

转导单链免疫毒素基因的 PBMCs 对人肝癌细胞 SMMC-7721 的体外杀伤活性

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In vitro cytotoxicity of PBMCs via genetic modification of single-chain immunotoxin

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Abstract

AIM: To investigate the selective cytotoxicity of single-chain immunotoxin (sFv-TNF- α fusion proteins) in cell line SMMC-7721.

METHODS: HCC-specific killer cells were generated by transducing the recombinant retroviral virus in supernatant of the virus producing cells into human peripheral blood mononuclear cells (PBMCs). PCR and RT-PCR were used to detect integration and transcription of the sFv-TNF- α gene in transduced PBMCs (PBMCs/PST). MTT method was used to detect antitumour activity of the sFv-TNF- α fusion proteins.

RESULTS: There was integrated sFv-TNF- α gene in the genome of PBMCs/PST, and PBMCs/PST were able to express the fusion sFv-TNF- α proteins. Cell killing was significant in HCC cells co-cultivated with PBMCs/PST, whereas the PBMCs/pLXSN control cells had no significant cytotoxic effects on HCC cells.

CONCLUSION: Expression of sFv-TNF- α fusion proteins in PBMCs/PST has cytotoxicity to HCC cells *in vitro*.

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摘要

目的: 用携带分泌型抗肝癌单链免疫毒素基因(sFv-TNF- α)的重组逆转录病毒感染人外周血单个核细胞(peripheral blood mononuclear cells, PBMCs), 使其表达并分泌针对人肝癌细胞的 sFv-TNF- α 融合蛋白, 观察转导的 PBMCs 对体外培养人肝癌细胞 SMMC-7721 的杀伤作用。

方法: 用感染性重组病毒产生细胞 C₂₂(PA317/PST) 产生的病毒上清转导人 PBMCs, 采用 PCR 和 RT-PCR 方法对转导的 PBMCs(PBMCs/PST) 进行 DNA 和 mRNA 水平的分析。PBMCs/PST 与 SMMC-7721 共培养, MTT 法检测 PBMCs/PST 表达产物对肝癌细胞的体外杀伤活性。

结果: PCR 及 RT-PCR 结果显示 PBMCs/PST 中扩增出外源目的基因对应的电泳条带。MTT 法检测结果, 分泌型抗肝癌单链免疫毒素对体外培养肝癌细胞 SMMC-7721 的杀伤率为 (38.2 \pm 6.7)%。

结论: 分泌型抗肝癌单链免疫毒素基因可以在 PBMCs 中整合并稳定表达, 其分泌的表达产物对 SMMC-7721 具有一定的体外杀伤作用。

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0 引言

肝细胞肝癌(hepatocellular carcinoma, HCC) 是常见的恶性肿瘤之一, 位居我国恶性肿瘤发病的第三位, 约占全世界肝癌病例的 42.5%, 每年死于肝癌的人数约为 11 万^[1-7]。迄今为止, 肝癌的病因和发病机制仍不十分清楚, 更无理想的治疗手段^[8-18]。随着现代分子生物学和免疫学的发展, 自 1990 年代初迄今, 已有百余项肿瘤基因治疗方案获准进入临床试验, 但纵观近 10 a 的研究发现, 其临床疗效与预期的目标之间还存在较大差距, 原因之一就是肿瘤细胞的特异性识别尚未解决。肿瘤特异性单链抗体的问世为基因治疗靶向性问题的解决提供了新的手段, 近年来成为肿瘤免疫基因治疗中的热点, 目前已有多种单链免疫毒素基因治疗计划进入临床试验^[19-27]。在本实验室成功克隆的抗肝癌单链抗体基础上, 我们构建了分泌型抗肝癌单链免疫毒素基

因逆转录病毒表达载体 pLXSN-sFv-TNF- (PST), 并用 PA317 细胞进行包装, 筛选出一株具有较高滴度的感染性重组病毒产生细胞 C₂₂(PA317/PST)^[28]. 用 C₂₂ 细胞产生的病毒上清转导人外周血单个核细胞(peripheral blood mononuclear cells, PBMCs), 使其表达并分泌针对人肝癌细胞的靶向性 sFv-TNF- 融合蛋白, 观察转导的 PBMCs 对体外培养肝癌细胞的杀伤作用.

