

# 胰腺癌组织 ChAT, GAD65 和 PKC 酶活性的表达

杨竹林, 王群伟, 邓星辉, 李代强, 吕芳, 李永国

杨竹林, 王群伟, 邓星辉, 吕芳, 李永国, 中南大学湘雅二医院肝胆疾病研究室 湖南省长沙市 410011  
李代强, 中南大学湘雅二医院病理科 湖南省长沙市 410011  
杨竹林, 男, 1962-08-08 生, 湖南省涟源市人, 汉族. 1998 年湖南医科大学博士研究生毕业, 教授. 主要从事消化系统疾病的病理研究.  
项目负责人: 杨竹林, 410011, 湖南省长沙市, 中南大学湘雅二医院肝胆疾病研究室.  
收稿日期: 2002-12-24 接受日期: 2003-01-14

## Enzymatic activities of ChAT, GAD65 and PKC in pancreatic carcinoma tissues

Zhu-Lin Yang, Qun-Wei Wang, Xing-Hui Deng, Dai-Qiang Li, Fang Lü, Yong-Guo Li

Zhu-Lin Yang, Qun-Wei Wang, Xing-Hui Deng, Dai-Qiang Li, Fang Lü, Yong-Guo Li, Research Laboratory of Hepatobiliary Diseases, Xiangya Second Hospital of Zhongnan University, Changsha 410011, Hunan Province, China

Correspondence to: Zhu-Lin Yang, Department of Pathology, Xiangya Second Hospital of Zhongnan University, Changsha 410011, Hunan Province, China.

Received: 2002-12-24 Accepted: 2003-01-14

## Abstract

AIM: To study on the expression characteristics of ChAT, GAD65 and PKC enzymatic activities and their clinicopathological significance in the tissues of chronic pancreatitis and pancreatic carcinoma.

METHODS: The enzymatic activities of ChAT, GAD65 and PKC were detected by immunohistochemical method of avidin-biotin complex on formalin-fixed and routine paraffin-embedded sections of specimens of chronic pancreatitis ( $n=10$ ) and pancreatic carcinoma ( $n=47$ ).

RESULTS: The positive rate and the score of ChAT, GAD65 and PKC were significantly lower in 10 cases of chronic pancreatitis than that of pancreatic carcinoma (ChAT, 0% vs 48.9%,  $0.2 \pm 0.4$  vs  $2.2 \pm 1.4$ ; GAD65, 10.0% vs 55.3%,  $0.6 \pm 0.9$  vs  $2.2 \pm 1.2$ ; PKC, 10.0% vs 57.4%,  $0.6 \pm 0.9$  vs  $2.1 \pm 1.6$ ). The score of ChAT was significantly higher in well-differentiated adenocarcinoma than that of poorly-differentiated adenocarcinoma ( $P < 0.05$ ). The positive rate and the score of GAD65 or PKC were significantly lower (GAD65,  $P < 0.05$ ; PKC,  $P < 0.01$ ) in cases of well-differentiated adenocarcinoma than in cases of poorly-differentiated. No difference was found for the enzyme expressions and the clinicopathological characteristics among different sex, age, with or without metastasis of pancreatic carcinoma. A highly positive correlation was found between the scores of GAD65 and PKC in pancreatic carcinoma.

CONCLUSION: The expression of enzymatic activities of ChAT, GAD65 or PKC might be related to the carcinogenesis, progression and biological behaviors of pancreatic carcinoma. They might be important biological markers of

pancreatic carcinoma.

