Pollen and Inorganic Characteristics of Bulgarian Unifloral Honeys

JULIANA ATANASSOVA¹, LILYANA YURUKOVA² and MARIA LAZAROVA²

¹Department of Botany, Faculty of Biology, University of Sofia "St. Kliment Ohridski", Sofia, Bulgaria; ²Institute of Biodiversity and Ecosystem Research, Bulgarian Academy of Sciences, Sofia, Bulgaria

Abstract

ATANASSOVA J., YURUKOVA L., LAZAROVA M. (2012): **Pollen and inorganic characteristics of Bulgarian unifloral honeys.** Czech J. Food Sci., **30**: 520–526.

The melissopalynological characteristics, three main physicochemical parameters (water content, pH, and electrical conductivity), and 19 macro- and microelements contents of 15 honey types from throughout Bulgaria that were collected from 2006 to 2009 were evaluated. The main honeys studied came from *Robinia pseudoacacia* L., *Helianthus annuus* L., *Brassica* spp., *Tilia* spp., and *Vicia* spp. The botanical origins of unifloral honey samples were identified as Lotus spp., *Coriandrum sativum* L., *Daucus*-type, *Stachys*-type, *Salix* spp., *Prunus* spp., *Castanea sativa* Mill., *Paliurus spina-christi* Mill., *Sophora japonica*, and *Amorpha* spp. Based on the physicochemical parameters and elements contents, one sample with high a percentage of *Trifolium* spp. pollen was identified as honeydew honey.

Keywords: unifloral honey; melissopalynology; physical-chemical parameters; macro- and microelements

Unifloral honey is a type of honey predominantly produced from the nectar of a single plant species. Unifloral honeys have an important commercial value because they are regarded as more valuable, often offered for sale at higher prices than the multifloral honeys (PERSANO ODDO & BOGDANOV 2004). Botanical origin strongly influences the quality of honey and all its physical and chemical characteristics (PŘIDAL & VORLOVÁ 2002; Bogdanov 2009; Primorac et al. 2011). It can be difficult to differentiate between polyfloral and unifloral honeys because multiple factors influence the honey composition. The European Directive (European Commission 2002) specifies some compositional limits for blossom and honeydew honeys, but it does not establish any legal criteria for unifloral honeys (Persano Oddo & BOGDANOV 2004). The classification of unifloral

honeys needs to include microscopic, physicochemical, and elemental analyses (BOGDANOV *et al.* 2004; PERSANO ODDO & PIRO 2004), however, the limits vary from country to country and may lead to some difficulties in the international trade in unifloral honey.

Melissopalynological investigations are rare in Bulgaria, and so are physicochemical and elemental analyses of honey (IVANOV & CHERVENAKOVA 1984; LAZAROVA & BOZILOVA 2001; ATANASSO-VA & KONDOVA 2004; ATANASSOVA *et al.* 2004, 2009). The goal of the present study was to define the melissopalynological and physicochemical parameters – water content, pH, and electrical conductivity (EC) as well as the elemental characteristics of Bulgarian unifloral honeys, and to compare them with the data obtained from other European regions.

Partly supported by the National Science Fund, Ministry of Education, Youth and Science, Bulgaria, Project No. N TK-B-1611.

MATERIAL AND METHODS

More than 200 honey samples were obtained, most of them directly from beekeepers all over Bulgaria, having been collected during the years of 2006–2009. Only 36 samples were identified as unifloral honeys according to their melissopalynological characteristics (Table 1) and they are the focus of the present paper. The samples were examined for their physicochemical and elemental properties. Organoleptic characteristics (colour and physical state) were described by sensorial analysis (Table 1).

Melissopalynolgical analysis. The method described by LOUVEAUX *et al.* (1978) was followed in the laboratory preparations and qualitative melissopalynological analysis. Along with the pollen identification, all honeydew elements detected (HDE) (algae and fungal elements) were counted. The frequency of the pollen types was expressed as percentage of the pollen sum (PN), which includes pollen grains only from nectar producing plants. To be considered unifloral, a honey sample should contain at least 45% of the corresponding pollen (Pd) (LOUVEAUX *et al.* 1978). The pollens of some plant species are known to be overrepresented in

