The Association between Sporadic Alzheimer's Disease and the Human ABCA1 and APOE Gene Polymorphisms in Iranian Population

HR Khorram Khorshid^{1*}, E Gozalpour¹, K Kamali², M Ohadi¹, M Karimloo³, MH Shahhosseiny⁴

¹Genetic Research Centre, University of Social Welfare and Rehabilitation Sciences, Tehran, Iran, ²Reproductive Biotechnology Research Centre, Avicenna Research Institute (ACECR), Tehran, Iran, ³Epidemiology and Biostatistics Department, University of Social Welfare and Rehabilitation Sciences, Tehran, Iran, ⁴Microbiology Department, Islamic Azad University, Shahr-e-Qods Branch, Tehran, Iran

Abstract

Background: Apolipoprotein E (APOE), which its ε4 allele has been reported as a risk factor in late onset Alzheimer's disease (AD), is the main cholesterol carrier in the brain. ATP-binding cassette transporter A1 (ABCA1) gene on chromosome 9, which has been known by genome-wide AD linkage study, has an important role in cellular cholesterol efflux. This study determines the association between sporadic AD and the human ABCA1 and APOE gene polymorphisms in Iranian population.

Methods: 154 AD cases and 162 control subjects from Iranian population were genotyped for APOE genotypes and ABCA1 polymorphism (R219K).

Results: The frequency of $\epsilon 2\epsilon 3$ genotype was higher in control subjects comparing AD patients but was not significant (13% versus 5.8%) and $\epsilon 3\epsilon 4$ genotype frequency was significantly higher in AD cases comparing with control subjects. APOE- $\epsilon 2$ allele frequency in cases was lower than control subjects but this difference was not significant (4.5% versus 8%). Individuals carrying $\epsilon 4$ allele, developed AD 6.5 times more than non-carriers (OR=6.52, 95%Cl=2.63-16.17). There was no significant association between ABCA1 polymorphism and AD.

Conclusion: Unlike other studies, R219K polymorphism was not dependent on gender and APOE-ɛ4 allele and there was no association between APOE and ABCA1 in AD patients compared to controls.

Keywords: Alzheimer's disease; Genetic association; Apolipoprotein E; Polymorphism; ATP-binding cassette transporter A1; Iran

Introduction

Alzheimer's disease (AD), which presents progressive cognitive defects such as memory loss, apraxia and personality changes, is the commonest cause of dementia in the mid and late ages.^{1,2} Two neuropathophysiological hallmarks of AD are intracellular neurofibrillary tangles and beta amyloid plaques in brain blood vessels. As hundreds genes have been known as the risk factors for late onset AD, the wellknown one is apolipoprotein E gene (APOE) which has been recognized as the most important risk factor in 65% of sporadic cases.³ Apolipoprotein E is the main part of very low density lipoproteins, intermediate density lipoproteins (IDL), chylomicrons and the main cholesterol carrier in the brain and its synthesis is independent in central nervous system (CNS) and lung. As APOE expression is stimulated by any CNS damages or diseases, it seems that APOE regulates cholesterol metabolism and distribution in the brain to repair and stabilize neurons' membrane and myelin.⁴⁻⁶ Several lines of evidence show that cholesterol metabolism and A β deposition are related to each other.

^{*}Correspondence: Hamid Reza Khorram Khorshid, MD, PhD, Genetic Research Centre, University of Social Welfare and Rehabilitation Sciences, Tehran, Iran. Tel/Fax: +98-21-22180138, e-mail: <u>hrkk1@uswr.ac.ir</u> Received: September 10, 2010 Accepted: November 1, 2010

Cholesterol reducing drugs such as statin decrease brain A β level and increase non-amyloidogenic α secretase cleavage of amyloid precursor protein and lead in reduced A β deposition.⁷⁻¹⁰ High cholesterol diet in transgenic animal models of AD leads in increased A β deposition in the brain.^{8,11,12} On the other hand, it was shown that the prevalence of AD is 60%-70% lower in cholesterol reducing drug consumers.^{1,13,14} Thus the risk of AD and A β deposition can be altered by factors associated with cholesterol level.

