Estimation and Comparison of *Anopheles maculipennis s.l.*(Diptera: Culicidae) Survival Rates with Light-trap and Indoor Resting Data

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Abstract

The main vector of malaria in Europe and Palearctic region is *Anopheles maculipennis* complex. In order to determine the survival rate of *An. maculipennis s.l.* this study was carried out in Garaboteh (Zanjan province, Iran) in July 2003. In this study anophelines were sampled for 31 consecutive days from five fixed animal shelters and five light traps. Out of 24481 collected *An. maculipennis s.l.*, 19703 (80.48%) were female. Relative density of female anopheline with 95% CI was 74.25-78.48 and 49.16-52.36 in light traps (LT) and pyrethrum space spray catch (PSC), respectively. A significant difference was in mean parous rate in LT (0.46) and PSC (0.50) samples (P< 0.01). Daily survival rate of anopheline mosquitoes in study area was 0.82-0.86. About 1.74% of female mosquitoes can transmit malaria after 10 d. In 4 d gonotrophic cycle, there was maximum correlation (>0.92) between parous and total females. Also there was significant correlation between nulliparous females and males in LT (r=0.96) and PSC (r=0.87) samples and with increasing anopheline abundance ratio of male/nulliparous significantly decreased (r=-0.35, r<0.05). Due to simplicity and feasibility of using graphical and parametric method, it can be placed in entomological surveys for monitoring parity, age structure and survival rate of vector anopheline in the world.

Keywords: Anopheles maculipennis s. l., Survival rate, Malaria, Iran

Introduction

Malaria is the most important vector borne disease which is transmitted by infected anophelinae mosquitoes, when they feed on human for their oviposition. Despite considerable progress in malaria control over the past decades, it is endemic in more than 100 countries of the world and kills 1.1-2.7 million people annually (1). Also more than 40% of the world populations live in areas with the risk of malaria transmission. The main vector of malaria in Europe and Palearctic region is Anopheles maculipennis complex (2). Out of nine known species of this complex, An. maculipennis s.l., the historical vector of malaria in Europe and the Middle East, was the first discovered sibling species among mosquitoes (3, 4). This complex has been identified as the major vector of malaria in the Caspian Sea littoral and Central Plateau of Iran (5-8), higher altitudes in the north of Iraq, mainly from Zahko, Amadia, Rowanduz and Penjwin (9). Recently it has been identified as secondary vector in the Biga Plains of Turkey (10).

Control programs against anopheline vectors, such as large-scale use of insecticide impregnated bednets and indoor residual spraying reduce mosquito density and survival rate. The entomological impact of such programs can therefore be evaluated by age structure (11) and life expectancy of anopheline vectors (12).

McDonald (13) suggested that mosquito vectors mainly die of predation or environmental factors rather than old age. Clemens and Patterson

(14) described two possible patterns of adult mosquito mortality. In the first pattern, mortality increases with age and there is a positive linear relationship between collection dates and mosquito age. Since anophelines are gonotrophically concordant after the first cycle, and the feeding cycle is equivalent to the oviposition cycle so parous rate can be used directly as an estimation of survival per feeding cycle (15-18).

In the second (Gillies' exponential model) the mortality rate does not vary with age. It can be estimated by mark-release-recapture experiments or laboratory multiple age-grading studies (19-23). The exponential model is one of the few methods applicable to gonotrophically disconcordant mosquito species. However, this model has several drawbacks, fundamental and potentially flawed assumption of the model is that the rate of female survivorship does not change with advancing age.

Pyrethrum space spraying and light-traps have been routinely used in studies of tropical and temperate mosquitoes (19, 24-28). Males of *An. maculipennis s.l.* like females are caught in light-trap. Indeed they rest commonly indoor and also are caught by pyrethrum space spray method. Ordinarily, male mosquitoes have low longevity (29, 30) and their activity is similar to nulliparous female specimens.

In this study the hypothesis that male anopheline frequency could estimate nulliparous female abundance, was tested in order to develop a more efficient use of the light trap and pyrethrum space spray for monitoring of age structure and parity of *An. maculipennis s. l.* and other malaria vectors in the world.