the honey (e.g. Castanea sativa Mill.), while others are underrepresented (e.g. Robinia pseudoacacia L), therefore different percentage values mark the honey as unifloral (Louveaux et al. 1978; Persano Oddo & PIRO 2004). For the quantitative analysis, the method described by MOAR (1985) was followed. Tablets containing a known number of spores of Lycopodium spp. were added to the sample. The absolute pollen concentration in 1 g of honey was derived from the ratio of total pollen × *Lycopodium* spores added to the number of Lycopodium spores counted. The frequency classes of honeys were identified according to MAURIZIO (1939). The pollen concentration of the Pd was calculated in the same way (Table 1). The ratio HDE/PN was also calculated (LOUVEAUX et al. 1978). Pollen identification was made by light microscopy and compared to the reference collections and with BEUG (2004).

Physicochemical analysis. The routine physicochemical analysis included water content (honey refractometer Atago HHR-2N 12–30%; Atago Co., Ltd., Tokyo, Japan), EC (mS/cm, \pm 1%) in 20% solution at 20°C (MultiLine P3; WTW, Weilheim, Germany), and pH (20% solution, \pm 0.01, Jenway pH-meter; Bibby Scientific Ltd., Staffordshire, UK) (Table 2). The Harmonised Methods of the

Table 1. Organoleptic and melissopalynological characteristics of the honeys studied

Numbers	Dominant pollen type	Colour	Physical state	Pd (%)	Concentration of Pd	Frequency class
1-6	Robinia	water white–light yellowish	liquid	34.7-53.5	1 193 (mean of 6 samples)	I–II
7	Trifolium	brownish	liquid	52.9	1 381 (1)	II
8	Lotus	light yellowish	liquid	49.6	4 482 (1)	II
9-10	Vicia	water white	liquid	51.8-45.0	1 588 (mean of 2 samples)	II
11	Sophora	light yellowish	liquid	74.8	1 767 (1)	II
12	Amorpha	light yellowish	liquid	63.8	2 118 (1)	II
13–17	Tilia	light amber–dark amber	liquid	36.8-87.8	1 769 (mean of 5 samples)	II
18-23	Helianthus	bright yellow– dark yellow	fine crystallised– large granulated	45.1-78.9	2 473 (mean of 6 samples)	II
24-29	Brassica	milky white– creamy white	fine crystalised	86.5-93.4	5 043 (mean of 6 samples)	II
30	Prunus	water white	liquid	78.8	1 659 (1)	II
31	Paliurus	dark yellow	liquid	67.7	3 890 (1)	II
32	Stachys	light yellow	fine crystalised	48.8	4 926 (1)	II
33	Castanea	dark amber	liquid	97.0	10 104 (1)	III
34	Coriandrum	amber	liquid	44.8	3 661 (1)	II
35	Daucus	dark yellow	fine crystalised	46.2	1 111 (1)	II
36	Salix	amber	fine crystalised	54.2	1 328 (1)	II

International Honey Commission (2009) do not specify any method for the identification of elements in honey. About 10 g of material was treated with 15 ml nitric acid (9.67M) overnight. The wet-ashing was continued with heating in a water bath, followed by the addition of 2 ml hydrogen peroxide. This treatment was repeated until reaching full digestion. The filtrate (through filter paper Filpap KA 2; Filpap, Štětí, Czech Republic) was diluted with double-distilled water (0.06 μ S/cm) up to 25 ml. All solutions were stored in plastic flasks. Macroelements K, Ca, Mg, P, S, and microelements Al, As, Cd, Co, Cr, Cu, Fe, Mn, Na, Ni, Pb, Sr, V, and Zn were determined by atomic emission spectrometry with the inductively coupled plasma system (ICP-AES) of VARIAN VISTA-PRO (Tables 3a and 3b). The detection limits were 0.002 mg/l for Mn and Sr, 0.004 mg/l for Cd, Co, Cr, Cu, Ni, and Zn, 0.02 mg/l for As and V, 0.03 mg/l for Pb, 0.04 mg/l for Al and Fe, and 0.5 mg/l for Ca, K, Mg, Na, P, and S. Analytical precision was checked by replications and blanks and by stock standard solutions (1000 µg/l Merck) for the preparation of working aqueous solutions.

