# Saving implants! BMP-2 application in revision total hip surgery

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#### Abstract

**Objective:** Besides others, there are two major problems in total hip replacement surgery which result in implant failure. First there is aseptic loosening due to a lack of implant biocompatibility or micromovements and second periimplant wear debris induced osteolysis which limits the survival rate of an implant. Regarding to recent data there are only limited therapeutic strategies to heal these bony defects without arthroplasty revision surgery. Since the investigation and characterization of adult mesenchymal stem cells (MSCs) from bone marrow, a cell and tissue engineering based therapy might be a promising solution to heal endoprosthesis associated bony defects. Moreover the application of growth factors in bone reconstructive surgery is another treatment concept to promote local bone regeneration.

**Patient and Methods:** We report about a 73-year old patient with a painful weight bearing and a large, wear debris induced pelvic osteolysis after total hip arthroplasty. To prevent from salvage surgical procedures and preserve bone, a healing attempted was performed by filling the critical bony defect zone with a BMP-2/MSC composit.

**Results:** Clinical and radiological follow-ups showed a progressive bony healing of the critical size defect area without any complications. Fifteen months after application the patient is still pain free, has no limitations in daily life or sport activities.

**Conclusion:** The case embarks on a strategy of non-embryonic stem cell and growth factor application to heal bony defects at patients with total hip endoprosthesis.

Keywords: Arthroplasty; BMP; Bone defect; Stem cells; Total hip replacement

## Introduction

Up to date total hip replacement is the golden standard for the treatment of painfull osteoarthritis of the hip in advanced stages. Although the average survival rate of most implants is more than 10 years, the increase of the average lifetime of the population will make revision surgery to a bigger problem in near future.

Besides low grade infections and septic loosenings, the debris and cement disease are major reasons for an early aseptic implant loosening after total hip replacement (Jäger et al., 2003). Depending on the particle size and chemical properties, polyethylene particles and metallosis can induce foreign body reactions which may induce local inflammation and peri-prosthetic osteolysis. As a biomechanical consequence, these local bone defects conduct progressive implant micromovements and may lead to endoprosthesis failure. Consequently various grades of critical bone defects (CSD) are more often

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noted in total hip revision surgery. Moreover the alternatives to reconstruct bony defects caused by wear debris are limited.

The use of allo- or autogenic bone grafts, the application of bone substitutes (biomaterials), osteosynthesis implants, or large revision arthroplasties are characterized by a lack of biological activity for bone regeneration and / or include biomechanical disadvantages. Therefore these techniques may lead to poor results in several cases. Especially the reconstruction of large bony defects at the pelvic site of a cup implant is technical demanding and bares a high risik of complications. Allogene transplantation of bone for the treatment of bony defects requires complex logistics affords. Besides high costs it is associated with several problems including infection risks, whereas autologous bone marrow transplantation is characterized by a donor site morbidity and limited resources. Furthermore it was shown in the literature that allograft bone transplants decreases in strength and weakens the periimplant region in vivo over time (Wheeler et al., 2005). As demonstrated by Kessler et al. (2003) specially the duration of the revision surgery appears as a predictive parameter for perioperative morbidity in revision total hip arthroplasty.

One promising option in the treatment of osseous defects in endoprothesis revision surgery may be the local application of bone marrow derived mesenchymal stem cells (MSCs). It was shown by several investigators that the human bone marrow accomodates multipotent progenitor cells, which can differentiate into osteoblasts, chondroblast, myoblasts and adipoblasts (Caplan et al., 1991; Gregory et al., 2005; Bruder et al., 1994; Pittenger et al., 1999; Gronthos et al., 2003; Majors et al., 1997). Other authors demonstrated that the number of bone marrow derived MSCs can be significantly increased by vacuum aspiration techniques (Muschler et al., 1997, 2001, 2002).

Although it has been shown in several pre-clinical studies that MSCs are potent to regenerate osteoblasts in vitro and in vivo (Bensaid et al., 2005) additional osteogenic stimuli may increase the osteoblastic potency of these cells. However, it is unclear if this strategy is helpful to heal large periimplant osteolytic defects.

