Seminal Plasma Lactoferrin Concentrations in Normal and Abnormal Semen Samples

WILLIAM M. BUCKETT,* MURRAY J. M. LUCKAS,* MEHMET R. GAZVANI,* IAN A. AIRD,† AND D. IWAN LEWIS-JONES†

From the *Reproductive Medicine Unit, Liverpool Women's Hospital, and the †Department of Obstetrics and Gynaecology, University of Liverpool, Liverpool, UK.

ABSTRACT: Although the iron-chelating protein lactoferrin is secreted by the seminal vesicles, the precise role of lactoferrin in semen is unclear. This study aimed to determine whether there is any association between seminal lactoferrin concentrations and normal and abnormal semen samples with and without leucocytospermia. Lactoferrin concentrations were measured by radial immunodiffusion of semen samples from 368 men attending a regional andrology referral center. Routine seminal analysis, including the presence of leucocytospermia, was also performed. Results showed increased seminal lactoferrin in samples showing oligospermia (13.3 mg/100 ml) and oligoasthenospermia (13.4 mg/100 ml) compared to normospermic samples (11.2 mg/100 ml). There were no significant differences in seminal lactoferrin between normospermic samples and azoospermic samples or asthenospermic samples with normal sperm density. Although there was a trend toward increased lactoferrin concentration with leucocytospermia, this was not significant. Possible causes for raised lactoferrin in association with oligospermia are discussed.

Key words: Leucocytospermia, oligospermia, asthenospermia, infertility.

J Androl 1997;18:302-304

Lactoferrin is a medium-molecular-weight protein (84 kDa) with iron-binding properties. It was first isolated in bovine milk and subsequently in human breast milk (Schäfer, 1951). The protein is present in many human secretions and is also found in the epithelial lining of the stomach, colon, lung, and genital tract, and in neutrophils (Groves, 1971). It was first identified in human seminal plasma by Masson et al (1966) and is secreted by the seminal vesicles (Hekman and Rümke, 1968; Tauber et al, 1975).

By virtue of its iron-chelating effect, lactoferrin is bacteriostatic (Masson and Heremans, 1966) and may therefore have a role in the defense against microorganisms (Masson, 1970). In breast milk, lactoferrin has been shown to act as an antimicrobial protein suppressing coliform bacteria in the neonate (Bullen et al, 1972). However, its role in semen remains unclear.

Lactoferrin levels have been reported to be raised in association with asthenospermia and oligospermia when compared to normospermia, although the numbers analyzed were small (Auterio et al, 1991). Wolff et al (1990) showed a significant relationship between leucocytospermia and a reduction in total numbers of sperm and sperm motility. Leucocytospermia is suggestive of genital-tract

302

infection (WHO, 1992), and lactoferrin concentrations may alter in such circumstances.

The purpose of this study was to determine any association between seminal plasma lactoferrin concentration and normal and abnormal semen samples with and without leucocytospermia.

Materials and Methods

Semen samples were obtained from 368 men attending a regional andrology infertility clinic. All semen samples were obtained by masturbation into a sterile nonspermicidal container after 4 days of sexual abstinence. The samples were allowed to liquefy for 30 minutes at room temperature. Routine seminal analysis, according to standard WHO criteria (1992), was then performed. In cases of complete azoospermia, serum folliclestimulating hormone (FSH) was also determined. Leucocytospermia was defined as $>1 \times 10^6$ white cells/ml by light microscopy.

Seminal lactoferrin concentrations were then determined on the supernatant after centrifugation at $1,000 \times g$ for 10 minutes. Single-radial-immunodiffusion assay was used (Mancini et al, 1965). Goat antisera to lactoferrin (Nordic Immunology Laboratories, Tilberich, Netherlands) was added to 1% agarose (Hyland Pharmaceuticals, Feltham, UK) and made into 1.5 mm plates that were then subdivided in two. Standardized human colostrum, lactoferrin concentration 10 mg/100 ml, was used as a standard for lactoferrin. Seminal plasma from each sample and standard colostrum was then added to three different plates. The diffusion through the agarose was measured (usually 20–30 mm)

Correspondence to: William M. Buckett, Reproductive Medicine Unit, Liverpool Women's Hospital, Crown St., Liverpool L8 7SS, UK.

Received for publication November 1, 1996; accepted for publication January 24, 1997.

Buckett et al · Seminal Lactoferrin in Male Infertility

Table 1. Number of samples displaying the different semen parameters and leucocytospermia

	Number of samples	Samples with leucocytospermia
Normospermia (>20 million/ml, >40% motility)	73	15 (20.5%)
Asthenospermia (>20 million/ml, <40% motility)	107	19 (17.4%)
Oligoasthenospermia (<20 million/ml, <40% motility)	115	17 (14.8%)
Oligospermia (<20 million/ml, >40% motility)	53	11 (20.8%)
Azoospermia (serum FSH > 9 IU/L)	10	1 (10.0%)
Azoospermia (serum FSH < 9 IU/L)	10	1 (10.0%)
Totals	368	64 (17.4%)

FSH, follicle-stimulating hormone.

after 48 hours' incubation, then the area was calculated and compared with the control in all plates, and the lactoferrin concentration calculated. The standard error of the lactoferrin concentration was earlier evaluated as 1.08%. Samples from all groups were run concurrently, and the technician performing the lactoferrin assay was blind to the results of the seminal analysis.