RESULTS AND DISCUSSION

According to the literature, over 100 plant species have been recorded as being used by bees to produce

unifloral honey in Europe (PERSANO ODDO & PIRO 2004). The data summarised in this study provide a good description of five Bulgarian unifloral honeys: black locust (R. pseudoacacia) – 6 samples, lime (Tilia spp.) – 5 samples, sunflower (Helianthus annuus L.) – 6 samples, rape (Brassica, most probably B. rapa L. ssp. oleifera, B. napus L. ssp. oleifera) -6 samples, and vetch (*Vicia* spp.) -2 samples. Further investigations are necessary to confirm the established characteristics of the other types of unifloral honeys represented by single samples only: coriander (Coriandrum sativum L.), willow (Salix spp.), Prunus spp. (including plums, cherries, peaches and apricots), chestnut (C. sativa), garland thorn (Paliurus spina-christi Mill.), Daucus-type (Apiaceae family), Stachys-type (Lamiaceae family), birdsfoot trefoil (Lotus spp.), japanese pagodatree (Sophora japonica L.), and amorpha (Amorpha spp.). One of the samples (7) with high percentage values of Trifolium spp. pollen was identified as honeydew honey.

Robinia honey is one of the most valued and purchased honeys on the Bulgarian market but it is difficult to identify. Six samples were identified as *Robinia* honey and were characterised by their light colour and liquid consistence, as were other Fabaceae honeys (*Lotus, Vicia, Sophora,* and *Amorpha*), with the exception of sample 7 (Table 1). The investigated *Robinia* honeys had the lowest concentration of the Pd and the lowest

Table 2. Main physicochemical parameters of the honeys studied

Number	Type of honey	Water content (%)	pН	Electrical conductivity (mS/cm)
1–6	Robinia (mean of 6 samples)	16.9	3.23	0.159
7	Honeydew (1)	17.5	4.28	0.961
8	Lotus (1)	15.5	3.22	0.338
9–10	Vicia (mean of 2 samples)	19.0	3.32	0.261
11	Sophora (1)	15.5	3.36	0.323
12	Amorpha (1)	19.0	3.20	0.200
13–17	<i>Tilia</i> (mean of 5 samples)	17.1	4.07	0.689
18-23	Helianthus (mean of 6 samples)	18.9	3.32	0.359
24-29	Brassica (mean of 6 samples)	19.7	3.33	0.181
30	Prunus (1)	15.2	3.19	0.185
31	Paliurus (1)	17.0	4.14	1.046
32	Stachys (1)	17.0	3.32	0.443
33	Castanea (1)	18.8	5.65	1.804
34	Coriandrum (1)	15.4	4.46	0.469
35	Daucus (1)	18.0	3.24	0.454
36	Salix (1)	20.0	3.61	0.399

total pollen concentration, belonging to class I and II (Table 1). Pollen in *Robinia* honey is usually underrepresented, because of the low pollen production (RICCIARDELLI D'ALBORE 1998). The recommendations of LOUVEAUX *et al.* (1978), and BSS (1990) were followed in this study, namely that 30% of *Robinia* pollen is considered sufficient to classify the honey as unifloral. *Vicia* honey and *Prunus* honey were also declared as *Robinia* honey by the beekeepers, probably because of its identical colour and physical state (Table 1). The mean values of the water content, pH, and EC were the lowest in this study (Table 2), coinciding with the results of PERSANO ODDO and PIRO (2004).

Three types of unifloral honey come from crop plants: sunflower, rape, and coriander. The rape honey was creamy white to milky white with fine crystals. The concentration of Brassica pollen was 5043/g on the average, higher than in the Helianthus honey (2473/g). Electrical conductivity of rape honeys was on the average 0.181 mS/cm, and of Helianthus honey on the average 0.359 mS/cm, which is close to the values cited by PERSANO ODDO and PIRO (2004). For Brassica honey, the maximum water content (19.7% on average) was established, followed by Helianthus honey (18.9% on average). Water content in blossom honey was found to be in the range of 15–20% (WHITE 1975; BOGDANOV 2009). The unifloral honey samples from different sites in Bulgaria revealed a similar range.