An option to enhance the recruitment and promote the differentiation of osteoblasts in progenitor cells is the application of growth factors. One promising group of growth factors are the multi-functional bone morphogenetic proteins (BMPs). Belonging to the transforming growth factor beta (TGF- $\beta$ ) superfamily, the roles of BMPs in embryonic development and for cellular functions in postnatal and adult animals have been extensively studied in recent years. BMP signaling plays a crucial role in heart, neural and cartilage development at different molecular levels. Moreover it is shown that the BMP-2 subtype is one of the most promising bone promoting growth factors, which was already applied to heal local bone defects in pre-clinical and clinical trials (Schmitt et al., 1999).

In orthopedic and dental surgery BMP-2 was successfully applied to heal open tibial fractures, promote spinal fusions and augmentate or preserve the alveolar bone in the dental ridge (Kraiwattanapong et al., 2005; Govender et al., 2002; Valentin-Opran et al., 2002).

Although BMP-2 can accelerate or improve bone healing when used alone, long healing times and abnormalities, such as cystic areas or abnormal microarchitecture of the new formed bone still occur (Liebermann et al., 2002; Sharp et al., 2005). Therefore there is a rational for an interest to combine BMPs with MSCs to heal bony defects.

Up to date there are only few clinical and experimental data about the application of BMPs in pelvis or hip. We report on a 73-year old male patient with severe debris induced acetabular osteolysis who has been treated by a BMP-2/MSC composit.

#### **Patient and Methods**

A 73-year old male patient with progressive pain in the left groin and gluteal region, which occurred since January 2004 was transmitted to our department. The patient underwent total hip replacement eight years ago (54 mm in diameter cementless threaded cup type "Münchner Schraubring" combined with a cementless stem type "Bicontact" / Merck, Germany and a ceramic head size medium).

The x-rays of the pelvis in two standard planes showed a condition after total hip arthroplasty with a large osteolytic tumor medial to the cup  $(7.5 \times 4.0 \times 3.5$ cm in diameter, corresponding to 90 cm<sup>3</sup>). Furthermore there were only slight signs of a radiolucent line at the proximal femur corresponding to Gruen zones 1, 2 and 7 (Gruen et al., 1979) without any additional signs of aseptic loosening (figure 1).

Further diagnoses were a paroxysmal atrial fibrillation, arterial hypertension and a prostata carcinoma, which was treated by a transurethral protatectomy five years ago.

To exclude a malignant tumor we took a biopsy of the osteolytic pelvic area performing an anterior surgical approach to the pelvis. The histopathological representative tissue scanned under polarization microscopy showed a birefringent material corresponding to polyethylene wear debris. The laboratory blood parameters were uneventfull. In a second operation we replaced the worn out polyethylene liner (figure 2). At this time the intraoperative examination of the prosthesis showed no sings of implant instability. To avoid a total cup replacement a healing attempt with a BMP-2/MSC composit was recommended to the patient. In accordance to the Declaration of Helsinki in its present form we informed the patient about BMP-2 associated perioperative risks including ectopic bone formation, allergic reactions and the formation of antibodies against BMP-2. The patient agreed with underwritten, informed consent.

Eight days later the osteolytic pelvic area was curettaged in a third stage surgery. Afterwards a total volume of 20 ml bone marrow aspirate at the posterior ilium was taken by Jamshidi-vacuum aspiration. Furthermore autologous spongiosa was obtained from the anterior iliac crest. Two collagen I/III sponges were sprinkled with 24 mg BMP-2 (Induct Os<sup>™</sup>/Sofamor).



Figure 1. The X - rays of the pelvis in anterior-posterior (a) and axial planes (b) showed a thread cup with a large, expansive critical bony defect corresponding to a wear induced osteolysis. Moreover there are slight radiolucent lines at the proximal femur in Gruen-zones 1 (G1), 2 (G2) and 7 (G7).

To guarantee a sufficient protein adherence on the collagen surface we incubated the sponges for 15 min. at room temperature.

