+}$ . The total antioxidation capacity was measured by the reaction of phenanthroline and Fe<sup>2+</sup> using a spectrophotometer at 520 nm. At 37 °C, a total antioxidation capacity unit was defined as the amount of antioxidant required to produce an absorbance increase of 0.01 in 1 mg protein.

Statistical analyses

The data were presented as Mean $\pm$ SD. The data between the model group and the control group were analyzed with independent-sample t test, and those between the model group, the treatment groups and EGB761 group were analyzed with one-way ANOVA (least-significant difference test) using SPSS 10.0 software. A P value less than 0.05 was considered to indicate statistically significant difference.

#### RESULTS

Effect of timosaponins on learning and memory dysfunction (Morris water maze test) At the beginning of the training, most of the swimming tracks of the rats presented a random pattern near the edge of the pool, instead of a shorter linear path. Towards the end of the 7-day training, most of the rats, with the exception of the rats in the model group, managed to find the platform through a roughly linear path. Typical tracks of the rats in the control, model and treatment groups in the Morris water maze are shown in Fig. 1. Apparently, the mean latencies were greatly longer in the model group than in the control group, or in other words, A $\beta$  (25-35)-treated rats took longer time to find the platform than the control rats did. Treatment with timosaponins and Ebg761 for 14 days could significantly shorten the latencies, which showed no significant difference between Egb761 and treatment groups (Tab.1).



d=749.079 cm, t=60.68 s d= 495.533 cm, t=26.82 s d=219.563 cm, t=13.88 s



d=913.219 cm, t=60.76 s d=433.923 cm, t=23.54 s d=252.737 cm, t=14.48 s



d=1 277.926 cm, t=66.19 s d=747.089 cm, t=33.27 s d=360.402 cm, t=18.92 s

- Fig.1 Typical examples of tracks made by the rats in the 4 groups in the Morris water maze test
  - A: Control group; B: Treatment group (100 mg/kg); C: Dementia model group. t: Latent period; d: Distance traveled

Tab.1	Latencies of	the rat	s to	reach	the	platform	in
-------	--------------	---------	------	-------	-----	----------	----

Morris water maze test					
Group	n	Latency (Mean±SD)			
Control	8	26.01±4.79**			
Model	8	33.59±8.29			
Egb761 (15.5 mg/kg)	8	26.04±5.55#			
Timosaponins (mg/kg)					
50	9	24.34±5.13**			
100	8	25.70±4.11#			
200	8	24.84±5.45**			

\*P<0.05, \*\*P<0.01 vs model group. The data between the model and the control groups were analyzed with independent-sample t test, and those between the model, treatment, and EGB761 groups with one-way ANOVA (LSD) by SPSS 10.0 software. SOD activity, total antioxidation capacity and MDA level in brain tissue After treatment with A $\beta$  (25-35), the activity of SOD and total antioxidation capacity in the rat brain tissues were lowered and MDA level increased compared with those in the control rats. Compared with the model group, the rats receiving timosaponins and Egb761 therapies showed remarkably increased SOD activity and total antioxidation capacity as well as reduced MDA level in the brain tissue (Tab. 2).

Group	п	SOD (U/mg,pro)	MDA (nmol/mg.pro)	Total antioxidation capacity (U/mg.pro)
Control	9	302.41±43.46**	4.60±1.09#	8.37±1.67**
Model	10	229.75±34.13	5.95±1.37	$2.75 \pm 1.18$
Egb761 (15.5 mg/kg)	9	298.51±45.40**	4.80±1.11"	4.50±2.30#
Timosaponins (mg/kg)				
50		275.53±31.90"	4.94±0.69"	$4.19 \pm 1.04$
100	10	277.23±63.55*	4.67±0.95**	8.39±2.37**
200	10	276.25±32.57"	4.85±0.77 <sup>#</sup>	9.48±1.94**

Tab.2 SOD activity, MDA level and total antioxidation capacity in the brain tissue of rats (Mean±SD)

\*P<0.05, \*\*P<0.01 vs model group. Statistical analysis of the data is specified in Tab.1.