The colour of *Tilia* honey ranged from light to dark-amber. The mean value of the pollen concentration (Pd) was 1767/g. We accepted 30% lime pollen as sufficient to classify the honey as unifloral (LOUVEAUX *et al.* 1978; BSS 1990). The mean values of pH and EC had maxima (Table 2) coinciding with the data of BOGDANOV (2009). In Switzerland, pH in honeys was on the average 4.5 ± 0.8 for blossom honey and 4.5 ± 0.26 for honey-dew honey (BOGDANOV & GFELLER 2006). The relatively low pH of honey is due to the presence of organic acids, which contribute to the honey flavour and stability against microbial spoilage. The values of pH above 4.5 were mentioned only for honeydew honey (BOGDANOV 2009).

Castanea sativa honey is specific for southwest areas of the country where small groves of sweet chestnut occur. One sample only (33) was identified as unifloral chestnut honey (declared by the beekeeper as honeydew honey). Many authors have noted that *Castanea* pollen is strongly overrepresented in honey, and requires at least 90%, with more than 10 000/g pollen concentration, for accepting this honey as unifloral. The high pollen concentration in sample 33 is the only one assigned to class III in this study (Table 1). The low ratio of HDE/PN (0.06) proved its nectar origin. The high EC (1.804 mS/cm) and pH (5.65%) coincide with the values given by PERSANO ODDO and PIRO (2004) and BOGDANOV (2009).

Sample 7 was declared by the beekeeper as honeydew honey, and its organoleptic characteristics corresponded to this type of honey in contrast to the palynological data (Table 1). The ratio HDE/PN was low (0.66). According to the high pH and EC (4.28% and 0.961 mS/cm, respectively), the elevated mineral contents of six elements (Mg, Mn, Na, P, Cu, Cr), and especially the high concentration of Mn, it could be assumed that sample 7 was honeydew honey. According to the European Honey Standard, the EC of the honeydew type is one of the most important characteristics (BOGDANOV et al. 2004), and should exceed 0.8 mS/cm. The average values of macro- and microelements in different unifloral honeys can be summarised as follows: The lime honeys were characterised on average by the highest contens of Ca, K, Mg, Mn, S, Sr, and Zn. The minimum average values of Ca, Fe, Mg, Mn, P, S, and Zn were found in Robinia honeys. The lowest concentrations of Al and K were found in the Brassica honeys (Table 3). Metals Ni and Mn varied by more than two orders of magnitude in different honey samples, followed by Cu (over 45 times). The heavy metals Cd, Co, and the microelement V were in all cases below the detection limits. The elements Zn, K, Mg, and Fe varied 25-30 times, Al, P, S, and Ca 6-12 times. The maximum values of the biogenic macroelements K and S were recorded in chestnut honey. According to the literature (BOGDANOV et al. 2008), the content of Ni in honey was up to 0.051 mg/100 g. The Czech, Slovak, and Polish honeys had higher Ni levels than the honeys originating from other parts of the world (LACHMAN et al. 2007). Maximum of Ni found in this study was in sample 8 (1 mg/1 kg), probably because of the high motor traffic in the area of the Plovdiv town. The heavy metals Pb and Cd and the toxic elements Cr and As could reflect the presence of environmental contamination or pharmacological treatment of bees, or incorrect procedures used for the honey preservation (PISANI et al. 2008). In Lithuanian honey, some heavy metals showed a wide range of values: Pb 2.9–22.1 µg/kg, Cd 4.1–14.6 µg/kg, Cu 119.6-342.9 µg/kg, Zn 514.0-5639.0 µg/kg, lead-