Data were assessed for normality using the Shapiro-Wilk test and described using the median and interquartile ranges. The Mann-Whitney U-test was used to compare groups.

Results

Of the 368 samples, 73 had normal semen parameters, 107 had asthenospermia with normal sperm density, 53 had oligospermia with normal sperm motility, 115 had oligoasthenospermia, and 20 had azoospermia (Table 1). A diagnosis of testicular germ cell hypofunction was made on the basis of raised serum FSH levels >9 IU/L in 10 of the azoospermia group, and the remaining 10 were presumed to have obstructive azoospermia.

Leucocytospermia was present in 64 samples (17.4%). The number of samples with leucocytospermia was 15 (20.5%) in the normospermic group, 19 (17.8%) in the asthenospermic group, 11 (20.8%) in the oligospermic group, 17 (14.9%) in the oligoasthenospermia group, and two (10%) in the azoospermic group. Leucocytospermia was not significantly raised in any of the groups.

There was no significant difference in median seminal lactoferrin concentrations between the normal group (11.2 mg/100 ml), the asthenospermia with normal sperm den-

Table 2. Median seminal lactoferrin (and interquartile ranges) in the different sperm groups

	Seminal lactoferrin (mg/100 ml)	
Normospermia $(n = 73)$	11.2 (6.3–15.7)	
Asthenospermia $(n = 107)$	12.6 (7.3–18.6)	
Oligoasthenospermia $(n = 115)$	13.3* (8. 9– 21.7)	
Oligospermia $(n = 53)$	13.4* (9.2–21.7)	
Azoospermia $(n = 20)$	11.1 (8.8–20.4)	

* Normal vs. oligoasthenospermia, P = 0.016; normal vs. oligospermia, P = 0.025.

sity group (12.6 mg/100 ml), and both azoospermia groups (11.2 mg/100 ml and 10.2 mg/100 ml). However, the oligospermia group (13.4 mg/100 ml) and the oligoasthenospermia group (13.3 mg/100 ml) both showed a significantly increased median seminal lactoferrin concentration (Table 2).

The median lactoferrin concentration in the leucocytospermia group (n = 64) was raised at 12.4 mg/100 ml compared to 11.4 mg/100 ml in the nonleucocytospermia group (n = 304), but this was not statistically significant.

Discussion

This large study analyzing 368 samples shows that seminal lactoferrin concentrations are raised in samples showing oligospermia and oligoasthenospermia. There was no significant difference in lactoferrin concentrations between complete azoospermia, whether presumed obstructive or associated with raised serum FSH levels, and normospermia samples. Auterio et al (1991) suggested that lactoferrin is raised in asthenospermia as well as oligospermia. Numbers in their study were small, and it is unclear whether the asthenospermic group had normal sperm density. Our study showed there was no significant difference between asthenospermia with normal sperm density and normospermic samples.

The reason for raised lactoferrin in individual semen samples remains unclear. Leucocytes are present in most ejaculates (Wolff and Anderson, 1988; Barratt et al, 1990); however, excessive presence of these cells (leucocytospermia) is traditionally associated with clinical and subclinical genital-tract infection (WHO, 1992). In this study, although median lactoferrin concentration was higher, there was no statistically significant difference in the between-samples exhibiting leucocytospermia and those that did not. This may be because the method for identifying leucocytes from other seminal cells and the various white-cell subsets is unreliable (Barratt et al, 1988). Alternatively, seminal leucocytes or increased lactoferrin concentrations may present in semen later than the period of acute infection. In this study, there was no history or clinical evidence of either vesiculitis or prostatitis.

Although lactoferrin concentrations were raised in oligospermia, there was no increase in lactoferrin concentration in cases of azoospermia. Lactoferrin is not present in semen from men with congenital absence of the seminal vesicles (Hekman and Rümke, 1968). A decrease in the seminal vesicle secretion caused by high-dose testosterone injection leads to lower lactoferrin concentrations in seminal plasma (Carpino et al, 1994). Therefore, seminal lactoferrin may reflect seminal vesicle function and the lack of a rise in cases of azoospermia may reflect concomitant abnormalities in the seminal vesicles (Lizana et al, 1987; Aumuller and Riva, 1992). Seminal fructose, which is also secreted from the seminal vesicles, has been used to assess seminal vesicle function (Gonzalez et al, 1989); however, only where there are significant abnormalities of the seminal vesicles is there a reduction of fructose concentrations.

Animal studies on the mouse epididymis show lactoferrin mRNA is stimulated by 17β -estradiol (Yu and Chen, 1993), and in the human female reproductive tract, lactoferrin concentration in vaginal mucus peaks just after the menses (Cohen et al, 1987), which also implies lactoferrin secretion is stimulated by estradiol. In men, serum and testicular estradiol have been shown raised in association with oligospermia (Krause, 1988; Levalle et al, 1994). Therefore, increased circulation of seminal estradiol could cause increased seminal lactoferrin secretion. Whether the rise in serum and testicular estradiol is directly involved in causing the oligospermia is at present uncertain.