Afterwards the bone marrow cell suspension was distributed over the collagen I/III carrier and incubated for another 5 min. The bony defect cavity was filled in a multilayer sandwich-technique by collagen sponges, BMP-2/MSC-composits and allogen and autogen spongious transplants as shown in figure 3. The total volume of the autologous bone transplant was less than one third of the whole pelvic defect.



Figure 2. The liner of explanted thread cup showed a decentralized polyethylene wear (arrrow). Corresponding to a liner-neck impingement of the implant we found a brown-grey change in color on the polyethylene rim. Because the metal cup was designed with a central gap polyethylene particles could migrate behind the implant and cause wear debris inducing osteolysis at the inner pelvic site.

#### Results

The postoperative phase was uneventfull without

any perioperative complication. The patient was mobilized on crutches with limited weight bearing of 20 kg at the left lower extremity for the first 12 weeks. After initial wound healing the patient was free of pain within 7 days. The patient was discharged after three weeks postoperatively and underwent a further physiotherapy program. X-ray controls of the pelvis in two standard planes after two, six, and 12 weeks showed a progressive osseous healing with newly formed bone in the osteolytic area (figure 4). There were no immunoreactive antibodies against BMP-2 shown by serum immunophoresis after 12 weeks.



Figure 3. a) The figure shows two collagen I/III strips (\*) which were impregnated by BMP-2, covered by MSCs containing autologous human bone marrow which was harvested by vacuum aspiration and augmented by spongiosa chips from iliac crest (arrows). b) The x-ray on a.p. view shows the multilayer sandwich-technique of refilling the critical bony osteolysis behind the cup. To lower the risk of an ectopic bone formation at the inner pelvic site collagen I/III sponges should act as a barrier and were positioned at the medical site of the reconstructive area (Col). Allograft bone was augmented by MSC containing bone marrow aspirate (MSC/allograft) and BMP-2 containing collagen I/III sponge, were incubated with MSCs (MSC/BMP-2). Col: collagen I/III sponge, MSC(s): bone marrow derived mesenchymal stem cell(s), ASp: autologous spongiosa, BMP: bone morphogenic protein.

Six months after BMP-2/MSC application CT scans proof that the osteolytic cavity was completely filled out with solid bone (figure 5). At this time and at latest follow up (fifteen months after surgery) the patient was free of pain and the range of motion (ROM) was unchanged compared to the preoperative status (flexion/extension:  $110^{\circ}/0^{\circ}/10^{\circ}$ , external/internal rotation:  $40^{\circ}/0^{\circ}/30^{\circ}$ , abduction/adduction:

 $30^{\circ}/0^{\circ}/10^{\circ}$ ). Three months after surgery the patient returned to sport activities (biking and hiking) and had no limitations in walking distances or daily life activities.



Figure 4. The figure shows x-rays in two standard planes of pelvis / hip. The roentgenological follow-ups before surgery and after two, six and 12 weeks after BMP-2/MSC-tretament show a progressive bony healing of the osteolytic area behind the cup.

As far as we know this is the first successfull BMP-2/MSC application in wear induced osteolysis of the pelvis.

## Discussion

In this study we showed that a local BMP-2/MSC application can promote bone formation within a wear debris induced peri-prosthetic osteolytic area. Our result corresponds to data of other authors in literature. Mont et al. (2004) applied BMP-enriched allografts in osteonecrosis of the femoral head to avoid donor site morbidity in 19 patients. They implanted a combination of demineralized bone matrix (DBM), processed allograft bone chips, and a thermoplastic carrier through

a window at the femoral head-neck junction. Mont et al. (2004) presented a clinical success in 86% of the cases after a mean follow-up of 48 months. The positive influence of MSCs for bone regeneration is confirmed by Muschler et al (2005) who showed in a canine spinal fusion model that a MSC-enriched matrix graft has advantages in bone healing compared to a matrix graft which is augmented by bone marrow.

In another study Bragdon et al. (2003) applied BMP-2 in a canine total hip arthroplasty model at the porous acetabular component and demonstrated a local promotion of the defect filling and bone ingrowth into the porous coating beneath the defect region by rhBMP-2/calcium phosphate cement ( $\alpha$ -BSM<sup>®</sup>).