1 材料和方法

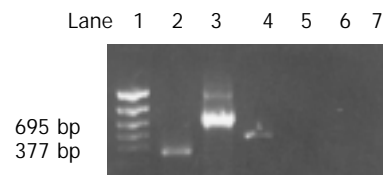
1.1 材料 人肝癌细胞系 SMMC-7721 为本室保存, RPMI1640 常规传代培养. 转染的逆转录病毒产生细胞系 C₂₂ 由本室构建^[28], 用 DMEM 常规传代培养. DMEM, G418, Superscript™ 逆转录试剂盒和 Trizol Reagent 为 Gibco 公司产品, Polybrene 为 Sigma 公司产品, MTT 为 Serva 公司产品, PHA 购自 Sigma 公司, 超级小牛血清购自杭州四季青生物工程材料研究所. 重组人 IL-2 为上海生物技术研究所产品. 淋巴细胞分离液购自上海试剂二厂.

1.2 方法 抽取正常人外周血 50 mL, 肝素抗凝, 加入淋巴细胞分离液 50 mL, 室温 1 400 g 离心 30 min, 吸取单个核细胞层悬液, 40 mL 无血清 RPMI1640 洗涤细胞 3 次(1 000 g 离心 10 min), 计数, 用含 200 mL/L 小牛血清的完全 RPMI1640 培养液稀释为活细胞 $1 \times 10^6 \cdot L^{-1}$, 加入 $5 \times 10^5 \cdot L^{-1}$ IL-2 及 $5 \times 10^3 \mu g \cdot L^{-1}$ PHA, 于 37 °C, 50 mL $\cdot L^{-1}$ CO₂ 培养 3 d, 换含 $5 \times 10^5 \cdot L^{-1}$ IL-2, 200 mL/L 小牛血清的完全 RPMI1640 培养液继续培养 3 d 后, 进行逆转录病毒的转导^[29]. 用 C₂₂ 细胞制备重组逆转录病毒上清^[28], 取 5 mL C₂₂ 细胞上清(不含 G418)及 8 mg $\cdot L^{-1}$ Polybrene 感染 1×10^6 个 PBMCs, 置 37 °C, 50 mL $\cdot L^{-1}$ CO₂ 孵箱内培养 6 h, 换普通完全 RPMI1640 培养液, 连续感染 3 d, 转导的 PBMCs 命名为 PBMCs/PST. 对照组 PBMCs 用 PA317/pLXSN(空白载体)细胞上清感染, 转导的 PBMCs 命名为 PBMCs/pLXSN. 病毒连续感染 3 d 后 4 d 转入含 500 mg $\cdot L^{-1}$ G418 的完全 RPMI1640 培养液中培养 1 wk. 分别提取重组逆转录病毒转导的 PBMCs/PST 和 PBMCs/pLXSN 细胞中的基因组 DNA^[28], 用 3 对引物(sFv-1 和 sFv-2, TNF-1 和 TNF-2 及 neo-1 和 neo-2), 分别扩增细胞基因组 DNA 中 sFv, TNF- 及 neo 基因, 以证实外源基因导入靶细胞. PCR 反应于 25 μL 体积中完成, 基因组 DNA 模板 2.5 μL , 引物 2.5 μL , 10 \times PCR 缓冲液 2.5 μL , 2.5 mmol $\cdot L^{-1}$ dNTP 2.5 μL , TaqDNA 聚合酶 0.5 μL , 加水补足 25 μL , 混匀后在 PCR 仪上扩增. 循环参数为: 94 °C 变性 60 s, 58 °C 退火 60 s, 72 °C 延伸 90 s, 经 30 个循环后, 72 °C 保温 10 min. PCR 产物于 15 g $\cdot L^{-1}$ 琼脂糖凝胶电泳. 紫外透射仪观察 PBMCs/PST 细胞基因组 DNA 中的外源基因. 采用 Trizol 一步法, 分别提取重组逆转录病毒转导的 PBMCs/PST 和 PBMCs/pLXSN 细胞中的总 RNA. 利用 Superscript™ 系统, 以细胞内总 RNA 为模板进行 cDNA 第一链的合成: 3.5 μL RNA, 2 μL 5 \times 逆转录缓冲液, 1 μL Oligo(dT)₁₂, 70