# DISCUSSION

The results of this study demonstrated that a single acute injection of A $\beta$  (25-35) into the lateral cerebral ventricle induced marked amnesic effect in rats as shown by the repeated training of the rats for spatial recognition in Morris water maze, one of the most widely employed paradigms to assess learning and memory function in rodents [30]. A $\beta$  (25-35), as we observed, induced impairment of the learning and memory abilities of the rats, and the rats in model group tended to circle around the pool whereas the control rats swam more directly to reach the platform in the later training days. Excessive oxygen-derived free radicals have been recognized for their potential to damage the neurons and reduce the density of neurons in the brain, leading to deteriorated learning and memory capacity in animals [31]. The central nervous system is especially vulnerable to free radical damage due to the high demand of oxygen consumption of the brain, its abundant lipid content, and the relative paucity of antioxidant enzymes compared with other tissues [32]. A $\beta$  in the brain of AD patient may promote the production of free radicals, which at least partially mediated the cytotoxic effects of AB on the neurons [33]. Some studies have also found that A $\beta$  is directly responsible for freeradical damage to the neuronal membrane systems, leading to subsequent neuronal loss in the AD brain[34]. Free radical attack on phospholipid polyunsaturated fatty acids (PUFA) can ultimately lead to the production of multiple aldehydes with different carbon chain length, including MDA[35]. In a sense, the level of MDA can be taken as an indicator for the state of lipid peroxidation. SOD converts superoxide radicals to H2O2, which is in turn broken down to water and oxygen in the presence of catalases. Thus, SOD and the catalases constitute the first defense mechanism against reactive oxygen species. A decrease in the activity of the antioxidant system would most likely lead to cellular death. Marcus et al[36] observed

significantly decreased SOD activity in frontal and AD temporal cortex. Our present study showed that in rats treated with A $\beta$  (25-35), the activity of SOD and the total antioxidative capacity decreased and the level of MDA increased, suggesting that A $\beta$  (25-35) could induce the overproduction of free radicals and this free radical metabolism disorder caused impairment of the brain function.

As the extract of a herbal drug with proven efficacy and safety, timosaponins, according to the results of this present study, resulted in a significant improvement of learning and memory capacity of rats treated with A $\beta$  (25-35) with such potency as comparable to that of EGB761. At the same time, timosaponins significantly enhanced SOD activities and total antioxidation capacity and decreased MDA level, similar to the effect of EGB761. These results demons- trate that timosaponins can remarkably enhance the learning and memory capacities, presumably through promoting scavenging of the free radicals. We had presumed that timosaponins possessed such potential to act on multiple targets, and our previous study also showed that timosaponins could ameliorate the dysfunction of the cholinergic system[18]. Timosaponins may still have some other potentials as intervening in cell apoptosis, a probability that is currently under investigation.

REFERENCES

[1] Selkoe DJ. The molecular pathology of Alzheimer's disease[J]. Neuron, 1991, 6(4): 487-98.

[2] Loo DT, Copani A, Pike CJ, et al. Apoptosis is induced by  $\beta$ -amyloid in culture central nervous system neurons [J]. Proc Natl Acad Sci USA , 1993, 90(17): 7951-5.

[3] Shearman MS, Ragan CI, Iversen LL. Inhibition of PC12 cell redox activity is a specific, early indicator of the mechanism of  $\beta$ -amyloid- mediated cell death[J]. Proc Natl Acad Sci USA , 1994, 91(4): 1470-4.

[4] Pike CJ, Burdick D, Walencewicz AJ, et al. Neurodegeneration induced by  $\beta$ -amyloid peptides in vitro: the role of peptide assembly state[J]. J Neurosci, 1993, 13(4): 1676-87.

 [5] Yatin SM, Varadarajan S, Butterfield DA. Vitamin E prevents Alzhei- mer's amyloid beta-peptide (1-42)-induced neuronal protein oxidation and reactive oxygen species production
[J]. Alzheimer Dis Assoc Disord, 2000, 2(2): 123-31.

