

PD98059对乙醛刺激的大鼠肝星状细胞增生的影响

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Effects of PD98059 on proliferation of rat cultured hepatic stellate cells stimulated by acetaldehyde

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Abstract

AIM: To study the effects of PD98059, the specific blocking agent of MEK₁, on the proliferation of hepatic stellate cells and expression of Proliferating Cell Nuclear Antigen in rat hepatic stellate cells (HSC).

METHODS: HSC stimulated by acetaldehyde were cultured. The cell growth was evaluated by MTT colorimetric assay. Proliferating cell nuclear antigen (PCNA) was examined by immunocytochemical staining.

RESULTS: PD98059 of 20 μmol/L had an inhibitory effect on proliferation of HSC ($P < 0.05$, 0.109 ± 0.020 vs 0.146 ± 0.030), which was more obvious when cells exposed to PD98059 at 50 and 100 μmol/L ($P < 0.05$, 0.081 ± 0.010 , 0.056 ± 0.020 vs 0.146 ± 0.030), and the expression of PCNA also showed a descending tendency with the increase of PD98059 concentration ($P < 0.05$, 0.62 ± 0.09 , 0.47 ± 0.04 , 0.34 ± 0.04 vs 0.74 ± 0.05)

CONCLUSION: PD98059 inhibits proliferation of HSC and expression of PCNA, which is correlated with the decreased activity of PCNA.

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摘要

目的: 探讨特异性 MEK1 阻断剂 PD98059 对乙醛刺激的肝星状细胞 (hepatic stellate cell, HSC) 增生及其细胞增生核抗原表达的影响。

方法: 用 PD98059 对乙醛刺激的 HSC 进行处理, 分别以 MTT 比色、免疫细胞化学法检测细胞增生及其细胞增生核抗原表达。

结果: PD98059 在 20 μmol/L 时即对 HSC 增生出现抑制作用 ($P < 0.05$, C 组 0.109 ± 0.020 vs B 组 0.146 ± 0.030), 50、100 μmol/L 时抑制作用逐渐增强 ($P < 0.05$, D、E 组 0.081 ± 0.010 、 0.056 ± 0.020 vs B 组 0.146 ± 0.030); HSC 中 PCNA 表达也随 PD98059 剂量增加而减弱 ($P < 0.05$, C、D、E 组 0.62 ± 0.09 、 0.47 ± 0.04 、 0.34 ± 0.04 vs B 组 0.74 ± 0.05)

结论: PD98059 对 HSC 增生及细胞增生核抗原表达具有抑制作用, 提示 Erk 信号传导通路是调控肝星状细胞增生的重要通道。

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0 引言

肝星状细胞(HSC)是形成肝纤维化的主要细胞, HSC的活化与增生是肝纤维化进展的中心环节^[1-5]. 乙醛是激活HSC导致酒精性肝纤维化的关键分子^[6,7]. 细胞外信号调节激酶(extracellular signal-regulated kinase, Erk)信号通路参与调控细胞增生与分化, 是多种信号交汇点或共同通路^[8-14], 但在乙醛刺激的HSC增生中的作用报道较少. 我们用不同剂量的特异性 MEK1 阻断剂 PD98059 阻断 Erk 活性后, 观察乙醛刺激的 HSC 增生及其细胞增生核抗原表达的变化, 探讨 Erk 信号通路调控乙醛刺激的 HSC 增生的分子机制, 为酒精性肝纤维化的防治提供理论依据.

1 材料和方法

1.1 材料 PD98059(NEB 公司); 增生细胞核抗原(PCNA)免疫组化法检测试剂盒(北京中山公司); 小牛血清及 DMEM/IMDM 培养基(美国 Gibco 公司); CFSC 大鼠肝星状细胞株由第三军医大学李小安博士惠赠.