Materials and Methods

Study area The investigation was carried out in over period of one month (July, 2003) in district of Gharaboteh (47° 40′ 10″ E, 37° 7′ 15″ N), Province of Zanjan. This district lies about 85 km Southwest of Zanjan, center of province, and has about 5100 inhibitants. The district is about 1160 m above sea level and lies near the

Ghezel-Ozan River. Rice fields, swampy places arising from irrigation waters, stagnant water, pools, stream fed pools, streams and fish rearing pools are the major sources of mosquito breeding in this area. The averages of maximum and minimum temperatures in summer are 32- 15° C and 5.3- -2.5° C in winter, respectively. The average yearly rainfall is about 300 mm. The district had not been under vector control program during the study period.

Light trap collections Five miniature CDC light-traps (Hausherr's Machine Works, Toms River, New Jersey and John W. Hock Company, Gainesville, Florida) were hung adjacent to mosquito nets. Light-traps operated by rechargeable batteries, were fitted with 12 V halogenated bulbs and 0.7 cm mesh grids to exclude larger insects. Traps were switched on at 21: 00 h local time, and were instructed to be switched them off at 06:00AM.

Pyrethrum space spray catches Five fixed animal shelters were used in this program. Every morning indoor rested mosquitoes in five animal shelters were collected by the standard method (25), using 0.2% pyrethrum spray.

Mosquito processing All collected mosquitoes were transferred into the plastic jars. The house and trap number and date of collection were recorded on the jar label and then jars were transferred into the cool box with ice packs. In the laboratory collections were identified under dissecting microscope (at x40) against standard taxonomic keys frequency of female and male of An. maculipennis s.l. in each sample were recorded and females were classified according to the blood digestion stages (abdominal conditions). Empty and freshly feed An. maculipennis s.l. were dissected for parity and classified as nulliparous and parous on the basis of the tracheolar skeins of the ovaries (31, 32).

Statistical analysis Survival rate and blood-meal frequency were estimated by using linear regression, to relate the numbers of mosquitoes biting on one day, to parous individuals collected one oviposition cycle later (17, 18, 33).

To test the efficiency of mosquito male frequency in estimating nulliparous females, graphical and parametric method were utilized (34) to examine bias and error in methods.

Results

A total of 24481 An. maculipennis s.l. were collected during July 2003. There was not significant difference in sex ratio of collected mosquitoes ($X^2 = 0.65$, P = 0.42) in both methods. Out of collected specimens, 19703 (80.48 %) were female. There were not significant differences in abundance of mosquitoes in five light traps (LT) and five indoor pyrethrum space spray catch (PSC) (Table 1). Relative density of An. maculipennis s.l. female and male within indoor resting with 95% confidence limits was 74.25-78.45 and 17.68-18.96, respectively. In light trap samples, the density was 49.16-52.36 and 11.91-13.11, respectively. In both studying methods, density of male anopheline was correlated with females (Fig. 1) and Pearson correlation coefficient between them was 0.7149.

Comparison of blood feeding stages of An. maculipennis s.l. caught by LT and PSC (Table 2) indicated that the proportion of blood fed captured mosquitoes by LT was lower (10%) than PSC (72.5%), ($X^2 = 14468.77$, df = 2, P = 0). Out of PSC captured An. maculipennis s.l., 8476 females were dissected for parity. In dissected females, 4295 (51%) were parous and in LT samples out of 7869 dissected females, parous rate was 0.46. A significant difference (z = 6.67, P < 0.01) in the mean parous rates was detected between LT and PSC captured mosquitoes (Table 3).

In 4 days gonotrophic cycle, there was maximum correlation (r= 0.92, for LT and r=0.99, for PSC) between parous and total female mosquitoes. Mean parous and survival rate during gonotrophic period in LT and PSC sampling methods were 0.46 and 0.50, respectively. This rate was unstable in first 10 days but afterwards was fixed (Fig. 2).

Pearson correlation coefficients for the relationship between abundance of log transformed nulliparous females and males in LT and PSC collections were calculated. There was significant correlation (Fig. 3) between nulliparous females and males in LT (r= 0.96) and PSC (r= 0.87) samples of An. maculipennis s.l. The potential bias in estimating nulliparous anopheline mosquito abundance by LT and PSC was examined graphically. There was significant decrease tendency (Fig. 4) for the ratio of male/nulliparous with increasing mosquito abundance in PSC (r= -0.35, P< 0.05) and LT (r= -0.58, P< 0.001) samples.