[6] Harman D. Ageing: a theory based on free radical and radiation chemistry [J]. Gerontol, 1957(1): 298-300.

[7] Biesalski HK. Free radical theory of aging[J]. Curr Opin Clin Nutr Metab Care, 2002, 5(1): 5-10.

[8] DeFeudis FV, Drieu K. Ginkgo biloba extract (EGb 761) and CNS functions: basic studies and clinical applications[J]. Curr Drug Targets, 2000, 1(1): 25-58.

[9] Smith PF, Maclennan K, Darlington CL. The neuroprotective pro- perties of the Ginkgo biloba leaf: a review of the possible rela- tionship to platelet-activating factor (PAF)[J]. J Ethnopharmacol, 1996, 50(3): 131-9.

[10] Wirth S, Stemmelin J, Will B, et al. Facilitative effects of EGb 761 on olfactory recognition in young and aged rats[J]. Pharmacol Bio- chem Behav, 2000, 65(2): 321-6.

[11] Le Bars PL, Katz MM, Berman N, et al. A placebo-controlled, double-blind, randomized trial of an extract of Ginkgo biloba for dementia. North American EGb Study Group[J]. JAMA, 1997, 278(16): 1327-32.

[12] Le Bars PL, Kieser M, Itil KZ. A 26-week analysis of a double-blind, placebocontrolled trial of the ginkgo biloba extract EGb 761 in dementia[J]. Dement Geriatr Cogn Disord, 2000, 11(4): 230-37.

[13] Christen Y, Maixent JM. What is Ginkgo biloba extract EGb 761? An overview - from molecular biology to clinical medicine[J]. Cell Mol Biol (Noisy-le-grand), 2002, 48(6): 601-

[14] DeFeudis FV, Drieu K. Ginkgo biloba extract (EGb 761) and CNS functions: basic studies and clinical applications[J]. Curr Drug Targets 2000, 1(1): 25-58.

[15] Luo Y. Ginkgo biloba neuroprotection: Therapeutic implications in Alzheimer's disease[J]. J Alzheimers Dis, 2001, 3(4): 401-7.

[16] Sastre J, Lloret A, Borras C, et al. Ginkgo biloba extract EGb 761 protects against mitochondrial aging in the brain and in the liver[J]. Cell Mol Biol (Noisy-le-grand), 2002, 48 (6): 685-92.

[17] 胡雅儿,孙启祥,夏宗勤. ZMS对老年大鼠脑内NGF和BDNF的影响[J]. 中国药理学通报, 2003, 19 (2): 149-51.

Hu YE, Sun QX, Xia ZQ. The effect of ZMS, an active component of Zhimu, on NGF and BDNF in brains of aged rats[J]. Chin Pharmacol Bull, 2003, 19 (2): 149-51.

[18] 徐江平. 知母皂苷对衰老大鼠M、N胆碱受体的调节作用[J]. 中国老年学杂志, 2001, 21: 379-80.

Xu JP. Regulation of saponins from Anemarrhean Asphodeloides Bge on cerebral M, Nacetylchol in receptors in aged rats[J]. Chin J Gerontol, 2001, 21: 379-80.

[19] Kaname N, Zhang J, Meng Z, et al. Effect of timosaponin E1 and E2 on superoxide generation induced by various stimuli in human neutrophils and on platelet aggregation in human blood[J]. Clin Chim Acta, 2000, 295(1-2): 129-40.

[20] Hoa NK, Phan DV, Thuan ND, et al. Insulin secretion is stimulated by ethanol extract of Anemarrhena asphodeloides in isolated islet of healthy Wistar and diabetic Goto-Kakizaki rats[J]. Exp Clin Endocrinol Diabetes, 2004, 112(9):520-5.

[21] Miura T, Ichiki H, Iwamoto N, et al. Antidiabetic activity of the rhizoma of Anemarrhena asphodeloides and active components, mangiferin and its glucoside[J]. Biol Pharm Bull, 2001, 24(9): 1009-11.