Yang ZL, Wang QW, Deng XH, Li DQ, Lü F, Li YG. Enzymatic activities of ChAT, GAD65 and PKC in pancreatic carcinoma tissues. *Shijie Huaren Xiaohua Zazhi* 2003;11(10):1554-1557

## 摘要

目的: 研究 ChAT, GAD65 和 PKC 酶活性在慢性胰腺炎和胰腺癌组织中表达特征及其临床病理意义。

方法: 胰腺癌( $n=47$ )和慢性胰腺炎( $n=10$ )手术切除标本经 40 g/L 中性甲醛固定后常规制作石蜡包埋切片, ChAT, GAD65 和 PKC 酶活性表达染色方法均为常规 ABC 免疫组化法。

结果: 胰腺癌 ChAT, GAD65 和 PKC 表达阳性率(48.9%, 55.3% 和 57.4%)及其评分( $2.2 \pm 1.4$ ,  $2.2 \pm 1.2$  和  $2.1 \pm 1.6$ )明显高于慢性胰腺炎阳性率(0%, 10.0% 和 10.0%)及其评分( $0.2 \pm 0.4$ ,  $0.6 \pm 0.9$  和  $0.6 \pm 0.9$ ), 均有显著或高度显著性差异( $P < 0.05$  或  $P < 0.01$ )。高分化腺癌 ChAT 评分值明显高于低分化腺癌( $P < 0.05$ ), 但阳性率之间无明显差异( $P > 0.05$ ); 高分化腺癌 GAD65, PKC 表达阳性率及其评分明显低于低分化腺癌, 均有显著或高度显著性差异( $P < 0.05$  或  $P < 0.01$ )。酶活性表达与胰腺癌患者性别、年龄、有无转移等临床特征均无明显关系。GAD65 评分与 PKC 评分存在高度密切关系( $r = 0.50$ ,  $P < 0.01$ )

结论: ChAT, GAD65 和 PKC 酶活性表达特征可能与胰腺癌发生发展及生物学行为有密切关系, 均为胰腺癌重要生物学标志物。

杨竹林, 王群伟, 邓星辉, 李代强, 吕芳, 李永国. 胰腺癌组织 ChAT, GAD65 和 PKC 酶活性的表达. *世界华人消化杂志* 2003;11(10):1554-1557

<http://www.wjgnet.com/1009-3079/11/1554.asp>

## 0 引言

胆碱乙酰转移酶(choline acetyltransferase, ChAT)在胆碱能神经元和神经纤维及细胞运动中起重要调节作用, 是胆碱能神经元特异性标志物<sup>[1-6]</sup>。谷氨酸脱羧酶(glutamate decarboxylase, GAD)有两种异构体(即 GAD65 和 GAD67), 二者与  $\gamma$ -氨基丁酸(GABA)能系统的功能代谢存在密切关系, 是 GABA 合成的关键酶<sup>[7-9]</sup>。蛋白激酶 C (protein kinase C, PKC) 在信号转换、细胞分化和生长过程中起重要作用<sup>[10-13]</sup>。新近发现 ChAT 表达活性与恶性肿瘤细胞发生及侵袭潜能有关<sup>[14-16]</sup>, GAD65 和 PKC 表达活性

与恶性肿瘤发生发展及生物学行为有较密切关系<sup>[17-30]</sup>. 国内外尚未见胰腺癌这方面研究的文献报道. 我们应用免疫组化方法研究胰腺癌和慢性胰腺炎组织中 ChAT, GAD65 和 PKC 酶活性表达及其生物学意义.

### 1 材料和方法

1.1 材料 收集湘雅医院和湘雅二医院1996-01/2001-12 手术切除胰腺癌标本 47 例 (术前均未进行化疗和放疗), 男 35 例. 女 12 例, 年龄 21-73 (平均 50 ± 12)岁. 均来源于胰腺导管上皮, 包括高分化腺癌 19 例, 中分化腺癌 8 例, 低分化腺癌 17 例和黏液腺癌 3 例. 临床和(或)病理证实发生胰腺外转移(区域淋巴结或胰腺外组织器官)36 例 (76.6 %). 另收集慢性胰腺炎手术切除标本 10 例, 男 6 例, 女 4 例, 年龄 35-65(平均 42 ± 7 岁). 上述标本经 40 g/L 中性甲醛固定后, 常规制作石蜡包埋连续切片, 切片厚 4 μm. 兔抗人 ChAT, GAD65 多克隆抗体, 鼠抗人 PKC 单克隆抗体, 生物素标记羊抗兔 IgG, 生物素标记羊抗鼠 IgG, ABC 试剂, 以及 DAB-HCL 显色试剂盒均购自武汉博士德公司.