Table 1: Analysis of deviance for the mean number of *An. maculipennis s. l.* collected in light traps and pyrethrum space spray method from Garaboteh (Zanjan, Iran), July 2003

	Pyrethrum space spray		Light trap	
-	Female	Male	Female	Male
Total number	11835	2839	7868	1939
Mean (95%CL)	74.25- 78.45	17.68- 18.96	49.16- 52.36	11.91- 13.11
F (4,151) ns*	0.14	0.05	0.16	0.06

ns* = not significant, P < 0.001

Table 2: Blood feeding condition of *An. maculipennis s. l.* caught by light trap and pyrethrum space spray from Garaboteh (Zanjan, Iran), July 2003

Abdominal condition	Light trap (%)	Pyrethrum space spray (%)	Total
Empty	6685 (85)	189 (1.5)	874
Freshly-fed	790 (10)	8586 (72.5)	9376
Gravid	393 (5)	3060 (26)	3453

Table 3: Parameter estimates for An. maculipennis s. l. collected from Garaboteh (Zanjan, Iran) in July 2003

Time delay Survival rate		Correlation index
yrethrum space spr	ay captured mosquitoes	
0	0.506	0.38
1	0.508	0.11
2	0.509	0.19
3	0.509	0.55
4	0.508	0.92
5	0.509	0.69
6	0.5011	0.44
7	0.5014	0.12
8	0.5015	0.33
9	0.5012	0.56
10	0.508	0.65
ght trap captured m	=	
0	0.461	0.26
1	0.463	0.05
2	0.463	0.35
3	0.462	0.68
4	0.460	0.99
5	0.459	0.68
6	0.460	0.36
7	0.461	0.32
8	0.464	0.35
9	0.462	0.41
10	0.457	0.39
		−□ −PSC Female
<u></u>		LT Female
		LT Male
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2 3 4 5 6 7 8	9 10 11 12 13 14 15 16 17 18	19 20 21 22 23 24 25 26 27 28 29 3
	Date (July 2003)	

Fig. 1: Abundance of *An. maculipennis s.l.* collected by light traps (LT) and pyrethrum space spray (PSC) in Garaboteh (Zanjan Iran) in July 2003.

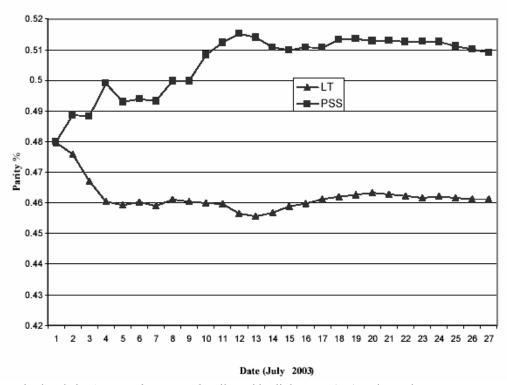


Fig. 2: Parity levels in *An. maculipennis s. l.* collected by light traps (LT) and pyrethrum space spray (PSS) in Garaboteh (Zanjan, Iran) in July 2003.

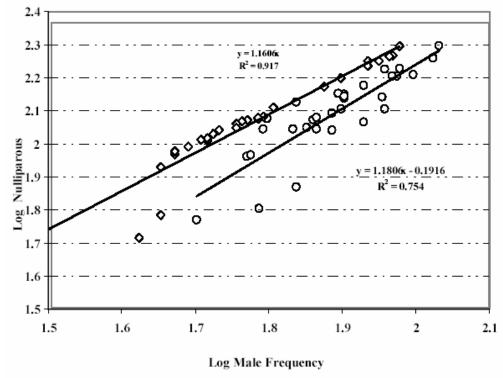


Fig. 3: Scatter distribution for the relationship between log transformed males and nulliparous females of *An. maculipennis s. l.*, collected with light traps (\Diamond) and pyrethrum space spray (O) in Garaboteh (Zanjan, Iran) in July 2003.

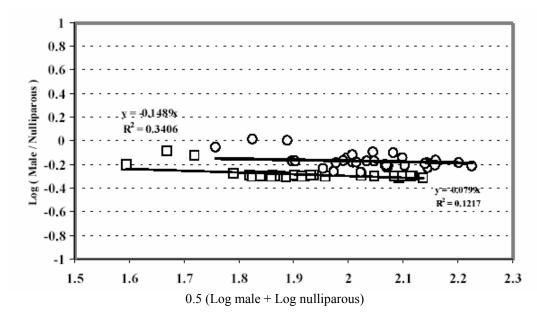


Fig. 4: Scatter distribution for the relationship as log ratio between male and nulliparous abundance and the William's mean (on logarithmic scale) of male and nulliparous females of *An. maculipennis s. l.*, catch by light traps (□) and pyrethrum space spray (O) in Garaboteh (Zanjan, Iran) in July 2003