[22] 孟志云, 李 文, 徐绥绪, 等. 知母的皂苷成分[J]. 药学学报, 1999, 34(6): 451-3.

Meng ZY, Li W, Xu SX, et al. Saponins from rhizomes of Ane- marrhena asphodeloides Bge[J]. Yao Xue Xue Bao, 1999, 34(6): 451-3.

[23] Zhang Y, Champagne N, Beitel LK, et al. Estrogen and androgen protection of human neurons against intracellular amyloid beta1-42 toxicity through heat shock protein 70[J]. J Neurosci, 2004, 24(23): 5315-21.

[24] Mattson MP, Cheng B, Davis D, et al.  $\beta$ -amyloid peptides desta- bilize calcium homeostasis and render human cortical neurons vulnerable to excitotoxicity[J]. J Neurosci, 1992, 12(2): 376-89.

[25] Varadarajan S, Kanski J, Aksenova M, et al. Differen mechanism of oxidative stress and neurotoxicity for Alzheimer's A beta (1-42) and A beta (25-35)[J]. J Am Chem Soc, 2001, 123(24): 5625-31.

[26] Giovannelli L, Casamenti F, Scali C, et al. Differential effects of amyloid peptides beta-(1-40) and beta-(25-35) injections into the rat nucleus basalis[J]. Neuroscience, 1995, 66(4): 781-92.

[27] Nabeshima T, Nitta A. Memory impairment and neuronal dysfunc- tion induced by betaamyloid protein in rats[J]. Tohoku J Exp Med, 1994, 174(3): 241-9.

[28] Frautschy SA, Yang F, Calderon L. Rodent models of Alzheimer's disease: rat Abeta infusion approach to amyloid deposits[J]. Neuro- biol Aging, 1996, 17(2): 311-21.

[29] Paxinos G, Watson C. The rat brain in stereotaxic coordinates[M]. Sydney: Academic Press, 1982. 22.

[30] Morris R. Developments of a water-maze procedure for studying spa- tial learning in the rat[J]. J Neurosci Methods, 1984, 11(1): 47-60.

[31] Song X, Bao M, Li D. Advanced glycation in D-galactose induced mouse aging model[J].

11.

J Horm Metab Res, 1999, 31(3): 278-82.

[32] Coyle JT, Puttfarcken P. Stress, glutamate and neurodegenerative disorders[J]. Science, 1993, 262(5134): 689-95.

[33] Butterfield DA, Lauderback CM. Lipid peroxidation and protein oxidation in Alzheimer's disease brain: potential causes and con- sequences involving amyloid beta-peptide-associated free radical oxidative stress[J]. Free Radic Biol Med, 2002, 32(11): 1050-60.

[34] Varadarajan S, Yatin S, Aksenova M, et al. Review: Alzheimer's amyloid betapeptide-associated free radical oxidative stress and neurotoxicity[J]. J Struct Biol, 2000, 130(2-3): 184-208.

[35] Esterbauer H, Schaur R, Zollner H. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde, and related aldehydes[J]. Free Radic Biol Med, 1991, 11(1): 81-128.

[36] Marcus DL, Thomas C, Rodriguez C, et al. Increased peroxidation and reduced antioxidant enzyme activity in Alzheimer's disease[J]. Exp Neurol, 1998, 150(1): 40-4.

REFERENCES

[1] Selkoe DJ. The molecular pathology of Alzheimer's disease[J]. Neuron, 1991, 6(4): 487-98.

[2] Loo DT, Copani A, Pike CJ, et al. Apoptosis is induced by  $\beta$ -amyloid in culture central nervous system neurons [J]. Proc Natl Acad Sci USA , 1993, 90(17): 7951-5.

[3] Shearman MS, Ragan CI, Iversen LL. Inhibition of PC12 cell redox activity is a specific, early indicator of the mechanism of  $\beta$ -amyloid- mediated cell death[J]. Proc Natl Acad Sci USA , 1994, 91(4): 1470-4.

