

Enzymatic synthesis of a CCK-4 tripeptide fragment

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Abstract: Objective To synthesize tripeptide derivative Phac-Met-Asp(OMe)-Phe-NH₂, which is a fragment of the gastrin C-terminal tetrapeptide CCK-4, by enzymatic reaction. **Methods** Three free enzymes, chymotrypsin, papain and thermolysin from acyl donor Phac-Met-OCam was involved in three steps. The choice of appropriate enzymes and solvents was selected. **Results** All enzymatic reactions were obtained in reasonable yields (63%-92%). FAB-MS and FD-MS verified the correct molecular mass of the peptides. **Conclusion** Studies on the chymotrypsin catalyzed coupling reaction between Phac-Met-OCam and H-Asp(OMe)₂ have focused on the low water content media. By papain catalyzed saponification of Phac-Met-Asp(OMe)₂, methyl ester of aspartic acid is selectively hydrolyzed to retain methyl ester, and Phac-Met-Asp(OMe)-OH and H-Phe-NH₂ can be coupled efficiently by thermolysin.

Key words: enzymatic synthesis; peptide bond; gastrin CCK-4; tripeptide derivative

Enzymatic approach has proved to be a promising alternative to chemical methods in peptide synthesis^[1-4] in regard to its numerous advantages over the chemical ones, such as the promotion of peptide bond formation under mild conditions, minimum side-chain protection, and elimination of racemization. The use of harmful organic reagents, in addition, is totally avoided to justify the technical applications of this approach.

It is important and difficult to choose from the reaction and separation conditions when total enzymatic or enzymochemical synthetic strategies are employed to produce moderate-length to large peptide. This fact is particularly significant when the convergent strategy is adopted because of the great number of the possibilities involved therein that await careful evaluation.

This investigation is designed to test the possibility of synthesizing peptides exclusively by enzymatic methods with reasonable yields. As the peptide for testing this synthesis approach, the tripeptide fragment of gastrin CCK-4 was chosen that has the same C-terminal sequence as that of gastrin and also displays the same physiological activities as the latter^[5].

Enzymatic synthesis of the tripeptide derivative Phac-Met-Asp(OMe)-Phe-NH₂ is reported in this paper. The phenylacetyl group (Phac) is used as the protection group for the amino group, and can be cleaved at the

end of the synthesis with penicillin G amidase without affecting the peptide bonds^[6]. Thus, beginning with Phac-Met-OCam (carboxamidomethyl ester), the target tripeptide derivative was synthesized successfully with 3 free enzymes, chymotrypsin, papain and thermolysin, and all reactions were carried out in reasonable yields (Fig. 1). The key steps in the synthesis of the tripeptide were the coupling of Phac-Met-OCam and H-Asp(OMe)₂ to form Met-Asp peptide bond catalyzed by chymotrypsin and the selective enzymatic hydrolysis of methyl ester of Phac-Met-Asp(OMe)₂ with papain.

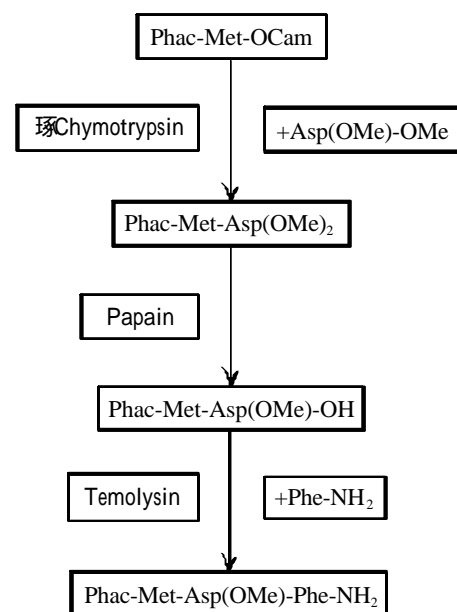


Fig. 1 Enzymatic synthesis of CCK-4 tripeptide fragment

MATERIALS AND METHODS

Materials

Chymotrypsin (EC3.4.21.1) from bovine pancreas (Type 域 crystallized for 3 times from chymotrypsinogen that had been crystallized for 4 times, dialyzed essentially into salt-free and lyophilized powder, with activity 40-60 U/mg of protein), papain (EC3.4.22.2) from *Carica papaya* (twice crystallized, lyophilized powder, 10-20 U/mg of BAE assay), and thermolysin (EC3.4.24.2) from *Bacillus thermoproteolyticus* rokko (crystallized and lyophilized powder containing calcium and sodium buffers salts, 50-100 U/mg of protein) were all from Sigma (USA). The amino acid derivatives, Phac-Met-OCam, H-Asp(OMe)-OMe and H-Phe-NH₂ HCl were synthesized in our laboratory by a standard procedure. All other reagents and solvents used in this study were of analytical grade.