Murakami et al. (2003) investigated the rhBMP-2

related bone formation using poly-D,L-lactic-acid-paradioxanone-polyethyleneglycol block co-polymer (PLA-DX-PEG) carrier in a canine model. To induce a critical femoral defect, half of the proximal femur was surgically resected and treated by the BMP-2/PLA-DX-PEG composit. They showed a bone repair after a 12 weeks follow-up and advocate the use of BMP-2-"hybrid" prosthesis as a new modality to repair bone defects in endoprosthesis revision surgery.





Figure 5. CT-scan before (a) and six months after (b) implantation of a BMP2/MSC-composit which was augmented by autologous and allogen spongious bone into an osteolytic pelvic cavity. a) The wear debris induced pelvic osteolysis lead to a pseudo-tumour with inflammative soft tissue. The inner cortical wall behind the cup is completely destructed whereas a cortical ring allows still a sufficient fixation of the thread cup. As a result to foreign body reaction ipsilateral pelvic organs are shifted by inflammatory tissue. The border of the expanding pseudotumour is marked by arrows. b) Six months after cuerettage of the pseudo-tumour and implantation of BMP-2/MSC-composit the initial critical size osteolysis is replaced by bone.

In contrast to the promising experimental and clinical

data mentioned before some authors report about negative results after BMP application. McGee et al. (2004) investigated in their pilot study the effect of BMP-7 on bone graft incorporation in sheep' femurs impacted with allograft after cemented hemiarthroplasty. They found an increased initial graft resorption and associated stem subsidence in the BMP-7 group. McGee et al. (2004) pointed out the demand of further studies examining dose response effects. They advised against clinical application of BMP-7 in femoral impaction grafting at revision hip arthroplasty.

In contrast to McGee et al. (2004) we used a BMP-2/ MSC composit to promote the local healing in wear debris induce osteolysis which was augmented by allograft an autologous spongiosa.

Although it is demonstrated in several studies that BMP-2 promotes an ectopic bone formation in animal MSCs, there is still a controversial discussion about the effects of BMP-2 on human MSCs in literature.

Diefenderfer et al. (2003) investigated the BMP signaling pathways in human bone marrow derived MSCs. In their study with MSC samples from more than a dozen patients, only one patient sample showed a significantly elevated alkaline phosphatase (ALP) after exposure to BMP whereas the rest responded to dexamethasone but not to BMP. As a consequence Diefenderfer et al. (2003) concluded that dexamethasone is required for induction of the osteoblast phenotype. The importance of dexamethasone presence and its necessity in osteoblast differentiation pathways (Diefenderfer et al., 2003) correspond to the data of our group published previously (Jäger et al., 2005).

Based on trauma related stress it is evident that the glucocorticoid levels are increased in patients after surgery. As demonstrated by Wellby et al. (1994) it is evident that the cortisol concentration increases after 30 minutes and peaked at four hours after elective surgeries.

In conclusion the in vivo bone formation promoted by BMP-2/MSC composit may not only base on BMP-2/ MSCs but also influenced by surgery related increased level of glucocorticoides. Furthermore sex and age of a patient may also influence BMP/MSC associated bone healing. Muschler et al. (1997, 2001, 2002) reported a significant age-related decline in the number of nucleated cells harvested per bone marrow aspirate for both men and women. However, they demonstrated that the number of colony forming units (CFU) which express alkaline phosphatase decreased significantly with age for women, but not for men.

Diefenderfer et al. (2003) showed advantages of BMP-2 for the differentiation of osteoblasts in MSCs compared to BMP-4 and -7. They presented a significantly higher induction of ALP in males.

Another factor which may influence the osteoinductive effects of BMP-2 and MSCs in vivo is the applied scaffold. In this case report we used a collagen I/III carrier as a BMP-2 and MSC scaffold. The finding that collagen I/III as a suitable carrier for MSCs with a low degree of cytotoxicity and osteoinductive properties is confirmed by our in vitro-studies published previously (Jäger et al. 2004, 2005, 2006). The qualification of three dimensional collagen I/III as an appropriate scaffold for MSCs based bone and cartilage engineering was also confirmed by Yoneno (2005) and Liu (2005). It is evident that especially collagen I, a normal constituent of bone, plays an important role in osteoblast differentiation and function. Kihara et al. (2006) showed on rat MSCs that osteoblastic differentiation is enhanced by adding solubilized type I collagen. Moreover Farrel et al. (2006) demonstrated that a collagen-I-glycosaminoglycan composit allows an osteogenic differentiation of rat MSCs in vitro. These data are coressponding to Chastain et al. (2006) who showed that collagen I acts as a mediator for MSCs to attach to polylactide scaffolds.