[4] Pike CJ, Burdick D, Walencewicz AJ, et al. Neurodegeneration induced by  $\beta$ -amyloid peptides in vitro: the role of peptide assembly state[J]. J Neurosci, 1993, 13(4): 1676-87.

 [5] Yatin SM, Varadarajan S, Butterfield DA. Vitamin E prevents Alzhei- mer's amyloid beta-peptide (1-42)-induced neuronal protein oxidation and reactive oxygen species production
[J]. Alzheimer Dis Assoc Disord, 2000, 2(2): 123-31.

[6] Harman D. Ageing: a theory based on free radical and radiation chemistry [J]. Gerontol, 1957(1): 298-300.

[7] Biesalski HK. Free radical theory of aging[J]. Curr Opin Clin Nutr Metab Care, 2002, 5(1): 5-10.

[8] DeFeudis FV, Drieu K. Ginkgo biloba extract (EGb 761) and CNS functions: basic studies and clinical applications[J]. Curr Drug Targets, 2000, 1(1): 25-58.

[9] Smith PF, Maclennan K, Darlington CL. The neuroprotective pro- perties of the Ginkgo biloba leaf: a review of the possible rela- tionship to platelet-activating factor (PAF)[J]. J Ethnopharmacol, 1996, 50(3): 131-9.

[10] Wirth S, Stemmelin J, Will B, et al. Facilitative effects of EGb 761 on olfactory recognition in young and aged rats[J]. Pharmacol Bio- chem Behav, 2000, 65(2): 321-6.

[11] Le Bars PL, Katz MM, Berman N, et al. A placebo-controlled, double-blind, randomized trial of an extract of Ginkgo biloba for dementia. North American EGb Study Group[J]. JAMA, 1997, 278(16): 1327-32.

[12] Le Bars PL, Kieser M, Itil KZ. A 26-week analysis of a double-blind, placebocontrolled trial of the ginkgo biloba extract EGb 761 in dementia[J]. Dement Geriatr Cogn Disord, 2000, 11(4): 230-37.

[13] Christen Y, Maixent JM. What is Ginkgo biloba extract EGb 761? An overview - from molecular biology to clinical medicine[J]. Cell Mol Biol (Noisy-le-grand), 2002, 48(6): 601-

[14] DeFeudis FV, Drieu K. Ginkgo biloba extract (EGb 761) and CNS functions: basic studies and clinical applications[J]. Curr Drug Targets 2000, 1(1): 25-58.

[15] Luo Y. Ginkgo biloba neuroprotection: Therapeutic implications in Alzheimer's disease[J]. J Alzheimers Dis, 2001, 3(4): 401-7.

[16] Sastre J, Lloret A, Borras C, et al. Ginkgo biloba extract EGb 761 protects against mitochondrial aging in the brain and in the liver[J]. Cell Mol Biol (Noisy-le-grand), 2002, 48 (6): 685-92.

[17] 胡雅儿,孙启祥,夏宗勤. ZMS对老年大鼠脑内NGF和BDNF的影响[J]. 中国药理学通报, 2003, 19 (2): 149-51.

Hu YE, Sun QX, Xia ZQ. The effect of ZMS, an active component of Zhimu, on NGF and BDNF in brains of aged rats[J]. Chin Pharmacol Bull, 2003, 19 (2): 149-51.

[18] 徐江平. 知母皂苷对衰老大鼠M、N胆碱受体的调节作用[J]. 中国老年学杂志, 2001, 21: 379-80.

Xu JP. Regulation of saponins from Anemarrhean Asphodeloides Bge on cerebral M, Nacetylchol in receptors in aged rats[J]. Chin J Gerontol, 2001, 21: 379-80.

[19] Kaname N, Zhang J, Meng Z, et al. Effect of timosaponin E1 and E2 on superoxide generation induced by various stimuli in human neutrophils and on platelet aggregation in human blood[J]. Clin Chim Acta, 2000, 295(1-2): 129-40.