1.2 方法 ChAT, GAD65 和 PKC 表达染色方法均为常规 ABC 免疫组化法(具体染色步骤略), 镜下观察 5 个高倍镜视野染色结果, 细胞质内含棕黄色颗粒者为 ChAT 和 GAD65 阳性细胞, 细胞膜含棕黄色颗粒者为 PKC 阳性细胞; 根据免疫组化着色强度评分(0 分, 无; 1 分, 弱; 2 分, 中; 3 分, 强)和阳性细胞率(0 分, 小于 5 %; 1 分, 5-10 %; 2 分, 11-20 %; 3 分, 21-50 %;

4 分, 大于 50 %)之和为该病例评分值, 且将评分值 0-2 分定为阴性表达病例(-), 大于 2 分定为阳性表达病例(+). 以 0.05 mol/L PBS 液(PH7.4)代替一抗作为染色阴性对照或替代对照, 以多次替代对照有效的乳腺癌切片作为染色阳性对照.

统计学处理 将所得数据输入 SPSS10.0 软件包进行  $\chi^2$  检验, t 检验及 Fisher's 精确概率法, 检验水准  $\alpha = 0.05$  和  $\alpha = 0.01$ .

### 2 结果

ChAT 和 GAD65 免疫组化反应产物主要定位于细胞质, 偶见细胞核着色(图 1, 2). PKC 免疫组化反应产物主要定位于细胞膜, 部分病例定位于细胞膜和细胞质(图 3). 三者 in 癌组织中分布呈较明显异质性, 同一切片中癌组织内阳性细胞着色强度和阳性细胞率可有较明显不同. 慢性胰腺炎导管上皮三者阳性率及其评分明显低于胰腺癌, 均有显著或高度显著性差异( $P < 0.05$  或  $P < 0.01$ , 表 1). 高分化腺癌 ChAT 表达阳性率及其评分明显高于中、低分化腺癌, 与低分化腺癌相比其评分有显著性差异( $P < 0.05$ ). 低分化腺癌 GAD65、PKC 表达阳性率及其评分明显高于中、高分化腺癌, 与高分化腺癌比较均有显著或高度显著性差异( $P < 0.05$  或  $P < 0.01$ ). 三者表达阳性率与性别、年龄及有无转移均无明显关系( $P > 0.05$ , 表 2). ChAT 表达评分与 GAD65、PKC 表达评分无明显相关性( $P > 0.05$ ), 但 GAD65 与 PKC 表达评分呈密切正相关( $r = 0.50$ ,  $P < 0.01$ ).

表 1 胰腺癌组织 ChAT, GAD65 和 PKC 酶活性表达

分组	n	阳性率(%)			评分值( $\bar{x} \pm s$ )		
		ChAT	GAD65	PKC	ChAT	GAD65	PKC
胰腺炎	10	0.0	10.0	10.0	0.2 ± 0.4	0.6 ± 0.9	0.6 ± 0.9
胰腺癌	47	48.9 <sup>b</sup>	55.3 <sup>a</sup>	57.4 <sup>a</sup>	2.2 ± 1.4 <sup>b</sup>	2.2 ± 1.2 <sup>b</sup>	2.1 ± 1.6 <sup>a</sup>

<sup>a</sup> $P < 0.05$ , <sup>b</sup> $P < 0.01$  vs 胰腺炎.