Discussion

Although malaria was officially declared eradicated from central plateau of Iran in 1975, its former vectors, mainly members of the *An. maculipennis* complex, are still distributed throughout the plateau and Caspian Sea littoral. The present situation of anophelism without malaria indicates that current socioeconomic and environmental conditions maintain the basic case reproduction number (Ro) below 1. Recently, it has been speculated that predicted climate changes may increase anopheline abundance and biting rates (as well as reduce the *Plasmodium* parasite extrinsic incubation period), allowing the reemergence of malaria transmission.

The findings showed that *An. maculipennis s.l.* had high density in the study area. These observations are consistent with the results of Manoucehri et al. (5) in central Plateau and Caspian Sea littoral. High reletive density has been reported for *An. gambiae* as 44-65 (33) and for *An. punctipennis* as 16-72 (35).

According to findings, outdoor light traps underestimated indoor resting anopheline. This

observation is consistent with works of Charlwood et al. (36) who found that the relative density of An. farauti collected in light traps was 25-80 from Papua New Guinea. High density of An. maculipennis s.l. in Garaboteh (Zanian) is expected, because in this area vector control program has not been performed over the past 30 yr and study area has potential breeding sites. Because the use of light traps in connection with mosquito nets decreased the number of blood fed mosquito caught, so 90% of females caught from light traps were unfed. Wekesa et al. (37) showed that only 28% of Culex tarsalis collected from different habitats were blood fed. In their studies this percentage for An. freeborni was 23%. Pyrethrum space spray unlike light traps caught >72% of freshly fed females. Sadandene et al. (28) in study of Korapat, Orisa, India: found that 82% of anopheline mosquitoes caught from indoor light traps were freshly fed. In this study an oviposition cycle of 4 d had high correlation coefficiency in parous and total An. maculipennis s.l. A significant oviposition cycle of 4 d was obtained for An. arabiensis at Mwea with potential breeding sites, however

that was on inland site with relatively low mean temperature (33). Overestimate on gonotrophic cycle of An. freeborni in Sacramento Valley, An. punctipennis in Maryland and An. quadrimaculatus in Florida were reported to be 4-6, 4-5, and five d (38-40). Unlike this study, lower estimation of gonotrophic cycle was reported from tropical areas. The length of oviposition cycle for An. gambiae and An. merus in rainy seasons was 2 d. In both cases the breeding places were close to sampling sites. An oviposition cycle of 3 d was obtained for An. gambiae at Misha, Kenya during dry seasons while there were not potential breeding sites (33). The additional time required for oviposition in Garaboteh (Zanjan) may reflect the influence of environmental conditions especially mean temperature.

A significant difference was seen in mean survival rate per oviposition cycle in the population of An. maculipennis s.l. caught from light traps (0.46) and pyrethrum spray catch (0.50). Lower parity was reported by Githeko et al. (41) in light trap samples. A significant difference in the mean parous rate later confirmed by McHug (39, 42) between unfed (0.36.4) and blood-fed (42.5) of *Culex tarsalis*. This discrepancy may be explained by difference in the genetic composition of two vector population with respect to the member of An. maculipennis complex, special behavior of nulliparous or parous mosquitoes and lowered parity of unfed females. However, low survival rate was reported for An. labranchiae, an important malaria vector of Italy (43, 44), some population of An. punctipennis from Pupua New Guinea (45). Mean daily survival rate obtained in this study (0.82-0.84) was higher than some populations of An. maculipennis from Iran (5) and An. labranchiae from Italy (43, 44). In case of other Anopheles species, the probability of daily survival was 0.89 for An. pharensis and 0.80 for An. multicolor in Egypt (46), 0.80-0.83 for An. pulcherrimus in Iran (47), 0.88 for An. psudopunctipennis in southern Mexico (48), 0.80- 0.88 for An. gambiae s.l. in Sudan (21) and 0.45- 0.68 for An.

vestitipennis in southern Mexico(49). The importance of daily survival of vectors for efficient transmission of infection is obvious. Mean temperature at Garaboteh (Zanjan) in July was 25°C and duration of sporogonic cycle of *Plasmodium vivax* is 10 d at 25° C. Therefor the probability of *An. maculipennis s.l.* survival through 10 d was 0.174, which means that 1.74% of female mosquitoes respectively may live long enough to transmit malaria.