Number	Honey type	Ca	Κ	Mg	Р	S I	Na Al	Fe	Mn	Zn
1-6	Robinia	32	126	6.0	24	12 8	.11 0.80	0.83	0.11	0.22
7	Honeydew	92	1121	97	124	41 1	6.3 0.47	1.73	12.7	0.47
8	Lotus	62	223	11	43	22 7	.99 0.56	4.37	0.10	0.56
9–10	Vicia	33	196	10	68	20 1	2.2 0.41	1.33	0.12	0.41
11	Sophora	78	264	15	71	32 9	.62 0.69	3.04	0.34	1.17
12	Amorpha	22	103	4.8	30	13 7	.22 0.24	0.58	0.06	0.08
1317	Tilia	77	792	21	49	24 7	.50 0.39	1.62	2.45	1.04
18-23	Helianthus	71	247	14	41	20 7	.58 0.36	1.93	0.36	0.61
24-29	Brassica	46	105	11	28	19 8	.49 0.36	1.01	0.17	0.25
30	Prunus	33	146	6.2	26	12 7	.50 0.37	0.35	0.10	0.17
31	Paliurus	62	1198	17	67	51 1	1.8 0.75	1.58	0.97	0.83
32	Stachys	57	403	20	71	32 8	.43 0.41	0.65	1.71	0.44
33	Castanea	66	1628	16	32	20 9	.55 0.64	0.59	3.73	0.20
34	Coriandrum	44	564	7.1	31	23 1	4.2 0.49	1.33	0.15	0.27
35	Daucus	110	339	15	41	25 8	.63 0.35	1.33	0.15	0.23
36	Salix	37	464	13	50	20 1	1.0 1.58	2.17	0.26	0.25
		As	Cd	Со	Cr	Cu	Ni	Pb	Sr	V
1-6	Robinia	< 0.1–0.16	< 0.01	< 0.01	< 0.01-0.01	< 0.01-0.1	5 < 0.01-0.08	< 0.08-0.15	0.15	< 0.05
7	Honeydew	0.320	< 0.01	< 0.01	0.020	0.45	0.14	< 0.08	0.39	< 0.05
8	Lotus	< 0.1	< 0.01	< 0.01	< 0.01	< 0.01	1.00	< 0.08	0.17	< 0.05
9–10	Vicia	< 0.1	< 0.01	< 0.01	< 0.01	< 0.01-0.4	3 0.05	< 0.08	0.12	< 0.05
11	Sophora	0.112	< 0.01	< 0.01	0.013	0.07	0.06	0.31	0.23	< 0.05
12	Amorpha	< 0.1	< 0.01	< 0.01	0.012	< 0.01	0.14	< 0.08	0.11	< 0.05
13-17	Tilia	< 0.1	< 0.01	< 0.01	< 0.01-0.01	0.12	< 0.01-0.92	< 0.08-0.19	0.33	< 0.05
18-23	Helianthus	< 0.1–0.17	< 0.01	< 0.01	< 0.01-0.01	< 0.01-0.0	7 < 0.01-0.98	< 0.08	0.21	< 0.05
24-29	Brassica	< 0.1–0.40	< 0.01	< 0.01	< 0.01-0.01	< 0.01-0.0	2 < 0.01-0.04	< 0.08	0.15	< 0.05
30	Prunus	0.220	< 0.01	< 0.01	< 0.01	< 0.01	0.57	< 0.08	0.13	< 0.05
31	Paliurus	< 0.1	< 0.01	< 0.01	< 0.01	0.07	< 0.01	< 0.08	0.27	< 0.05
32	Stachys	< 0.1	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.08	0.19	< 0.05
33	Castanea	< 0.1	< 0.01	< 0.01	< 0.01	0.09	< 0.01	< 0.08	0.40	< 0.05
34	Coriandrum	< 0.1	< 0.01	< 0.01	< 0.01	0.05	< 0.01	< 0.08	0.18	< 0.05
35	Daucus	< 0.1	< 0.01	< 0.01	< 0.01	0.08	< 0.01	< 0.08	0.34	< 0.05
36	Salix	0.268	< 0.01	< 0.01	< 0.01	0.23	< 0.01	< 0.08	0.13	< 0.05

Table 3. Concentrations of macro-and microelements in the honeys studied (mg/kg fresh weight)

ing to the conclusion that the maximum values were found in the urban or industrial areas, and that the overall content of these microelements in Lithuanian honey is lower than in the honeys of other EU countries (STANIŠKIENĖ *et al.* 2006). In comparison, the maximum concentrations found in this Bulgarian study were slightly higher for Cu (0.45 mg/kg) in the honeydew honey, and almost three times lower for Zn (1.861 mg/kg) in the *Tilia* honeys. The most pronounced difference in the trace elements contents was that between honeydew and blossom honeys (IVANOV & CHERVENA-KOVA 1984; FELLER-DEMALSY *et al.* 1989; SEVLIMLI *et al.* 1992). The differences in the contents of metals between the individual monofloral honey samples could be related to different compositions of organic compounds and their concentrations in the pollen. Some constituents such as phenolic compounds and flavonoids contained in the pollen could form complexes with metals (LACHMAN *et al.* 2010). This can explain higher levels of Al, As, Cu e.g. in *Salix* type honey.