表 2 胰腺癌临床病理特征与 ChAT, GAD65 和 PKC 表达的关系

	n	阳性率(%)			评分值( $\bar{x} \pm s$ )		
		ChAT	GAD65	PKC	ChAT	GAD65	PKC
男	35	48.6	51.4	57.1	2.3 ± 1.5	2.1 ± 1.3	2.1 ± 1.5
女	12	50.0	66.7	58.3	2.0 ± 1.1	2.6 ± 0.9	2.1 ± 1.7
年龄小于 45	15	40.0	60.0	66.7	2.2 ± 0.9	2.5 ± 1.0	2.4 ± 1.5
大于或等于 45	32	53.1	53.1	53.1	2.2 ± 1.6	2.1 ± 1.3	2.0 ± 1.6
高分化腺癌	19	63.1	42.1	47.3	2.7 ± 1.4	1.8 ± 1.4	2.0 ± 1.5
中分化腺癌	8	37.5	50.0	37.5	1.9 ± 1.5	2.2 ± 0.9 <sup>b</sup>	1.2 ± 1.5
低分化腺癌	17	35.3	76.5 <sup>a</sup>	82.4 <sup>a</sup>	1.8 ± 1.2 <sup>a</sup>	2.8 ± 0.9 <sup>b</sup>	2.8 ± 1.5 <sup>c</sup>
黏液腺癌	3	66.7	33.3	33.3	2.0 ± 1.7	1.3 ± 1.5	1.3 ± 1.5 <sup>d</sup>
转移 无	11	45.5	54.5	63.6	2.5 ± 1.6	2.1 ± 1.6	2.1 ± 1.6
有	36	50.0	55.6	55.6	2.1 ± 1.3	2.3 ± 1.1	2.1 ± 1.6

<sup>a</sup> $P < 0.05$ , <sup>b</sup> $P < 0.01$  vs 高分化腺癌, <sup>c</sup> $P < 0.01$ , <sup>d</sup> $P < 0.01$  vs 低分化腺癌.

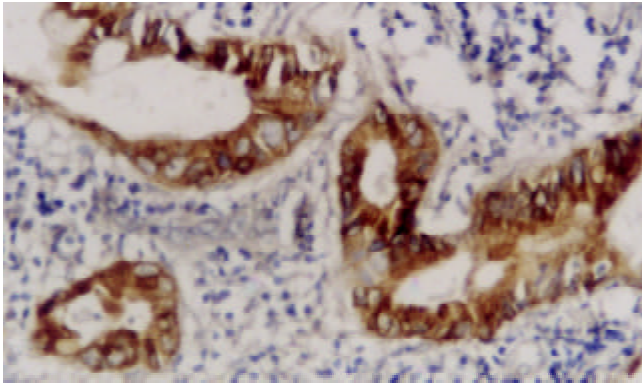


图1 胰腺高分化腺癌 ChAT 酶活性阳性表达 ABC 法  $\times 100$ .

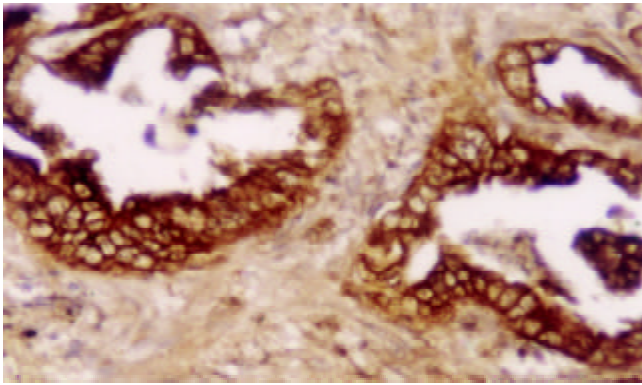


图2 胰腺中分化腺癌 GAD65 酶活性阳性表达 ABC 法  $\times 100$ .