A promising alternative to BMP-2 application to enhance the osteogenic potency of MSCs may the use of BMP-2 transfected stem cells. Tsuda et al. (2005) reported about an enhanced osteoinduction by MSCs transfected with a fiber-mutant adenoviral BMP-2 gene in rats. In

another study Dai et al. (2005) demonstrated on a goat diaphyseal defect that a combination of biphasic calcined bone (BCB) and MSCs transduced with hBMP-2 significantly improved the bone healing. However, it is unclear if gene therapeutic strategies will fulfil the demands of biological safety and efficiency under clinical application.

Although this case showed a rapid healing of the critical size osseous defect, there is no evidence that BMP-2 application was solely responsible for healing of the severe osteolysis.

However, there are some arguments that BMP-2 supports the bone healing significantly in our case: The patient was an elderly men and not a young person with high bone healing potential (1). The defect zone was critical in size for a spontaneous healing (2) and the total volume of the autologous bone graft was less than one third of the whole defect (3). The presence of wear debris and inflammative soft tissue has a negative influence on bone healing (4), but the patient showed a rapid bone healing within 12 weeks after the MSC/BMP-2 transplantation (5).

It is not clear if the patient would have benefit from a two-stage surgical procedure (1. biopsy, 2. implant exchange and defect filling) which would reduce costs and hospital stay. Overall, individual factors (co-morbidity, size of defect, quality of bone, surgical opportunities and experiences, demands of the patient for sports and daily activities) has to be considered in decision making for or against an BMP-2 application in the treatment of patients with a periimplant osseous defect. In contrast to a conventional one-stage revision surgery including an implant exchange and defect filling by bone substitutes (autologous / allogene bone grafs, hydroxyapatite, tricalciumphosphate) a two or three-stage procedure has the disadvantage of repititive anaesthesias and prolonged hospital stay.

#### Conclusion

Considering individual facts the local application of

BMP-2/MSC may restore lost bone mass encountered in revision arthroplasty.