The ranking of averages of all the analysed macroelements, heavy metals, and toxic elements in the *Robinia* honey was K > Ca > P > S > Na > Mg > Fe > Al > Zn > Sr > Mn, Ni, Cu, Cr, As, Pb, V, Co, Cd. In the *Tilia* honey, the ranking was: K > Ca > P > S > Mg > Na > Mn > Fe > Zn > Al >Sr > Cu; in the studied *Helianthus* honeys and the Brassica honeys, the order was similar to that of the *Robinia* honey, except with Mn occupying the last place. Potassium concentration, the highest one in various honey samples, was in accordance with the standards set by the Food and Agriculture Organization of the United Nations. No hygienic norms have been set for heavy metals and toxic elements in honey in Bulgaria. According to the data published by the European countries and some Maximum Admitted Levels, or Maximum Residue Limits (Čelechovská & Vorlová 2001; MATEI et al. 2004; BRATU & GEORGESCU 2005), however, the contents of heavy metals and toxic elements in the studied honeys from Bulgaria were low and did not present any health hazard

CONCLUSIONS

Unifloral honeys are rare in Bulgaria, because it is a common practice of the small honey producers to harvest honey only two times during the apicultural period. The botanical origin of honeys declared by the producers often did not correspond to our results of melissopalynological and physicochemical analyses.

The present study showed that the most common unifloral honey types in different regions of Bulgaria during 2006–2009 were black locust, lime, sunflower, and rape honeys. The ranking of average pollen concentrations for the honeys studied was *Brassica* > *Helianthus* > *Tilia* > *Robinia*. The highest pollen density was established for chestnut honey. The lime honey was found to have the highest average pH and EC and the maximum contents of Ca, K, Mg, Mn, S, Sr, and Zn. The highest EC (1.804 mS/cm) and pH (5.65) were observed in a single honey sample of chestnut. Based on the physico-chemical parameters and elements contents, one of the samples with a high percentage (59%) of *Trifolium* spp. pollen was identified as honeydew honey.

The present study showed low contents of heavy metals and toxic elements in all the honeys studied. The strongly varying contents of macro- and microelements could not be related only to different botanical origins of the honeys, but it also reflects different geographical origins as well as the environmental contamination of the localities in the respective geographical regions.

Acknowledgements. The authors are grateful to Dr. P. ZHELEV for collecting chestnut honey, to the staff of the Analytical Laboratory at the Institute of Biodiversity and Ecosystem Research, BAS for the laboratory work, and to Prof. PETRA MUDIE for improving the English text.

References

- ATANASSOVA J., KONDOVA V. (2004): Pollen and chemicalphysical analysis of unifloral honey from different regions of Bulgaria. Phytologia Balcanica, **10**: 45–50.
- ATANASSOVA J., BOZILOVA E., TODOROVA S. (2004): Pollen analysis of honey from the region of three villages in Western Bulgaria. Phytologia Balcanica, **10**: 247–252.
- ATANASSOVA J., YURUKOVA L., LAZAROVA M. (2009): Palynological, physical and chemical data of honey from the Kazanlak region (central Bulgaria). Phytologia Balcanica, 15: 107–114.
- BEUG H-J. (2004): Leitfaden der Pollenbestimmung fur Mitteleuropa und angrenzende Gebiete. Verlag Dr. Friedrich Pfeil, München.
- BOGDANOV S. (2009): The Book of Honey. Bee Product Science. Available at www.bee-hexagon.net
- BOGDANOV S., GFELLER M. (2006): Classification of honeydew and blossom honeys by discriminant analysis. Agroscope Liebefeld Posieux, **500**: 3–7.
- BOGDANOV S., JURENDIC T., SIEBER R., GALLMANN P. (2008): Honey for nutrition and health: a review. American Journal of the College of Nutrition, **27**: 677–689.
- BOGDANOV S., RUOFF K., PERSANO ODDO L. (2004). Physicochemical methods for the characterisation of unifloral honeys: a review. Apidologie, **35**: 4–17.
- BRATU I., GEORGESCU C. (2005): Chemical contamination of bee honey-identifying sensor of the environment pollution. Journal of Central Europaean Agriculture, 6: 95–98.
- BSS (1990): Bulgarian State Standard 2673-89. Bee Honey. Committee of Quality with the Bulgarian Council of Ministers.