保温 10 min, 冰浴 2 min, 加入 1 μL 100 mmol $\cdot L^{-1}$ DTT, 1 μL 10 mmol $\cdot L^{-1}$ dNTP, 1 μL RNase inhibitor, 0.5 μL MMLV 逆转录酶, 37 °C 反应 1 h. 以 cDNA 第一链为模板进行 sFv 和 TNF- 基因的扩增, PCR 反应于 50 μL 体积中完成, cDNA 模板 2.5 μL , 5 $\mu mol \cdot L^{-1}$ 引物 5 μL , 10 \times PCR 缓冲液 5 μL , 2.5 mmol $\cdot L^{-1}$ dNTP 5 μL , TaqDNA 聚合酶 0.5 μL , 加水补足 50 μL , 混匀后在 PCR 仪上扩增. PCR 产物于 15 g $\cdot L^{-1}$ 琼脂糖凝胶电泳, 紫外透射仪观察 sFv 和 TNF- 基因的 mRNA. 选择对数生长期的人肝癌细胞 SMMC-7721 接种于 24 孔培养板中, 每孔 2×10^4 , 将 PBMCs/PST 按每孔 2×10^5 接种于孔径 0.4 μm 的 Millicell 滤膜培养皿中, 并将 Millicell 置 24 孔培养板中, 使 PBMCs/PST 与 SMMC-7721 共培养^[29,30]. 设 PBMCs/pLXSN 与 SMMC-7721 共培养为空白对照, PBMCs 与 SMMC-7721 共培养为空白对照, 每种细胞设 20 复孔. 培养板置 37 °C, 50 mL $\cdot L^{-1}$ CO₂ 孵箱内培养, 72 h 后取出 Millicell, 加 MTT 每孔 40 μL 至 24 孔培养板中, 继续培养 4 h, 弃上清, 加入 0.2 mL DMSO 终止反应, 充分振荡后在酶标仪上检测 A_{550nm} 值, 计算细胞杀伤率^[29].

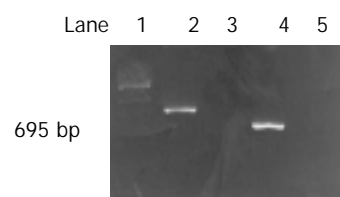
2 结果

2.1 PBMCs/PST 基因组 DNA 中外源基因扩增 PCR 产物电泳后观察, 可见相应的 327 bp, 470 bp, 790 bp 大小的条带, 分别与 neo, TNF-, sFv 基因片段大小相符, 而 PBMCs/pLXSN 中未扩增出这 3 种大小的条带(图 1), 证实 PBMCs/PST 中有外源性目的基因的导入.



1 PCR markers 2 PBMCs/PST (Neo, sFv, TNF-)
5-7 空载体对照 PBMCs/pLXSN (Neo, sFv, TNF-)
图 1 基因组 DNA 中 sFv、TNF- 及 neo 基因 PCR 扩增结果.

2.2 外源基因在 PBMCs/PST 中转录水平的分析 RT-PCR 产物电泳后, 可见相应的 470 bp, 790 bp 大小的条带, 分别与 TNF-, sFv 基因片段大小相符, 对照组未扩增出这两种大小的条带(图 2), 证实 PBMCs/PST 中外源性目的基因有 mRNA 表达.



1 PCR markers 2 PBMCs/PST (sFv) 3 空载体对照 PBMCs/pLXSN (sFv)
4 PBMCs/PST (TNF-) 5 空载体对照 PBMCs/pLXSN (TNF-)
图 2 sFv 和 TNF- 基因 mRNA 的 RT-PCR 检测.