#### References

- Bensaid W, Oudina K, Viateau V, Pertier E, Bousson V, Blanchat C, Sedel L, Guillemin G, Petite H. De novo reconstruction of functional bone by tissue engineering in the metatarsal sheep model. Tissue Eng 2005; 11(5-6): 814-24.
- Bragdon CR, Doherty AM, Rubash HE, Jasty M, Li XJ, Seeherman H, Harris WH. The efficacy of BMP-2 to induce bone ingrowth in a total hip replacement model. Clin Orthop Relat Res 2003; 417: 50-61.
- Bruder SP, Find DJ, Caplan AI. Mesenchymal stem cells in bone development, bone repair, and skeletal regeneration therapy. J Cell Biochem 1994; 56: 283-94.
- 4. Caplan AI. Mesenchymal stem cells. J Orthop Res 1991; 9: 641-50.
- Chastain SR, Kundu AK, Dhar S, Calvert JW, Putnam AJ. Adhesion of mesenchymal stem cells to polymer scaffolds occurs via distinct ECM ligands and controls their osteogenic differentiation. J Biomed Mater Res A 2006 (in press).
- Dai KR, Xu XL, Tang TT, Zhu ZA, Yu CF, Lou JR, Zhang XL. Repairing of goat Tibial Bone Defects with BMP-2 Gene-Modified Tissue-Engineered Bone. Calcif Tissue Int 2005; 77(1): 55-61.
- Diefenderfer DL, Osyczka AM, Reilly GC, Leboy PS. BMP responsiveness in human mesenchymal stem cells. Connect Tissue Res 2003; 44 Suppl 1: 305-11.
- Diefenderfer DL, Osyczka AM, Garino JP, Leboy PS. Regulation of BMPinduced transcription in cultured human bone marrow stromal cells. J Bone Joint Surg 2003; 85-A Suppl 3: 19-28.
- Farrell E, O'brien FJ, Doyle P, Fischer J, Yannas I, Harley BA, O'connell B, Prendergast PJ, Campbell VA. A Collagen-glycosaminoglycan Scaffold Supports Adult Rat Mesenchymal Stem Cell Differentiation Along Osteogenic and Chondrogenic Routes. Tissue Eng. 2006; 12(3): 459-68.
- 10. Govender S, Csimma C, Genant HK, Valentin-Opran A, Amit Y, Arbel R, Aro H, Atar D, Bishay M, Borner MG, Chiron P, Choong P, Cinats J, Courtenay B, Feibel R, Geulette B, Gravel C, Haas N, Raschke M, Hammacher E, van der Velde D, Hardy P, Holt M, Josten C, Ketterl RL, Lindeque B, Lob G, Mathevon H, McCoy G, Marsh D, Miller R, Munting E, Oevre S, Nordsletten L, Patel A, Pohl A, Rennie W, Reynders P, Rommens PM, Rondia J, Rossouw WC, Daneel PJ, Ruff S, Ruter A, Santavirta S, Schildhauer TA, Gekle C, Schnettler R, Segal D, Seiler H, Snowdowne RB, Stapert J, Taglang G, Verdonk R, Vogels L, Weckbach A, Wentzensen A, Wisniewski T. Recombinant human bone morphogenetic protein-2 for treatment of open tibial fractures: a prospective, controlled, randomized study of four hundred and fifty patients. J Bone Joint Surg 2002; 84-A(12): 2123-34.
- Gregory CA, Prockop DJ, Spees JL. Non-hematopoietic bone marrow stem cells: molecular control of expansion and differentiation. Exp Cell Res 2005; 306(2): 330-5.
- 12. Gronthos S, Zannettino AC, Hay SJ, Shi S, Graves SE, Kortesidis A, Sim-

mons PJ. Molecular and cellular characterisation of highly purified stromal stem cells derived from human bone marrow. Cell Sci 2003; 116(Pt 9): 1827-35.

- Gruen TA, McNeice GM, Amstutz HC (1979) Modes of failure of cemented stem-type femoral components: a radiographic analysis of loosening. Clin Orthop 1979; 141: 17-26.
- Jäger M, Wilke A. Comprehensive biocompatibility testing of a new PMMA-hA bone cement versus conventional PMMA cement in vitro. J Biomater Sci Polym Ed 2003; 14(11): 1283-98.
- Jäger M, Feser T, Denck H, Krauspe R. Proliferation and Osteogenic Differentiation of Mesenchymal Stem Cells Cultured onto Three Different Polymers in vitro. Ann Biomed Eng 2005; 33(10): 1348-61.
- Jäger M, Fischer J, Schultheis A, Krauspe R. Extensive H+ release by bone substitutes affects biocompatibility in vitro testing. J Biomed Mater Res A 2006; 76(2): 310-22.
- Kessler S, Kinkel S, Kafer W, Puhl W, Schochat T. Influence of operation duration on perioperative morbidity in revision total hip arthroplasty. Acta Orthop Belg 2003; 69(4): 328-33.
- Kihara T, Hirose M, Oshima A, Ohgushi H. Exogenous type I collagen facilitates osteogenic differentiation and acts as a substrate for mineralization of rat marrow mesenchymal stem cells in vitro. Biochem Biophys Res Commun. 2006 Mar 24; 341(4): 1029-35.
- Kraiwattanapong C, Boden SD, Louis-Ugbo J, Attallah E, Barnes B, Hutton WC. Comparison of Healos/bone marrow to INFUSE(rhBMP-2/ ACS) with a collagen-ceramic sponge bulking agent as graft substitutes for lumbar spine fusion. Spine 2005; 30(9): 1001-7.
- Lieberman JR, Daluiski A, Einhorn TA: The role of growth factors in the repair of bone: Biology and clinical applications. J Bone Joint Surg 2002; 84-A: 1032-44.
- Liu Y, Sun S, Singha S, Cho MR, Gordon RJ. 3D femtosecond laser patterning of collagen for directed cell attachment. Biomaterials 2005; 26(22): 4597-605.
- Majors AK, Boehm CA, Nitto H, Midura RJ, Muschler GF. Characterization of human bone marrow stromal cells with respect to osteoblastic differentiation. J Orthop Res 1997; 15(4): 546-57.
- 23. McGee MA, Findlay DM, Howie DW, Carbone A, Ward P, Stamenkov R, Page TT, Bruce WJ, Wildenauer CI, Toth C. The use of OP-1 in femoral impaction grafting in a sheep model. J Orthop Res 2004; 22(5): 1008-15.
- Mont MA, Etienne G, Ragland PS. Outcome of nonvascularized bone grafting for osteonecrosis of the femoral head. Clin Orthop 2003; (417): 84-92.
- 25. Murakami N, Saito N, Takahashi J, Ota H, Horiuchi H, Nawata M, Okada T, Nozaki K, Takaoka K. Repair of a proximal femoral bone defect in dogs using a porous surfaced prosthesis in combination with recombinant BMP-2 and a synthetic polymer carrier. Biomaterials 2003; 24(13): 2153-9.
- Muschler GF, Boehm C, Easley K. Aspiration to obtain osteoblast progenitor cells from human bone marrow: the influence of aspiration volume. J Bone Joint Surg 1997; 79-A(11): 1699-709.
- Muschler GF, Nitto H, Boehm CA, Easley KA. Age- and gender-related changes in the cellularity of human bone marrow and the prevalence of