Further studies need to measure serum and seminal estradiol as well as seminal lactoferrin in men with oligospermia to determine whether raised estradiol is the causative factor of raised lactoferrin. Also, better identification of white cells, in particular activated neutrophils and other leucocyte subgroups, is needed in order to determine whether lactoferrin is secreted as part of an acute or chronic infective condition.

Acknowledgments

We thank D. B. McClelland, J. M. McGrath, and R. R. Samson for kindly providing the standardized human colostrum.

References

Aumuller G, Riva A. Morphology and functions of the human seminal vesicle. Androlagia 1992;24:183-196.

Auterio, M, Sansone G, Abrescia P. Relative ratios of lactoferrin, albumin,

and acid phosphatase seminal levels as sperm quality markers in fertile and infertile men. J Androl 1991;12:191-200.

- Barratt CLR, Bolton AE, Cooke ID. Functional significance of white blood cells in the male and female reproductive tract. *Hum Reprod* 1990;5:639-644.
- Barratt CLR, Li TC, Monteiro EF. Diagnosis and detection of male accessory gland infection. In: Barratt CLR, Cooke ID, eds. Advances in Clinical Andrology. Lancaster: MTP Press; 1988:91–97.
- Bullen JJ, Rogers HJ, Leigh L. Iron-binding proteins in milk and resistance to Escherichia coli infection in infants. Br Med J 1972;264:69– 75.
- Carpino A, Sisci D, Aquila S, Beraldi E, Sessa MT, Siciliano L, De Luca G, Ando S. Effects of short-term high dose testosterone propionate administration on medium molecular-weight proteins of human seminal plasma. Androlagia 1994;26:241-245.
- Cohen MS, Britigan BE, French M, Bean K. Preliminary observations on lactoferrin secretion in human vaginal mucus: variation during the menstrual cycle, evidence of hormonal regulation, and implications for infection with Neisseria gonorrhoeae. Am J Obstet Gynecol 1987;157:1122-1125.
- Gonzalez GF, Garcia-Hjarles MA, Gutierrez R, Guerra-Garcia R. The secretory activity of the seminal vesicles and its relationship to sperm motility: effects of infection in the male reproductive tract. Int J Androl 1989;12:286-294.
- Groves ML. Minor milk proteins and enzymes. In: McKenzie HA, ed. Milk Proteins, Chemistry and Molecular Biology. Vol 2. New York: Academic Press; 1971:367-418.
- Hekman A, Rümke P. The antigens of human seminal plasma. Prot Biol Fluids 1968;16:549-552.
- Krause W. Concentration of oestradiol in serum and testis tissue in patients with fertility disorders. *Fertil Steril* 1988;49:926–927.
- Levalle OA, Zylberstein C, Aszpis S, Mariana V, Ponzio R, Aranda C, Guitelman A, Scaglia HE. Serum luteinizing hormone pulsatility and intratesticular testosterone and oestradiol concentrations in idiopathic infertile men with high and normal follicle stimulating hormone serum concentrations. *Hum Reprod* 1994;9:781–787.
- Lizana J, Eneroth P, Bystrom B, Bygdeman M. Studies on the consistency of transudated and locally produced proteins in human seminal plasma. Int J Fertil 1987;32:71-76.
- Mancini G, Carbonara AO, Heremans JF. Immunological quantitation of antigens by single radial immunodiffusion. *Immunochemistry* 1965;2: 235-254.
- Masson PL. La Lactoferrine. Brussels, Belgium: Editions Arscia SA; 1970.
- Masson PL, Heremans JF. Studies on lactoferrin, the iron-binding protein of secretions. *Prot Biol Fluids* 1966;14:115-124.
- Masson PL, Heremans JF, Dive C. An iron-binding protein common to many external secretions. *Clin Chim Acta* 1966;14:735-739.
- Schäfer KH. Elektrophoretische untersuchungen zum milcheiweissproblem. Monatsschr Kinderheilk 1951;99:69.
- Tauber PF, Zaneveld LJD, Propping D, Schumacher GFB. Components of human split ejaculates. I. Spermatozoa, fructose, immunoglobulins, albumin, lactoferrin, transferrin and other plasma proteins. J Reprod Fertil 1975;43:249-267.
- Wolff H, Anderson DJ. Immunohistochemical characterization and quantitation of leukocyte subpopulations in human semen. *Fertil Steril* 1988;49:497-504.
- Wolff H, Politch JA, Martinez A, Haimovici F, Hill JA, Anderson DJ. Leucocytospermia is associated with poor semen quality. *Fertil Steril* 1990;50:129–132.
- World Health Organization. WHO Laboratory Manual for the Examination of Human Semen and Sperm-Cervical Mucus Interaction. 3rd ed. Cambridge: Cambridge University Press; 1992.
- Yu LC, Chen YH. The developmental profile of lactoferrin in mouse epididymis. *Biochem J* 1993;296:107-111.