1.2 方法 细胞用含 150 ml/L 小牛血清的 DMEM/IMDM (1:1) 培养基, 另添加适量的 HEPES、抗生素(1×10^5 单位/L 青霉素和 100 mg/L 链霉素)至 37°C 、50 ml/L CO_2 混合气体的孵箱中培养, 细胞换液时间为 2-3 d, 传代时间为 3-5 d, 传代前用 2.5 g/L 胰酶消化. 实验分为: 空白对照组(A 组)加无血清的 DMEM/IMDM(1:1)培养液; 乙醛对照组(B 组): 无血清培养液中加乙醛 200 $\mu\text{mol/L}$; 实验组 1(C 组): 在 B 组基础上培养液中加入 PD98059 (浓度为 20 $\mu\text{mol/L}$); 实验组 2 (D 组): 在 B 组基础上培养液中加入 PD98059 (浓度为 50 $\mu\text{mol/L}$); 实验组 3 (E 组): 在 B 组基础上培养液中加入 PD98059 (浓度为 100 $\mu\text{mol/L}$). 取大鼠 HSC, 调整浓度至 $1 \times 10^8/\text{L}$ 接种于 96 孔细胞培养板中, 每孔 200 μL 细胞悬液, 细胞生长至 80% 以上融合度时, 4 ml/L 血清培养基同步化处理 24 h, 实验组经 PD98059 预处理 1 h 后, 加乙醛 200 $\mu\text{mol/L}$ (终浓度), CO_2 培养箱中继续孵育 24 h 后(乙醛每 12 h 补充), 每孔加 5 g/L 的 MTT 20 μL , 反应 4 h, 用快速翻板法去除培养液, 加 DMSO 200 μL , 30 min 后用酶标仪(E-Liza Mat-3000)双波长测定其 A 值, 测定波长为 570 nm, 参考波长为 630 nm, 酶标仪所示 A 值为 A_{570} 减去 A_{630} , 以消除非特异性光吸收效应. 免疫组化: 以 $5 \times 10^8/\text{L}$ 接种 12 孔培养板, 每孔 1 ml, 作细胞爬片. 细胞生长至 80% 以上融合度行同步化处理, 按上述分组法加药继续培养 24 h. 将培养板孔中的盖玻片取出, PBS 洗涤两次, 用 40 g/L 多聚甲醛固定 10 min, 按 SP 试剂盒操作方法进行免疫细胞化学 PCNA 检测, DAB 显色, 同时用 PBS 代替一抗作阴性对照. 结果判定: 胞核呈棕黄色为阳性, 并对染色后的细胞片进行显微镜下观察, 摄片, IDA-2000 软件分析平均灰密度, 相对定量 PCNA 表达的强度.

统计学处理 所有指标均用($\bar{x} \pm s$)表示, 采用 SPSS11.0 软件分析. 成组设计多样本均数的比较用单因素方差分析.

2 结果

2.1 MTT 结果 加入乙醛后明显刺激 HSC 增生($P < 0.05$, B 组 0.146 ± 0.030 vs A 组 0.030 ± 0.010); 随着 PD98059 用量增加 C (0.109 ± 0.020)、D(0.081 ± 0.010)、E (0.056 ± 0.020) 3 组细胞增生均受到抑制($P < 0.05$, vs B 组), 呈明显的剂量效应关系, 但 E 组细胞增生仍高于 A 组, 表明 100 $\mu\text{mol/L}$ PD98059 仍不能完全抑制乙醛刺激的 HSC 增生.

2.2 PCNA 免疫组织化学染色 乙醛对照组 HSC 胞核着色最深, 呈棕黄色(图 1), 实验组随着 PD98059 用量的增加胞核着色逐渐减弱, PD98059 100 $\mu\text{mol/L}$ (完全抑制 Erk 活性剂量)胞核呈淡黄色(图 2). 经病理分析软件分析显示, 乙醛组 PCNA 平均灰密度较对照组明显增高($P < 0.05$, B 组 0.74 ± 0.05 vs A 组 0.24 ± 0.03); 实验组随着 PD98059 用量的增加 HSC 内 PCNA 平均灰密度较乙醛组明显降低($P < 0.05$), E 组 PCNA 平均灰密度

(0.34 ± 0.04)仍高于 A 组($P < 0.05$), 结果表明 PD98059 具有抑制作用, 但 100 $\mu\text{mol/L}$ PD98059 仍不能完全抑制乙醛刺激的 HSC 内 PCNA 表达.