Analysis with high-performance liquid chromatography (HPLC)

HPLC (Gilson), Column: Nucleosil 100RP-18, 5 μ m and 100 mm \times 4 mm column (Macherey-Nagel). Mobile Phase (System 玉): Solvent A, 0.05 mol/L HCOONH₄ (pH 6.5); solvent B, 80% (V/V) MeOH and 20% water, elution gradient from 45% to 85% B; Mobile Phase (System II): Solvent A, H₂O (0.1% TFA); solvent B, 80% Acetonitril (0.1% TFA), elution gradient from 30% to 70% B; Flow rate 0.3 ml/min; UV detection at 260 nm

Enzymatic synthesis of the peptides

Phac-Met-Asp(OMe)₂ H-Asp(OMe)₂ (350 mg, 2.18 mmol) and Phac-Met-OCam (400 mg, 1.23 mmol) were added into a 20-ml flask, and dissolved with ethyl acetate (10 ml) which contained 0.05 mol/L Tris HCl buffer (pH 9.0, 150 μ l). After the addition of chymotrypsin (15 mg), the mixture was shaken at room temperature until the peak of Phac-Met-OCam was not observed in HPLC (mobile phase system 玉 and 域). The reaction mixture was diluted by ethyl acetate (50 ml), and the organic layer was washed with 5% Na₂CO₃, 10% citric acid and saturated NaCl solution in succession, and dried over anhydrous sodium sulphate. After concentrated in vacuum, a white solid substance was obtained (320 mg, 63%). mp 128-129 $^{\circ}$ C, FAB-MS: 411 (M+H)⁺.

Phac-Met-Asp(OMe)-OH Phac-Met-Asp(OMe)₂

(1.6 g, 4 mmol) was suspended in 0.2 mol/L KH₂PO₄ buffer (pH 6.0, 50 ml) containing 2-mercaptoethanol (100 μ l). The pH of the mixture was adjusted to 6.0 with 1 mol/L HCl. Papain (50 mg) was subsequently added to the suspension, which was shaken at room temperature under constant pH control (pH 6.0) until the peak of Phac-Met Asp (OMe)₂ disappeared as observed by HPLC (Mobile Phase System 域). The pH of the mixture was then adjusted to 2 to 3 with 6 mol/L HCl, followed by extraction with ethyl acetate (2 \times 100 ml) and washing the extract with 10% citric acid and saturated NaCl solution in succession. The organic layer was dried over anhydrous sodium sulphate and concentrated in vacuum, and finally a white powder was obtained after lyophilization (1.46 g, 92%), mp 141-144 $^{\circ}$ C, FD-MS: 396 (M⁺).

Phac-Met-As (OMe)-Phe-NH₂ Phac-Met-Asp(OMe)-OH (800 mg, 2 mmol) and H-Phe-NH₂ HCl (600 mg, 3 mmol) were suspended in distilled water (20 ml). The pH of the mixture was adjusted to 7.0 with 4 mol/L NaOH. Thermolysin (10 mg) was added to the resultant cloudy solution, upon which white precipitate was immediately produced. The reaction mixture was kept in 40 $^{\circ}$ C of water bath overnight. After the mixture was cooled in ice bath, the precipitate was collected by filtration, washed carefully with cold 1% NaHCO₃ (3 \times 10 ml), water (2 \times 10 ml), and 5% citric acid (3 \times 10 ml), respectively. The precipitate was again washed with water until it turned neutral, and a white powder was resulted (895 mg, 82.6%). mp: 210-213 $^{\circ}$ C, FAB-MS: 543 (M+H)⁺.

RESULTS AND DISCUSSION

The synthesis was initiated at the amino terminal of the peptide, opposite to the way generally used in chemical peptide synthesis. The chemical synthesis is always inherits the danger of racemization during the activation of the carboxyl group of the peptide, a problem not seen in enzymatic methods. This is one of the major advantages of enzyme-catalyzed peptide synthesis.

Enzymatic synthesis of Phac-Met-Asp(OMe)₂

Usually, it is not easy to form the peptide bond of Met-Asp by means of coupling of Met and Asp derivatives catalysed by enzymes^[4]. In the initial approach,

the synthesis was attempted in aqueous buffer phase or in organic solvents-buffer by coupling of Phac-Met-OR (R=H, Me or Et) and H-Asp(OMe)₂ under the catalysis by 3 enzymes (chymotrypsin, papain or thermolysin), but it was not successful, and only the hydrolysis products of ester (Phac-Met-OR) were obtained.

Using such activated ester as the carbamoylmethyl (Cam) ester as the acyl donor to control peptide bond formation in the reaction kinetics, the dipeptide Phac-Met-Asp(OMe)₂ could be obtained through chymotrypsin-catalyzed coupling of Phac-Met-OCam and free H-Asp(OMe)₂ in a yield above 63%, and the reaction proceeded in organic solvent ethyl acetate containing 0.05 mol/ml Tris-HCl buffer (less than 1.5%, V/V, pH 9.0). Water is essential for the enzyme to retain its bioactivity in organic solvents^[10], and adequate water activity^[11] in the synthetic reaction system may facilitate the formation of the peptide bonds. The dipeptide Phac-Met-Asp(OMe)₂ could not be obtained in reasonable yields when the content of Tris-HCl buffer (0.05 mol/ml) exceeded 2% in the organic solvent ethyl acetate, and when the buffer content reached 10% (V/V) in the ethyl acetate, a sticky mixture would result. It should be noted that no product could be obtained when the reaction was carried out in waterless ethyl acetate or acetonitrile even in the presence of the same enzyme.