The findings showed significant correlation between males and nulliparous data. There was significant tendency for the ratio of male/ nulliparous, so decrease arises with increasing mosquito abundance. Also there was difference in the log ratio on male/nulliparous between light trap and pyrethrum space spray catch data. The mean log ratio for LT and PSC data were -0.3 and -0.2, whose antilogies were 2 and 1.5, respectively. This means that on average the catch 1 male from LT or PSC method were 2 and 1.5 times that of nulliparous in that sample. Graphical and parametric method has been successfully used for detecting relationship between anopheline abundance in light traps and indoor resting or outdoor human landing collections (50-54). Due to its simplicity and feasibility on anopheline parity detection, this method can be placed in public health evaluation plans and declare the age structure and survival rate of vector anopheline in different geographical areas.

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References

- 1. World Health Organization (2000). WHO Expert Committee on Malaria. Twentieth Report. WHO Technical Report Series 892.
- 2. White GB (1978). Systematic reappraisal of the *Anopheles maculipennis* complex. *Mosquito Systematics*, 10: 13-44.

- 3. Sedaghat MM, Linton YM, Oshaghi MA et al. (2003). The *Anopheles maculipennis* complex (Diptera:Culicidae) in Iran: molecular characterization and recognition of new species. *Bull Entomol Res*, 93: 527-35.
- 4. Harbach RE (2004). The classification of genus *Anopheles* (Diptera: Culicidae) a working hypothesis of phylogenetic relationships. *Bull Entomol Res*, 94: 537-53.
- 5. Nicolescu G, Linton YM, Viadimireescu A et al (2004). Mosquitoes of the *Anopheles maculipennis* group (Diptera: Culicidae) in Romania with the discovery and formal recognition of new species based on molecular and morphological evidence. *Bull Entomol Res*, 94: 525-35.
- 6. Manouchehri AV, Zaini A, Motaghi M (1976). Resistance of *Anopheles maculipennis* to insecticides in northern Iran. *Mosquito News*, 36: 51-5.
- 7. Manouchehri AV, Zaim M, Emadi AM (1992). A review of malaria in Iran, 1975-90. *Journal of the American Mosquito Control Association*, 4: 381-85.
- 8. Zaim M (1987). Malaria control in Iran-present and future. *J Am Mosq Control Assoc*, 3: 392-96.
- 9. Zahar AR (1990). Vector bionomics in the epidemiology and control of malaria: Part II; the WHO European region & the WHO Eastern Mediterranean Region. *WHO/VBC/90.3 -MAL/90.3*.
- 10. Alten B, Caglar SS and Özel O (2000). Malaria and its vectors in Turkey. *European Mosquito Bulletin*, 7: 27-33.
- 11. World Health Organization (2002). Malaria Entomology and Vector Control, Learner's Guide, Social Mobilization and Training Control, Prevention and Eradication Department, Communicable Diseases Cluster, World Health Organization. Geneva, p. 114.
- 12. Garret-Jones C, Grab B (1964). The assessment of insecticidal impact on the malaria mosquito's vectorial capacity, from

- the data on the proportion of parous females. *Bulletin of World Health Organization*. 31: 71-86.
- 13. MacDonald G (1957). *The Epidemiology* and Control of Malaria. Oxford University Press, London.
- 14. Clements AN, Paterson GD (1981). The analysis of mortality and survival rates in wild population of mosquitoes. *J Applied Ecology*, 18: 373-99.
- 15. Birley MH, Rajagopalan PK (1981). Estimation of the survival and biting rates of *Culex quinqufasciatus* (Diptera: Culicidae). *J Med Entomology*, 18: 181-86.
- 16. Birley MH, Boorman JP (1982). Estimating the survival and biting of haematophagus insects with particular reference of *Culicoides obosletus* group (Diptera: Ceratopogonidae) in Southern England. *J Animal Ecology*, 51:133-48.
- 17. Holmes PR, Birely MH (1987). An improved method for survival rate analysis from time series of haematophagus dipteran populations. *J Animal Ecology*. 56: 427-40.
- 18. Lord CC, Baylis M (1999). Estimation of survival rates in haematophagus insects. *Med Vet Entomol*, 13: 225-33.
- 19. Service MW (1993). *Mosquito Ecology*. *Field Sampling Methods*. 2nd ed. London, Elsevier Applied Science Publishers Ltd. London., P. 988.
- 20. Day JF, Edman JD, Scott TW (1994). Reproductive rates and survivorship of *Aedes aegypti* (Diptera: Culicidae) maintained on blood, with field observations from Thailand. *J Med Entomol*,31:611-7.
- 21. Constantini C, Li SG, Della-Torre A et al. (1996). Density, survival and dispersal of *Anopheles gambiae* complex mosquitoes in a West African Savana village. *Med Vet Entomol* 10: 203-19.
- 22. Muir LE, Kay BH (1998). *Aedes aegypti* survival and dispersal estimated by mark-release-recapture in northern Australia. *Am J Trop Med Hyg*, 58: 277-87.

