

The Cysteinyl Leukotriene (CysLT) Pathway in Allergic Rhinitis

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ABSTRACT

The cysteinyl leukotrienes (CysLTs), leukotriene C₄(LTC₄), leukotriene D₄(LTD₄) and leukotriene E₄(LTE₄) are released in response to specific allergen in nasal secretions from patients with active allergic rhinitis. The symptoms and inflammation of allergic rhinitis can be induced by inhalation of CysLTs. Inflammatory cells from patients with allergic rhinitis express both the synthetic and signaling proteins for the CysLT pathway. CysLTs activate cell migration, in particular eosinophils, endothelial or epithelial cell adhesion and release of cytokines and other oxidative inflammatory mediators. Cytokines may also activate the release of CysLTs from eosinophils and other myeloid cells and also enhance the expression of the CysLT₁ receptor creating an inflammatory amplification cycle. Systemic CysLT₁ receptor antagonists can reduce the inflammation and symptoms of both allergic rhinitis and asthma.

KEY WORDS

allergic rhinitis, CysLT receptors, CysLTs, cytokines, inflammation

INTRODUCTION

Activation of the leukotriene synthetic pathway occurs in inflammatory or allergic cells following stimulation of intracellular calcium release by specific plasma membrane activating molecules such as bacterial cell wall antigens on neutrophils or IgE on mast cells.^{1,2} Calcium release induces the translocation of five-lipoxygenase (5-LO) and cytosolic phospholipase A₂ to the nuclear membrane where arachidonic acid is released and presented to 5-LO by five-lipoxygenase-activating protein (FLAP).³ FLAP is one of the membrane associated proteins of eicosanoid and glutathione metabolism (MAPEG) family of which there are six members that are involved in either detoxification or eicosanoid synthesis. It is possible that FLAP evolved from an early evolutionary detoxification role to a specific cofactor for the production of leukotriene A₄(LTA₄). In the presence of FLAP, 5-LO efficiently catalyzes a two-step reaction to produce LTA₄ that can be converted by perinuclear-bound LTC₄ synthase to LTC₄ or by cytosolic LTA₄ hydrolase to leukotriene B₄(LTB₄).³ LTB₄ is a direct activator of neutrophils, monocytes and macrophages via activation of the BLT1 and BLT2 high affinity G protein-coupled receptors (GPCRs).⁴ LTC₄ is ex-

ported from cells via the MRP anion pump and converted in blood by γ -glutamyltransferase to LTD₄ and then by dipeptidase action to LTE₄. LTC₄, LTD₄ and LTE₄ are collectively called the CysLTs and they are the components of slow reacting substance of anaphylaxis (SRS-A). The CysLTs act on monocytes, macrophages, eosinophils and mast cells and on lung and intestinal smooth muscle cells via specific activation of the CysLT₁ and/or CysLT₂ GPCRs.⁵⁻⁹ Several cellular processes may be activated by CysLT receptor stimulation but intracellular calcium release appears to be the primary signaling pathway.⁹

The CysLT synthetic and signaling proteins are expressed on inflammatory cells, including mast cells, eosinophils and monocytes/macrophages in nasal secretions from patients with active allergic rhinitis and in mucosa from patients undergoing turbinectomies for obstructed nasal passages.^{10,11} The three committed proteins in CysLT synthesis, namely 5-LO, FLAP and LTC₄ synthase, are expressed on 40% of mast cells, 70% of eosinophils and 70% of monocytes/macrophages in nasal secretions.¹⁰ Both CysLT receptors are expressed at approximately the same ratios as the synthetic proteins on these inflammatory cells. The CysLT₁ receptor and 5-LO and FLAP (but not the CysLT₂ receptor or LTC₄ synthase) are ex-