2.3 细胞杀伤率检测 用 MTT 法检测与 PBMCs/PST, PBMCs/pLXSN 共培养 SMMC-7721 的 A_{550nm} (A)值及与 PBMCs 共培养 SMMC-7721 的 A_{550nm} 值(A_0), 计算对 SMMC-7721 细胞的杀伤率(%). 计算公式为: 细胞杀伤率 (%) = $(1 - A/A_0) \times 100\%$. PBMCs/PST($n = 20$)细胞杀伤率为 $(38.2 \pm 6.7)\%$, 而 PBMCs/pLXSN($n = 20$)细胞杀伤率为 $(15.7 \pm 3.1)\%$, 经 t 检验, $P < 0.01$.

3 讨论

实现人类实体瘤基因治疗的途径主要有 2 种, (1)在靶细胞的转染(in vivo), 即直接基因治疗途径, 例如将细胞因子或肿瘤抗原的基因表达载体直接体内注射或将 MHC-I 基因直接注射入瘤体内等治疗肿瘤; (2)离体靶细胞的转染(ex vivo), 即间接基因治疗途径, 必须分离并培养离体的靶细胞, 并在接受基因转移后移植回人体以治疗疾病, 例如应用体外基因修饰的 TIL、瘤苗、成纤维细胞等体内回输或接种的方法治疗肿瘤^[31-34]. 我们选择逆转录病毒介导的体外基因转移方法进行基因治疗的研究, 要求基因转移的靶细胞既要容易获得, 又要容易回输到人体, 因此, 我们采用 PBMCs 作为基因转移的靶细胞. 由于目前所常用的基因转移方法^[35-41](包括逆转录病毒载体-包装细胞转移系统)效率均不高, 获得大量的靶细胞是重要的先决条件, 因此, 要求该细胞易于体外培养, 而且具有一定的分裂和增生能力. 此外, 逆转录病毒载体对处于活跃分裂状态的细胞感染率和基因转移率较高, 而对未分化的分裂不十分活跃的细胞感染率和基因转移率较低, 因此, 基因治疗中应尽量选择处于活跃分裂状态的细胞作为靶细胞. PBMCs 具备上述的各项条件, 目前是一种较理想的基因治疗靶细胞^[42-44]. 虽然有研究表明, TIL 抗肿瘤的活性是 PBMCs 的 50-100 倍, 而且具有良好的肿瘤病灶趋向性和浸润性^[45-49], 但由于其不易分离和培养, 在某些肿瘤如肝癌中 TIL 数量极少, 无法满足基因治疗中反复操作的要求, 因而其应用受到很大的限制.

我们将正常人 PBMCs 分离并用 PHA 和 IL-2 进行体外刺激培养, 用携带分泌型抗肝癌单链单链免疫毒素基因的重组逆转录病毒感染, 使其表达并分泌针对人肝癌细胞的 sFv-TNF- 融合蛋白, 采用 PCR, RT-PCR 方法, 对转导的 PBMCs 进行 DNA 和 mRNA 水平的分析, 证实外源目的基因在 PBMCs 中整合并表达. 将转导的 PBMCs 与 SMMC-7721 共培养, 由于 Millicell 底部为孔径 $0.4 \mu\text{m}$ 的滤膜, 细胞不能通过该滤膜, 而其分泌物可以相互透过^[29], 因此, 用 MTT 法可检测 PBMCs/PST 表达产物对肝癌细胞的杀伤作用, 结果表明, 分泌型抗肝癌单链单链免疫毒素对体外培养的肝癌细胞系 SMMC-7721 具有杀伤作用. 本实验结果表明, 转导的人 PBMCs 可以表达并分泌特异性的抗肝癌 sFv-TNF- 融合蛋白, 并且对体外培养的肝癌细胞系 SMMC-7721 具有杀伤活性. 因此, 采用人 PBMCs 作

为靶向细胞因子的表达细胞进行肝癌基因治疗是可行的.

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