- 23. Morrison AC, Costero A, Edman JD, Clark GG and Scott TW (1999). Increased fecundity of *Aedes aegypti* (Diptera: Culicidae) fed human blood prior to release in a mark recapture study in Puerto Rico. *J Am Mosq Control Assoc.* 15: 98-102.
- 24. Zaim M, Ershadi MRY, Manouchehri AV, Hamdi MR (1986). The use of CDC light traps and other procedures for sampling malaria vectors in southern Iran. *J Am Mosq Control Assoc*, 2: 511-15.
- 25. World Health Organization (1990). The epidemiology and control of malaria. The WHO, Eastern Mediterranean Region. Mimeographed document. WHO/VBC/90.2-MAL/90.3.
- 26. Takken W, Charlwood JD, Billingsley PF, Gort G (1998). Dispersal and survival of *Anopheles funestus* and *A. gambiae s.l.* (Diptera: Culicidae) during the rainy season in southeast Tanzania. *Bulletin of Entomological Research*. 88: 561-66.
- 27. Hii JL, Smith T, Mai A et al. (2000). Comparison between anopheline mosquitoes (Diptera: Culicidae) caught using different methods in a malaria endemic area of Papua New Guinea. *Bull Entomol Res*, 90: 211-19.
- 28. Sadanandane C, Jambulingam P, Subramanian S (2004). Role of modified CDC miniature light-traps as an alternative method for sampling adult anophelines (Diptera: Culicidae) in the National Mosquito Surveillance Program in India. *Bull Entomol Res*, 94: 55-63.
- 29. Mahmood F, Reisen WK (1982). *Anopheles stephensi* (Diptera: Culicidae): changes in male mating competence and reproductive system morphology associated with aging and mating. *J Med Entomology*, 19: 573-88.
- 30. Grieco JP, Nicole LA, Ireneo B, Russell K, Richard A, Donald R, Rejmankova E (2003). Comparison of life table attributes from newly established colonies of *Anopheles albimanus* and *Anopheles*

- vestitipennis in northern Belize. J Vector Ecology 28: 200-207.
- 31. Detinova TS (1962). Age-grouping Methods in Diptera of Medical Importance. World Health Organization. p. 216.
- 32. World Health Organization (1975). *Manual on Practical Entomology in Malaria*. WHO Offset Publ. 13. p. 160.
- 33. Mutero CM, Birely MH (1987). Estimation of the survival rate and oviposition cycle of field population of malaria vectors in Kenya. *J Applied Ecology*, 24: 853-63.
- 34. Bland J, Altman DG (1995). Comparing methods of measurement: why plotting difference against standard method is misleading. *Lancet*; 346: 1085-87.
- 35. Jensen T, Dritz DA, Fritz GN Washino RK, Reeves W (1993). Lake vera revisited: parity and survival rates of *Anopheles punctipennis* at the site of malaria outbreak in the Sierra Nevada foothills of California. *Am J Trop Med & Hyg*, 59: 591-94.
- 36. Charlwood JD, Birley MH, Dagoro H, Paru R, Holmes PR (1985). Assessing survival rates of *Anopheles farauti* (Diptera: Culicidae) from Popua New Guinea. *J Animal Ecology*, 54: 1003-16.
- 37. Wekesa JW, Yual B, Washino RK, de Vasquez AM (1997). Blood feeding patterns of *Anopheles freeborni* and *Culex tarsalis* (Diptra: Culicidae) effect of habitat and host abundance. *Bull Entomol Res*, 39: 633-41.
- 38. Washino RK and Bailey SF (1970). Over wintering of *Anopheles punctipennis* (Diptera: Culicidae) in California. *J Med Entomol*, 7: 95-98.
- 39. McHugh CP (1989). Ecology of a semiisolated population of adult *Anopheles freeborni*: abundance, trophic status, parity, survivorship, gonotrophic cycle length and host selection. *Am J Trop Med Hyg*, 41: 169-76.
- 40. Jensen T, Kaiser PE, Barnard DR (1993). Short-term changes in the abundance