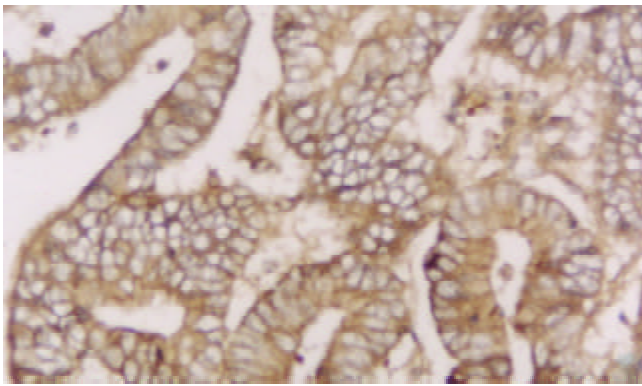


图3 胰腺高分化腺癌 PKC 酶活性阳性表达 ABC 法  $\times 100$ .

### 3 讨论

ChAT 可能与肿瘤发生发展及生物学行为有密切关系, 人类原发和转移性黑色素瘤存在 ChAT 和蝇蕈碱乙酰胆碱受体的表达, 而正常皮肤黑色素细胞则缺乏, 人类黑色素瘤细胞株(SK-mel28)中, ChAT 通过其受体调节瘤细胞收缩和运动, 认为与瘤细胞侵袭潜能有密切关系. 人类小细胞肺癌、畸胎瘤和神经母细胞瘤中 ChAT 能促进癌细胞生长和增强其侵袭和发生转移的能力<sup>[14-16]</sup>. ChAT 在其他恶性肿瘤中是否表达尚未文献报道. 我们发现慢性胰腺炎导管上皮均呈 ChAT 阴性表达, 胰腺癌阳性表达率 48.9%, 且高分化腺癌 ChAT 评分明显高于低分化腺癌, 但与胰腺癌是否转移无明显关系. 提示部分胰腺癌表达 ChAT, 说明部分上皮性恶性肿瘤象黑色

素瘤一样存在 ChAT, ChAT 可能通过其受体作用调节肿瘤细胞生长、分化、运动及其生物学行为, 其确切作用机制有待深入研究.

GAD65 为  $\gamma$ -氨基丁酸(GABA)合成的关键酶, 而 GABA 源性系统在调节细胞生长发育等方面起重要作用, 应用 GABA 受体拮抗剂处理后可抑制多种细胞的生长发育<sup>[7-9]</sup>. 近年研究发现恶性肿瘤(如结肠癌、乳腺癌、胃癌等) GAD65 表达水平明显高于其正常组织和良性病变, 认为 GAD65 表达与恶性肿瘤发生、生物学行为等有密切关系<sup>[17-20]</sup>. 我们发现慢性胰腺炎导管上皮 GAD65 表达阳性率及其评分明显低于胰腺癌, 高分化腺癌阳性率及其评分明显低于低分化腺癌, 但与是否转移无明显关系. 其结果与国外文献报道较一致, 说明 GAD65 表达特征与胰腺癌发生发展及生物学行为存在密切关系, 为胰腺癌重要生物学标志物之一.

PKC 是一大类结构功能不同的丝氨酸/苏氨酸激酶家族, 至少有 12 种同工酶亚型. PKC 广泛分布于哺乳动物组织细胞中, 具有多种生理功能, 是一系列细胞级联信号途径的关键环节<sup>[10-13]</sup>. 近年研究发现 PKC 表达在恶性肿瘤生物学行为及肿瘤细胞多药耐药性形成方面有重要作用, 恶性肿瘤 PKC 表达水平明显高于正常组织及其良性病变, 多药耐药胃癌细胞株 PKC 表达水平明显高于药物敏感细胞株<sup>[21-30]</sup>. 本结果显示慢性胰腺炎中导管上皮 PKC 表达阳性率及其评分明显低于低分化腺癌, 但与有无转移无明显关系. 说明 PKC 表达与胰腺癌发生及其恶性程度有密切关系, 检测 PKC 表达水平对指导临床胰腺癌化疗或放疗可能具有重要临床应用价值. 其机制可能与 PKC 本身生物学作用有关.