The brain has the highest cholesterol content of the body (20%) and because of blood brain barrier, the cholesterol homeostasis is independent from cholesterol level of plasma. Most of the brain cholesterol is immobilized in the myelin and the remaining is in neurons, glials and extracellular lipoproteins.⁶ Every day, 6-7 mg of excess cholesterol is converted to 24 S-hydroxycholesterol by 24 S-hydroxilase to transport out of the brain through blood brain barrier and the gene encoding 24 S-hydroxilase has found to be associated with the risk of AD.^{15,16} Cellular cholesterol is transported out of the cell by a mechanism called cholesterol efflux in which a membraneassociated protein, ATP-binding cassette transporter A1 (ABCA1), transports the cholesterol to high density lipoproteins (HDL).¹³ ABCA1 gene is located in 9q31.3 position which has been shown to be linked with AD by previous studies.^{17,18} Loss of ABCA1 function causes Tangier's disease which is characterized by absence of HDL, coronary artery disease and neuropathy. Like APOE, the expression of ABCA1 is regulated by RXR-LXR heterodimer and there are some evidences suggesting lack of APOE secretion from microglia, when ABCA1 is not expressed, so it seems that ABCA1 can influence APOE and cholesterol metabolism in the CNS.^{5,19}

In support for a link between AD and cholesterol metabolism, it has been supposed that ABCA1 polymorphisms may influence brain cholesterol homeostasis and risk of developing AD. Association of ABCA1 polymorphisms and AD has been studied in different population and there are some positive and some negative results.^{7,8,13,15,17,20,21} Raygani *et al.* showed that APOE- ϵ 4 allele was a risk factor in developing AD in Iranian population but the protective role for APOE- ϵ 2 against AD in this population was not statistically significant.²² This study determines the association between sporadic AD and the human ABCA1 and APOE gene polymorphisms in an Iranian population.

Materials and Methods

This case and control study involved 154 AD cases (mean age=78.55±7.80 years) and 162 control subjects (mean age=77.14±6.95 years) in which AD cases recruited using DSM IV criteria and control subjects were included if they were older than 65 vears old with no known neuropsychiatry disorders. The informed consent was signed by all of them or their legal guardians. The criterion for inclusion as a case was the diagnosis of AD diagnosed by the expert psychiatrist and lacking any neurologic or psychiatric disorders for the control group. Subjects were excluded if they had any family history of dementia or neurologic diseases. AD and control subjects were recruited from Alzheimer's society of Iran and Geriatric centers of Farzanegan, Mehrvarzan, Shayestegan, Kahrizak, Hasheminejhad and Rheumatism Center in Tehran, Iran from 2007 to 2008. The information regarding the age, sex, ethnicity, job and education were asked and recorded and finally 5 ml of peripheral blood sample was collected in tubes containing 200 µl of 0.5 M EDTA. Genomic DNA was extracted from peripheral blood leukocytes using salting-out method. APOE was genotyped by PCR-RFLP method which had been described by Wenham et al.²³ DNA was amplified by polymerase chain reaction (PCR) using forward primer: 5'-TCC AAG GAG CTG CAG GCG GCG CA-3'; and Reverse primer: 5'-ACA GAA TCC GCC CCG GCC TGG TAC ACT GCC A-3'. The 227 bp PCR products were digested by Hha I (10 U/µl, Fermentas) and loaded on a 12% polyacrylamide gel for electrophoresis; finally the gels were stained using silver staining method.

To genotype ABCA1 in AD cases and control subjects, a part of exon 7 of ABCA1 was amplified by polymerase chain reaction (PCR) using forward primer: 5'-CCT GTC ATT GTG CCT TGT G -3'; and reverse primer: 5'-GGA TTG GCT TCA GGA TGT C -3'. The 372 bp PCR product was digested by *Sty* I (10 U/µl, Fermentas) and loaded on an 8% polyacrylamide gel for electrophoresis; finally the gels were stained by silver staining method.