Fig.2 describes the kinetic time course of chymotrypsin-catalyzed Phac-Met-Asp(OMe)₂ synthesis, in which it is manifest that the hydrolysis product (Phac-Met-OH) of Phac-Met-OCam was produced during the reaction. Results of this study showed that Phac-Met-Asp(OMe)₂ was stable in ethyl acetate containing 0.05 mol/ml Tris-HCl buffer (1.5%, V/V, pH 9.0) in the presence of chymotrypsin within 1 or 2 days without amide or ester hydrolysis reaction.

Enzymatic synthesis of Phac-Met-Asp(OMe)-Phe-NH₂

The synthesis of the protected tripeptide, Phac-Met-Asp(OMe)-Phe-NH₂, was accomplished by coupling of Phac-Met-Asp(OMe)-OH and H-Phe-NH₂ in a yield of 82.7% with thermolysin in water. Phac-Met-Asp(OMe)-OH was obtained by papain-catalyzed saponification of Phac-Met-Asp(OMe)₂ in 0.2 mol/ml KH₂PO₄ pH 6.0 buffer, and methyl ester of aspartic acid was hydrolyzed while methyl ester

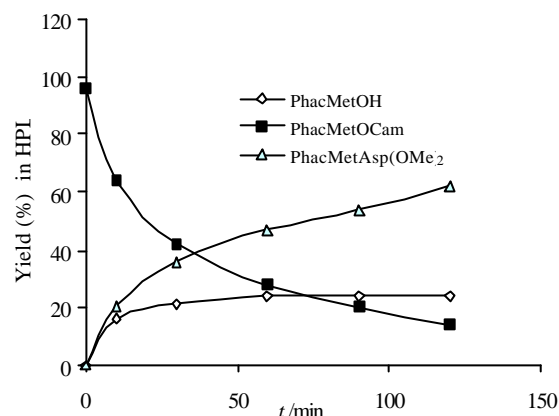


Fig.2 Time course of the enzymatic reaction between Phac-Met-OCam and H-Asp(OMe)₂

The reaction was carried out in ethyl acetate containing 0.05 mol/ml Tris-HCl buffer (1.5%, V/V, pH 9.0) with chymotrypsin. Acyl donor Phac-Met-OCam; Dipeptide Phac-Met-Asp(OMe)₂; Hydrolysis product Phac-Met-OH.

remained steady under the reaction condition. The resulted dipeptide, Phac-Met-Asp(OMe)-OH was important since it could easily bind H-Phe-NH₂ in high yield (>82.6%) in the presence of thermolysin, and produced almost no by-products. In addition, the tripeptide Phac-Met-Asp(OMe)-Phe-NH₂ was not obtained successfully with enzyme-catalyzed coupling of Phac-Met-Asp(OMe)₂ and H-Phe-NH₂ as shown in Tab.1.

Tab.1 The results of the synthesis of Phac-Met-Asp(OMe)-Phe-NH₂ with enzymes catalyzed coupling of Phac-Met-Asp(OMe)₂ and H-Phe-NH₂-HCl

Enzyme	Medium	Result
chymotrypsin	0.2 mol/L NaHCO ₃ buffer, pH 9.5	No product
Thermolysin	H ₂ O, pH 7.0	Yield < 1%
Papain	0.2 mol/L KH ₂ PO ₄ buffer, pH 8.5	Yield < 10%

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酶催化合成 CCK-4 三肽片段

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摘要 目的 探讨酶法合成胃泌激素 CCK-4C- 末端片段三肽衍生物 Phac-Met-Asp(OMe)-Phe-NH₂ 遥方法 以 Phac-Met-OCam 为起始原料 采用三种蛋白酶 羧基蛋白酶 木瓜蛋白酶 嗜热菌蛋白酶 chymotrypsin, papain, thermolysin 经过三步酶促反应得到目标三肽化合物 研究了酶催化条件下 Met-Asp 肽键的形成以及天门冬氨酸双酯中 羧基酯的酶催化条件下的选择性水解 对酶以及反应介质的选择等酶促合成条件进行了研究 结果 三步酶促反应均可得到较合适的收率 3%~92% 产物经 FAB-MS 和 D-MS 质谱确认 结论 以羧基蛋白酶为酶催化剂 在含有 1.5% 0.05 mol/L Tris-HCl 缓冲液的乙酸乙酯介质中 Phac-Met-OCam 和 H-Asp(OMe)₂ 缩合可得到收率大于 63% 的缩合产物 木瓜蛋白酶催化下可选择性水解 Phac-Met-Asp(OMe)₂ 中的 羧基酯而保留 苄基酯 嗜热菌蛋白酶能有效地催化 Phac-Met-Asp(OMe)-OH 与 H-Phe-NH₂ 生成肽键 遥

关键词 酶促合成 肽键 胃泌激素 CCK-4C- 三肽衍生物