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Received 18 February 2004.
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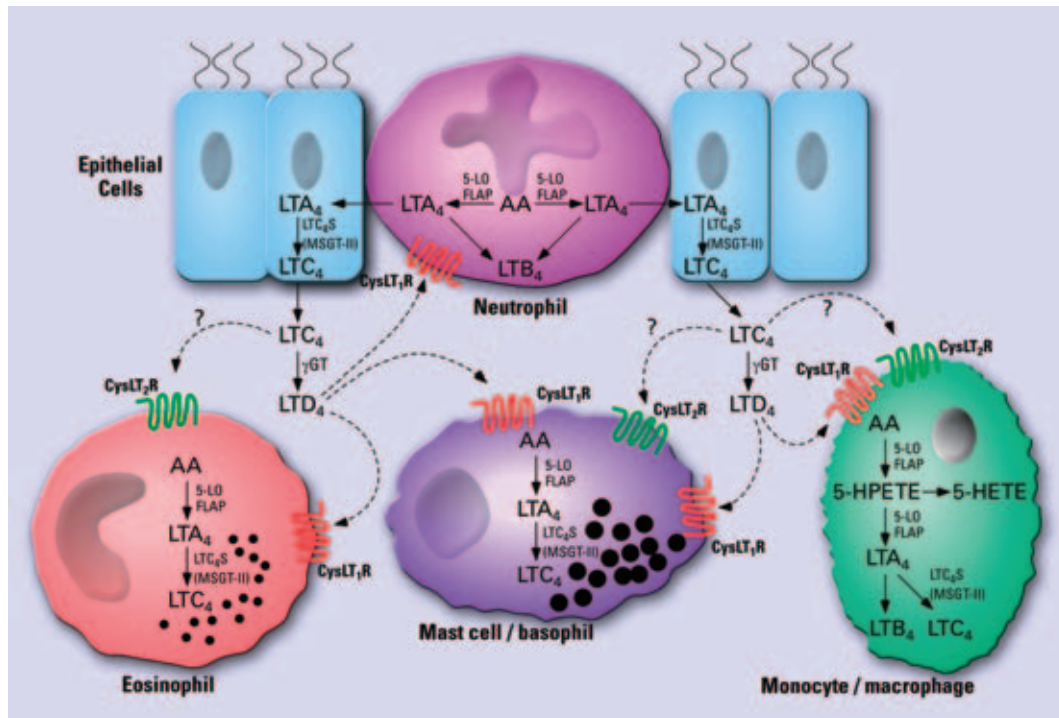


Fig. 1 Potential CysLT signaling pathways in inflammatory cells in allergic rhinitis. Reprinted with permission from Figueroa DJ *et al.* Expression of cysteinyl leukotriene synthetic and signaling proteins in inflammatory cells in active seasonal allergic rhinitis. *Clin. Exp. Allergy* 2003; **33**: 1380-1388.

pressed on 40–60% of neutrophils recovered in nasal secretions from patients suffering from active allergic rhinitis.¹⁰ The latter finding was not expected since previously negligible CysLT₁ receptor expression had been observed on peripheral blood neutrophils from normal individuals.¹² It is possible that CysLT₁ receptor expression is enhanced during trafficking from the blood into the nasal passages of patients suffering from allergic rhinitis.¹⁰ Since 80–90% of eosinophils in nasal secretions expressed both CysLT₁ and CysLT₂ receptors it is possible that these receptors could form heterodimers as well as homodimers or oligomers, but none of these structures has been confirmed. The CysLT₁ receptor is also present on neutrophils, eosinophils, mast cells and vascular cells, but not on epithelial cells or submucosal glands, in nasal mucosa samples from congested patients requiring turbinectomies.¹¹ A cartoon of the expression of the CysLT pathway proteins and potential interaction of CysLTs with their receptors in allergic nasal tissues is depicted in Figure 1.

In allergic subjects, CysLTs (but not LTB₄) are elevated following specific ragweed allergen challenge in a ragweed pollen dose-dependent fashion.¹³⁻¹⁶ The elevations in CysLTs correlate temporally with the severity of the symptoms of allergic rhinitis.¹⁵ Intranasal challenge with LTD₄ results in nasal secretions (rhinorrhea) peaking at 5 minutes af-

ter LTD₄ challenge and returning to control levels within 20 minutes.¹⁶ In addition, intranasal LTC₄ challenge in 5 patients increased nasal airway resistance in a dose-dependent fashion.¹³

The severity of nasal blockage in allergic rhinitis is proportional to the nasal eosinophil concentration.¹⁷ Activated eosinophils produce high amounts of CysLTs which can act in an autocrine or paracrine manner to activate eosinophil chemotaxis, migration, degranulation and adhesion to β_2 integrins on endothelial or epithelial cells.¹⁸⁻²¹ LTD₄ promotes chemotaxis of eosinophils with the rank order of potency of agonists that reflects a CysLT₁ receptor activation profile, namely LTD₄ > LTC₄ > LTE₄.¹⁸ LTD₄ enhanced eosinophil-derived neurotoxin release induced by IL-5.¹⁹ In addition, LTD₄ up-regulates expression of adhesion molecules such as Mac-1 and can enhance the survival of eosinophils *in vitro* to a similar degree as treatment with GM-CSF.²⁰⁻²² CysLT₁ receptor antagonists reverse the ability of CysLTs to prolong eosinophil life *in vitro*.²²