- ČELECHOVSKÁ O., VORLOVÁ L. (2001): Groups of honey – physicochemical properties and heavy metals. Acta Veterinaria Brno, 70: 91–95.
- Codex Alimentarius (2001). Alinorm 41/0: Revised standard for honey., Alinorm, 1: 19–26.
- European Commission Council Directive 2001/110/EC (2002). Official Journal of the European Communities , L10: 47–52.

FELLER-DEMALSY M., VINCENT B., BEAULIEU F. (1989): Mineral content and geographical origin of Canadian honeys. Apidologie, **20**: 77–91.

Harmonised Methods of the International Honey Commission (2009). Available at http://www.bee-hexagon. net/en/network.htm

IVANOV T., CHERVENAKOVA Y. (1984): Content of some macro-, oligo- and microelements in bee honey, royal jelly and pollen. Journal of Animal Science, **21**: 65–69.

LACHMAN J., KOLIHOVÁ D., MIHOLOVÁ D., KOŠATA J., TITĚRA D., KULT K. (2007): Analysis of minority honey components: possible use for the evaluation of honey quality. Food Chemistry, **101**: 973–979.

LACHMAN J., HEJTMÁNKOVÁ A., SÝKORA J., KARBAN J., ORSÁK M., RYGEROVÁ B. (2010): Content of major phenolic and flavonoid antioxidants in selected Czech honey. Czech Journal of Food Sciences, 28: 412–426

- LAZAROVA M., BOZILOVA E. (2001): Pollen and chemical analysis of honey from different floral regions of Bulgaria, Phytologia Balcanica, 7: 101–112.
- LOUVEAUX J., MAURIZIO A., VORWOHL G. (1978): Methods of melissopalynology. Bee World, **59**: 139–157.

MATEI N., BIRGHILA S., DOBRINAS S., CAPOTA P. (2004): Determination of C vitamin and some essential trace elements (Ni, Mn, Fe, Cr) in bee products. Acta Chimica Slovenica, **51**: 169–175. MAURIZIO A. (1939): Untersuchungen zur quantitativen Pollenanalyse des Honing. Mitteilungen aus dem Gebiete der Lebensmittel Untersuchung und Hygiene, **30**: 27–72

MOAR N. (1985): Pollen analysis of New Zealand honey. New Zealand Journal of Agricultural Resarch, **28**: 39–70.

PERSANO ODDO L., BOGDANOV S. (2004): Determination of honey botanical origin: problems and issues. Apidologie, **35**: 2–3

PERSANO ODDO L., PIRO R. (2004): Main European unifloral honeys: descriptive sheets. Apidologie, **35**: 38–81.

PISANI A., PROTANO G., RICCOBONO F. (2008): Minor and trace elements in different honey types produced in Siena County (Italy). Food Chemistry, **107**: 1553–1560.

PRIMORAC L., FLANJAK I., KENJERIĆ D., BUBALO D., TOPOLNJAK Z. (2011): Specific rotation and carbohydrate profile of Croatian unifloral honeys. Czech Journal of Food Sciences, **29**: 515–519.

PŘIDAL A., VORLOVÁ L. (2002): Honey and its physical parameters. Czech Journal of Animal Science, 47: 439– 444.

RICCIARDELLI D'ALBORE G. (1998): Mediterranean Melissopalynology. Universita degli studi di Perugia, Faculta di Agraria, Perugia.

SEVLIMLI H., BAYULGEN N., VARINIOGLU A. (1992): Determination of trace elements in honey by INAA in Turkey. Journal of Radioanalytical and Nuclear Chemistry, **165**: 319–325.

STANIŠKIENĖ B., MATUSEVIČIUS P., BUDRECKIENĖ R. (2006): Honey as an indicator of environmental pollution. Environmental Research, Engineering and Management, 36: 53–58.

WHITE J.W. (1975): Composition of Honey. A Comprehensive Survey. Heinemann Press, London: 157–206.

> Received for publication February 2, 2012 Accepted after corrections April 17, 2012

Corresponding author

JULIANA ATANASSOVA, Assoc. Prof., University of Sofia "St. Kl. Ohridski", Faculty of Biology, Department of Botany, 8 Dragan Tzankov Blvd., Sofia -1164, Bulgaria

tel. + 359 281 673 52, e-mail: atanassova_juliana@abv.bg