[20] Hoa NK, Phan DV, Thuan ND, et al. Insulin secretion is stimulated by ethanol extract of Anemarrhena asphodeloides in isolated islet of healthy Wistar and diabetic Goto-Kakizaki rats[J]. Exp Clin Endocrinol Diabetes, 2004, 112(9):520-5.

[21] Miura T, Ichiki H, Iwamoto N, et al. Antidiabetic activity of the rhizoma of Anemarrhena asphodeloides and active components, mangiferin and its glucoside[J]. Biol Pharm Bull, 2001, 24(9): 1009-11.

[22] 孟志云, 李 文, 徐绥绪, 等. 知母的皂苷成分[J]. 药学学报, 1999, 34(6): 451-3.

Meng ZY, Li W, Xu SX, et al. Saponins from rhizomes of Ane- marrhena asphodeloides Bge[J]. Yao Xue Xue Bao, 1999, 34(6): 451-3.

[23] Zhang Y, Champagne N, Beitel LK, et al. Estrogen and androgen protection of human neurons against intracellular amyloid beta1-42 toxicity through heat shock protein 70[J]. J Neurosci, 2004, 24(23): 5315-21.

[24] Mattson MP, Cheng B, Davis D, et al.  $\beta$ -amyloid peptides desta- bilize calcium homeostasis and render human cortical neurons vulnerable to excitotoxicity[J]. J Neurosci, 1992, 12(2): 376-89.

[25] Varadarajan S, Kanski J, Aksenova M, et al. Differen mechanism of oxidative stress and neurotoxicity for Alzheimer's A beta (1-42) and A beta (25-35)[J]. J Am Chem Soc, 2001, 123(24): 5625-31.

[26] Giovannelli L, Casamenti F, Scali C, et al. Differential effects of amyloid peptides beta-(1-40) and beta-(25-35) injections into the rat nucleus basalis[J]. Neuroscience, 1995, 66(4): 781-92.

[27] Nabeshima T, Nitta A. Memory impairment and neuronal dysfunc- tion induced by betaamyloid protein in rats[J]. Tohoku J Exp Med, 1994, 174(3): 241-9.

[28] Frautschy SA, Yang F, Calderon L. Rodent models of Alzheimer's disease: rat Abeta infusion approach to amyloid deposits[J]. Neuro- biol Aging, 1996, 17(2): 311-21.

[29] Paxinos G, Watson C. The rat brain in stereotaxic coordinates[M]. Sydney: Academic Press, 1982. 22.

[30] Morris R. Developments of a water-maze procedure for studying spa- tial learning in the rat[J]. J Neurosci Methods, 1984, 11(1): 47-60.

[31] Song X, Bao M, Li D. Advanced glycation in D-galactose induced mouse aging model[J].

11.

J Horm Metab Res, 1999, 31(3): 278-82.

[32] Coyle JT, Puttfarcken P. Stress, glutamate and neurodegenerative disorders[J]. Science, 1993, 262(5134): 689-95.

[33] Butterfield DA, Lauderback CM. Lipid peroxidation and protein oxidation in Alzheimer's disease brain: potential causes and con- sequences involving amyloid beta-peptide-associated free radical oxidative stress[J]. Free Radic Biol Med, 2002, 32(11): 1050-60.

[34] Varadarajan S, Yatin S, Aksenova M, et al. Review: Alzheimer's amyloid betapeptide-associated free radical oxidative stress and neurotoxicity[J]. J Struct Biol, 2000, 130(2-3): 184-208.

[35] Esterbauer H, Schaur R, Zollner H. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde, and related aldehydes[J]. Free Radic Biol Med, 1991, 11(1): 81-128.

[36] Marcus DL, Thomas C, Rodriguez C, et al. Increased peroxidation and reduced antioxidant enzyme activity in Alzheimer's disease[J]. Exp Neurol, 1998, 150(1): 40-4.

# 回结果列表