### 4 参考文献

- 1 Fujii T, Tajima S, Yamada S, Watanabe Y, Sato KZ, Matsui M, Misawa H, Kasahara T, Kawashima K. Constitutive expression of mRNA for the same choline acetyltransferase as that in the nervous system, an acetylcholine-synthesizing enzyme, in human leukemic T-cell lines. *Neurosci Lett* 1999;259:71-74
- 2 Sailer M, Oppitz M, Drews U. Induction of cellular contractions in the human melanoma cell line SK-mel 28 after muscarinic cholinergic stimulation. *Anat Embryol* 2000;201:27-37
- 3 Pahud G, Bontron S, Eder-Colli L. Modulation of choline acetyltransferase synthesis by okadaic acid, a phosphatase inhibitor, and KN-62, a CaM kinase inhibitor, in NS-20Y neuroblastoma. *Neurochem Int* 2001;38:75-82
- 4 Bugnard E, Sors P, Roulet E, Bloc A, Loctin F, Dunant Y. Morphological changes related to reconstituted acetylcholine release in a release-deficient cell line. *Neuroscience* 1999;94:329-338
- 5 Fujii T, Ushiyama N, Hosonuma K, Suenaga A, Kawashima K. Effects of human antithymocyte globulin on acetylcholine synthesis, its release and choline acetyltransferase transcription in a human leukemic T-cell line. *J Neuroimmunol* 2002;128:1-8
- 6 Miettinen RA, Kalesnykas G, Koivisto EH. Estimation of the total number of cholinergic neurons containing estrogen receptor-alpha in the rat basal forebrain. *J Histochem Cytochem* 2002;50:891-902
- 7 Matsuba T, Yano M, Abiru N, Takino H, Akazawa S, Nagataki S, Yasukawa K. Expression of recombinant human glutamic acid decarboxylase (GAD) in myeloma cells and enzyme-