CysLT treatment can increase the expression of TH₂ inflammatory cytokines such as IL-4 and IL-5 from eosinophils and mast cells.^{23,24} In reverse, TH₂ cytokines can increase CysLT synthesis and CysLT₁ receptor expression creating a cycle of inflammatory mediator amplification.²⁵⁻²⁷ IL-13 and IL-4 have been shown to increase CysLT₁ receptor expression in hu-

man monocytes and macrophages.²⁶ IL-13 increases CysLT₁ receptor expression in human fetal lung fibroblasts lung fibroblasts and IL-13 has a synergistic effect with LTC₄ on eotaxin production from these cells.²⁷ This eotaxin production was blocked by the CysLT₁ receptor antagonists montelukast and pranlukast.²⁷ These CysLT₁ receptor antagonists can also inhibit IL-5 expression in ragweed and mite-stimulated peripheral blood mononuclear cells.^{28,29} Interestingly montelukast also inhibited the synthesis of CysLTs in peripheral blood mononuclear cells suggesting that antagonism of the receptor decreased the number of CysLT producing cells.²⁹

The CysLT-stimulated production of non TH₂ cytokines such as MIP-1 β and TNF- α may be blocked by treatment with CysLT₁ receptor antagonists.²³ Other inflammatory markers such as NO have been shown to be elevated by LTC₄ treatment of human polymorphonuclear leukocytes *in vitro*.³⁰ Both montelukast and pranlukast have been shown to reduce exhaled NO in asthmatics.^{31,32} Superoxide radicals have the potential to initiate oxidative damage in cells and LTD₄ has been shown to increase these unstable oxidation products in human eosinophils *in vitro*.³³ The increase in superoxide radicals was blocked by treatment with the CysLT₁ receptor antagonist pranlukast.³³

As described above CysLT₁ receptor antagonists block effects of CysLT activation of eosinophils, mast cells and neutrophils. Pranlukast has been shown to be effective in the prevention of LTD₄-induced nasal secretion, nasal airway resistance and nasal eosinophil infiltration in a guinea pig model of allergic rhinitis.³⁴ Pranlukast has also been shown cause a significant decrease in eosinophil infiltration and cytokine production in human nasal mucosa of patients with perennial allergic rhinitis.³⁵

Montelukast at 10 mg once daily dosing has shown efficacy against daytime and nighttime symptoms and improved quality of life scores in multi-center, randomized, double-blind, placebo-controlled seasonal allergic rhinitis trials held in spring and fall.³⁶⁻³⁸ In the spring trial montelukast was also shown to decrease peripheral blood eosinophil counts indicating both nasal and systemic anti-inflammatory effects in this trial.³⁷

The pathophysiology of rhinitis has many common features with that of asthma and 20–50% of patients with rhinitis also have asthma, whereas 80% or more of patients with asthma have concomitant rhinitis.³⁹ This has led to the hypothesis that systemic drugs such as CysLT₁ receptor antagonists may treat both upper and lower airway disease treating the inflammatory ‘one airway’ symptoms for patients with both asthma and allergic rhinitis.⁴⁰⁻⁴³ Urinary LTE₄ has been regarded as an index of systemic CysLT synthesis and severe nasal blockage was shown to be associated with increased urinary LTE₄ concentration in

patients with seasonal allergic rhinitis.⁴⁴ It will be of interest to measure both activated leukocyte CysLT synthesis and urinary CysLT production from patients with both asthma and seasonal allergic rhinitis during both active and inactive allergen seasons. More sophisticated analysis of LT products or LT pathway polymorphisms may allow us in the future to better define the population who would benefit most by systemic CysLT antagonism or inhibition.

ACKNOWLEDGEMENTS

The author gratefully acknowledges the skilled scientific help of David Figueroa, Helen Galczenski, and Dr. Marie Gleason (Merck, West Point, PA, USA), the talented artistic skills of Kevin Clark (Merck Frosst, Montreal, Canada) and the superb administrative and personal support of Sandy Camburn.

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