osteoblastic progenitors. J Orthop Res 2001; 19(1): 117-25.

- Muschler GF, Midura RJ. Connective tissue progenitors: practical concepts for clinical applications. Clin Orthop 2002; 395: 66-80.
- Muschler GF, Matsukura Y, Nitto H, Boehm CA, Valdevit AD, Kambic HE, Davros WJ, Easley KA, Powell KA. Selective retention of bone marrowderived cells to enhance spinal fusion. Clin Orthop 2005; 432: 242-51.
- Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, Moorman MA, Simoneti DW, Craig S, Marshak DR. Multilineage Potential of Adult Human Mesenchymal Stem Cells. Science 1999; 284:143-7.
- Schmitt JM, Hwang K, Winn SR, Hollinger JO. Bone morphogenetic proteins: an update on basic biology and clinical relevance. J Orthop Res 1999; 17(2): 269-78.
- 32. Sharp JG, Murphy BO, Jackson JD, Brusnahan SK, Kessinger A, Neff JR. Promises and pitfalls of stem cell therapy for promotion of bone healing. Clin Orthop 2005; 435: 52-61.
- 33. Tsuda H, Wada T, Yamashita T, Hamada H. Enhanced osteoinduction by mesenchymal stem cells transfected with a fiber-mutant adenoviral

BMP2 gene. J Gene Med 2005; 7(10): 1322-34.

- Valentin-Opran A, Wozney J, Csimma C, Lilly L, Riedel GE. Clinical evaluation of recombinant human bone morphogenetic protein-2. Clin Orthop 2002; 395:110-20.
- Wellby ML, Kennedy JA, Barreau PB, Roediger WE. Endocrine and cytokine changes during elective surgery. J Clin Pathol 1994; 47(11): 1049-51.
- 36. Wheeler DL, Enneking WF. Allograft bone decreases in strength in vivo over time. Clin Orthop 2005; 435: 36-42.
- Wild A, Jäger M, Lensing-Höhn S, Werner A, Krauspe R. Growth behaviour of human mononuclear cells derived from bone marrow and cord blood on a collagen carrier for osteogenic regeneration. Biomed Tech (Berl) 2004; 49(9): 227-32.
- 38. Yoneno K, Ohno S, Tanimoto K, Honda K, Tanaka N, Doi T, Kawata T, Tanaka E, Kapila S, Tanne K. Multidifferentiation potential of mesenchymal stem cells in three-dimensional collagen gel cultures. J Biomed Mater Res A 2005; 75(3): 733-41.