- and parity rate of *Anopheles quadri-maculatus* species C (Diptera: Culicidae) in a central Florida swamp. *J Med Ento-mol*, 30: 1038-42.
- 41. Githeko AK, Service MW, Mbogo CM, Atieli FA, Juma FO (1994). Sampling *Anopheles arabiensis, An. gambiae s.l.* and *An. funestus* (Diptera: Culicidae) with CDC light traps near a rice irrigation area and a sugarcane belt in western Kenya. *Bulletin of Entomological Research*, 84: 319-24.
- 42. McHugh CP (1990). Survivorship and gonotrophic cycle length of *Culex tarsalis* (Diptera: Culicidae) near Sheridan, Placer County, California. *J Med Entomol*, 27: 1027-30.
- 43. Romi R (1999). *Anopheles labranchiae*, an important malaria vector in Italy, and other potential malaria vectors in southern Europe. *European Mosquito Bulletin*, 4: 8-10.
- 44. Romi R, Pierdominici G, Severini C, Tamburro A, Cocchi M, Menichetti D, Pili E, Marchi A (1997). Status of malaria vectors in Italy. *J of Medical Entomology*, 34: 263-71.
- 45. Bockarie MJ, Aleander N, Bockarie F, Ibam E, Banish G, Alpers M (1996). The late biting habit of parous *Anopheles* mosquitoes and pre-bedtime exposure of human to infective mosquitoes. *Trans Roy Soc Trop Med Hyg*, 90: 23-4.
- 46. Kenawy MA (1991). Development and survival of *Anopheles pharoensis* and *An. multicolor* from Faiyum, Egypt. *J Am Mosq Control Assoc*, 7: 551-55.
- 47. Zaim M, Zahirnia AH, Manouchehri AV (1993). Survival rate of *Anopheles culicifacies*. and *Anopheles pulcherrimus* in sprayed and unsprayed in Ghassreghand District, Buluchistan, Iran, 1991. *J Am Mosq Control Assoc* 9: 421-25.
- 48. Fernandes-Salas I, Rodrigues MH and Roberts DR (1994) Gonotrophic cycle and survivorship of *Anopheles pseudopunc*-

- *tipennis* (Diptera: Culicidae) in the Tapachura foothills of Southern Mexico. *J Med Entomol*, 31: 340-47.
- 49. Arredondo-Jimenez JI, Rodriguez MH, Washino RK (1998). Gonotrophic cycle and survivorship of *Anopheles vestitipennis* (Diptera: Culicidae) in two different ecological areas of southern Mexico, *J Med Entomol* 35: 937-42.
- 50. Lines JD, Curtis CF, Wilks TJ, Njunwa, KJ (1991). Monitoring human biting mosquitoes (Diptera: Culicidae) in Tanzania with light traps hung beside mosquito nets. *Bull Entomol Research*, 81: 77-84.
- 51. Mbogo CM, Glass GE, Forster D, Kabiru EW, Githure JI, Ouma JH, Beier J (1993). Evaluation of light traps for sampling anopheline mosquitoes in Kilifi, Kenya. *Journal of the American Mosquito Control Association*, 9: 260-63.
- 52. Davis JR, Hall T, Chee EM, Majala A, Minjas J, Shiff CJ (1995). Comparison of sampling anopheline mosquitoes by light trap and human bait collections indoors at Bagamoyo, Tanzania. *Medical and Veterinary Entomology*, 9: 249–55.
- 53. Costantini C, Sagnon NF, Sanogo E, Merzagora L, Coluzzi M (1998). Relationship to human biting collections and influence of light and bednet in CDC light-trap catches of West African malaria vectors. *Bulletin of Entomological Research*, 88: 503–11.
- 54. Magbity EB, Magbity EB, Lines JD, Marbiah MT, David K, Peterson E (2002). How reliable are light traps in estimating biting rates of adult *Anopheles gambiae s.l.* (Diptera: Culicidae) in the presence of treated bed nets? *Bull Entomol Research*, 92: 71-6.