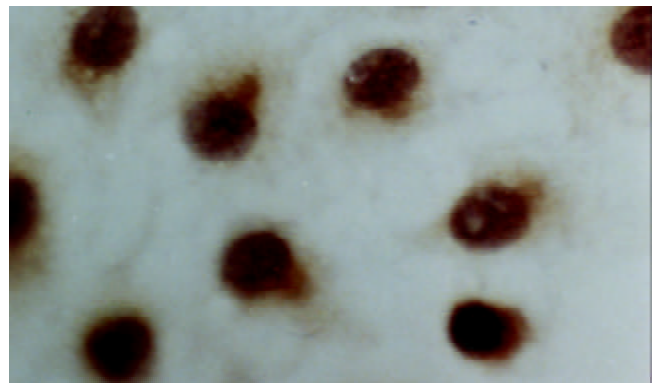


图 1 乙醛组 HSC 免疫细胞化学染色胞核着色呈棕黄色(×400 SP 法).

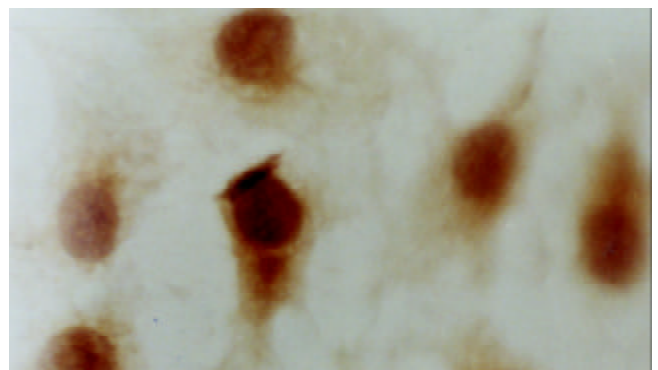


图 2 100 $\mu\text{mol/L}$ PD98059 予处理乙醛刺激的 HSC 后胞核着色呈淡黄色(×400 SP 法).

3 讨论

肝纤维化的发生机制较复杂^[15], 目前关于 ERK 信号传导通路的激活与肝纤维化发生机制的研究甚少. 文献表明: Erk 信号传导通路参与调控 PDGF 等刺激的 HSC 增生^[16-19]. 乙醛作为有效刺激信号是激活 HSC 导致酒精性肝纤维化发生的关键分子, 但 ERK 信号通路在乙醛刺激的 HSC 增生中的意义报道较少, 我们采用 MTT 实验观察 PD98059 对乙醛刺激的 HSC 增生的影响, 结果表明阻断 Erk 信号通路后 HSC 增生明显抑制, 且有剂量依赖关系, 提示 Erk 信号通路是调控乙醛刺激的 HSC 增生的重要通道, 同时 100 $\mu\text{mol/L}$ PD98059 (完全抑制 Erk 活性剂量)仍不能完全抑制 HSC 增生, 提示 Erk 通路并非调控乙醛刺激的 HSC 增生的唯一通道.

增生细胞核抗原(proliferating cell nuclear antigen, PCNA), 又称细胞周期蛋白, 是真核细胞 DNA 合成时所必需的一种 36 KD 的酸性核蛋白^[20], 他在细胞周期调控方面发挥着重要作用^[21-26], 其表达亦可反应细胞增生的活跃程度^[27-30], 目前 Erk 信号通路调控乙醛刺激的 HSC 增生的分子机制尚不明了, 我们用不同浓度 PD98059(20, 50, 100 $\mu\text{mol/L}$)分别加入乙醛刺激的 HSC 内 24 h 后, 免疫细胞化学染色法检测发现,

PD98059 组与乙醛对照组相比, PCNA 表达逐渐减弱, 100 $\mu\text{mol/L}$ 的 PD98059 组 PCNA 表达仍弱阳性, 提示 PD98059 抑制乙醛刺激的 HSC 增生可能与一定程度上抑制细胞周期蛋白 PCNA 合成有关。

目前对 Erk 信号通路调控乙醛刺激的 HSC 增生的分子机制研究刚刚起步, 其确切机制尚需进一步研究。

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