- linked immunosorbent assay (ELISA) for autoantibodies to GAD. *J Biochem* 1997;121:20-24
- 8 Pinal CS, Cortessis V, Tobin AJ. Multiple elements regulate GAD65 transcription. *Dev Neurosci* 1997;19:465-475
- 9 Salazar P, del Carmen Sanchez-Soto M, Hiriart M, Tapia R. Biochemical characteristics of the gamma-aminobutyric acid system in the insulinoma cell lines HIT-T15, RIN-m5F, beta TC3, and comparison with rat brain. *Arch Med Res* 2001;32:419-428
- 10 Kohn EA, Yoo CJ, Eastman A. The protein kinase C inhibitor Go6976 is a potent inhibitor of DNA damage-induced S and G2 cell cycle checkpoints. *Cancer Res* 2003;63:31-35
- 11 Darley RL, Pearn L, Omidvar N, Sweeney M, Fisher J, Phillips S, Hoy T, Burnett AK. Protein kinase C mediates mutant N-Ras-induced developmental abnormalities in normal human erythroid cells. *Blood* 2002;100:4185-4192
- 12 Gopalakrishna R, Gundimedda U. Antioxidant regulation of protein kinase C in cancer prevention. *J Nutr* 2002;132:3819S-3828S
- 13 Wang SI, Mukhtar H. Gene expression profile in human prostate LNCaP cancer cells by (-) epigallocatechin-3-gallate. *Cancer Lett* 2002;182:43-51
- 14 Song P, Sekhon HS, Jia Y, Keller JA, Blusztajn JK, Mark GP, Spindel ER. Acetylcholine is synthesized by and acts as an autocrine growth factor for small cell lung carcinoma. *Cancer Res* 2003;63:214-221
- 15 Fisher HS, Berti I, Schatz DS, Humpel C, Saria A. Retinoic acid treatment enhances the acetylcholine contents in the human teratocarcinoma cell line NTera-2. *Regul Pept* 2000;96:59-63
- 16 Nagai A, Suzuki Y, Baek SY, Lee KS, Lee MC, McLarnon JG, Kim SU. Generation and characterization of human hybrid neurons produced between embryonic CNS neurons and neuroblastoma cells. *Neurobiol Dis* 2002;11:184-198
- 17 Matuszek M, Jesipowicz M, Kleinrok Z. GABA content and GAD activity in gastric cancer. *Med Sci Monit* 2001;7:377-381
- 18 Opolski A, Mazurkiewicz M, Wietrzyk J, Kleinrok Z, Radzikowski C. The role of GABA-ergic system in human mammary gland pathology and in growth of transplantable murine mammary cancer. *J Exp Clin Cancer Res* 2000;19:383-390
- 19 Mazurkiewicz M, Opolski A, Wietrzyk J, Radzikowski C, Kleinrok Z. GABA level and GAD activity in human and mouse normal and neoplastic mammary gland. *J Exp Clin Cancer Res* 1999;18:247-253
- 20 Kleinrok Z, Matuszek M, Jesipowicz J, Matuszek B, Opolski A, Radzikowski C. GABA content and GAD activity in colon tumors taken from patients with colon cancer or from xenografted human colon cancer cells growing as s.c. tumors in athymic nu/nu mice. *J Physiol Pharmacol* 1998;49:303-310
- 21 Clark AS, West KA, Blumberg PM, Dennis PA. Altered protein kinase C (PKC) isoform in non-small cell lung cancer cells: PKC delta promotes cellular survival and chemotherapeutic resistance. *Cancer Res* 2003;63:780-786
- 22 Yoshikawa N, Nakamura K, Yamaguchi Y, Kagota S, Shinozuka K, Kunitomo M. Effect of PKC412, a selective inhibitor of protein kinase C, on lung metastasis in mice injected with B16 melanoma cells. *Life Sci* 2003;72:1377-1387
- 23 Orlandi L, Binda M, Folini M, Bearzatto A, Villa R, Daidone MG, Zaffaroni N. Ribozyme-mediated inhibition of PKC alpha sensitizes androgen-independent human prostate cancer cells to cisplatin-induced apoptosis. *Prostate* 2003;54:133-143
- 24 Biswas DK, Martin KJ, McAlister C, Cruz AP, Graner E, Dai SC, Pardee AB. Apoptosis caused by chemotherapeutic inhibition of nuclear factor-kappaB activation. *Cancer Res* 2003;63:290-295
- 25 Metzger E, Muller JM, Ferrari S, Buettner R, Schule R. A novel inducible transactivation domain in the androgen receptor: implications for PRK in prostate cancer. *EMBO J* 2003;22:270-280
- 26 Chen Y, Wu Q, Song SY, Su WJ. Activation of JNK by TPA promotes apoptosis via PKC pathway in gastric cancer cells. *World J Gastroenterol* 2002;8:1014-1018
- 27 Simard M, Zhang W, Hinton DR, Chen TC, Weiss MH, Su YZ, Gopalakrishna R, Law RE, Couldwell WT. Tamoxifen-induced growth arrest and apoptosis in pituitary tumor cells in vitro via a protein kinase C-independent pathway. *Cancer Lett* 2002;185:131-138
- 28 Volm M, Sauerbrey A, Zintl F, Koomagi R, Efferth T. Protein expression profile of newly diagnosed acute lymphoblastic leukemia in children developing relapses. *Oncol Rep* 2002;9:965-969
- 29 Shimizu T, Taira N, Senou M, Takeda K. Involvement of diverse protein kinase C isoforms in the differentiation of ML-1 human myeloblastic leukemia cells induced by the vitamin D3 analogue KH1060 and the phorbol ester TPA. *Cancer Lett* 2002;186:67-74
- 30 韩英, 时永全, 李玲. 蛋白激酶 C 同工酶 PKC-A 及 PKC-BI 在胃癌及其耐药细胞中的表达和功能. *中华肿瘤杂志* 2001;23:103-106