APOE and ABCA1 alleles and genotypes frequencies were analyzed through logistic regression, χ^2 or Fisher's Exact tests. Statistical significance was assumed when *p* value was less than 0.05. The statistical analysis and the odd ratios (OR) were determined using SPSS software (versian13, Chicago, IL, USA) and free online epidemiological software of Open Epi (2.2.1).

Results

Distribution of age, sex, jobs, educational level and genetic background was almost the same in both groups, so there was no need to use any method for adjustment of cases and controls (Table 1). The mean age and number of females were slightly higher in patients compared to control subjects. Our data showed that the highest frequency of AD was observed in housewives and the lowest was among farmers. People with academic education had the lowest frequency among patients and illiterate individuals had the most frequency. The samples were consisted of 5 Iranian genetic backgrounds in which Fars was the most common population.

The frequencies of APOE genotypes and alleles in AD cases and control subjects were shown in Table 2. The frequency of $\varepsilon 2\varepsilon 2$ genotype in control subjects was lower than that in AD cases but it was not significant (*p*=0.444). The distribution of $\varepsilon 2\varepsilon 3$ genotype was not significantly different in both groups (13% in controls versus 5.8% in AD, *p*=0.128) and OR was found to be 0.53 (95%CI=0.23-1.21). The genotype frequency of $\varepsilon 3\varepsilon 3$ was higher in control subjects compared with patients (Reference Group). The $\varepsilon 3\varepsilon 4$ genotype frequency in AD cases was significantly

Table 1: Comparison of mean age, sex, jobs, education levels and genetic backgrounds between AD cases and control subjects.

Parameter		AD patients (No.=154)	Control subjects (No.=162)	P value
Age		78.55±7.80 ^a	77.14±6.95	0.091
Sex (M/F) ^b		63/91	63/99	0.714
	Housewife	55.8%	56.2%	
Jobs	Own business	23.4%	21.0%	
	Worker	9.2%	8.6%	0.938
	Farmer	3.2%	3.1%	
	Employee	8.4%	11.1%	
	Illiterate	41.6%	43.2%	
Education	Primary school	29.2%	29.6%	
levels	Secondary school	16.2%	12.3%	0.427
	Diploma	11.1%	9.3%	
	Academic	1.9%	5.6%	
Genetic back-	Fars	61.0%	63.6%	
ground	Turk	25.3%	25.3%	
0	Kurd	3.9%	1.8%	0.490
	Lor	0.7%	2.5%	
	Gilak and Mazani	9.1%	6.8%	

^aMean±SD, ^bMale/Female

Table 2: The genotype and allele frequencies were compared between AD cases and control subjects
--

Genotype/Allele	Alzheimer No.=154	Control No.=162	P value	Odds Ratio
Genotype				
£3£3	69.5%	82.1%	Rf [*]	
ε2ε2	1.3%	0.6%	0.444	2.48 (0.22-27.8)
ε2ε3	5.8%	13%	0.128	0.53 (0.23-1.21)
ε2ε4	0.6%	0.6%	0.439	1.24 (0.08-20.1)
ε3ε4	20.8%	3.7%	0.001	6.52 (2.63-16.17)
ε4ε4	2%	0	0.182	undefined
Allele				
ε3	82.8%	90.1%	Rf [*]	
ε2	4.5%	8%	0.243	0.67 (0.34-1.32)
ε4	12.7%	1.9%	0.001	7.44 (3.1-17.9)

*Reference Group.

higher than that in control group (20.8% versus 3.7%, p=0.001). The distribution of $\epsilon 2\epsilon 4$ genotype was the same in both groups and different distribution of $\epsilon 4\epsilon 4$ genotype in the groups was not significant (2% versus 0, p=0.182).

The APOE- ε 4 allele frequency was significantly higher in AD cases compared with control subjects (12.7% versus 1.9%, *p*=0.001). Comparing allele frequency in APOE- ε 4 allele carriers with non-carriers, OR was found to be 6.52 (95%CI=2.63-16.17). The frequency of APOE- ε 3 allele in patients was lower than that in control group (Reference Group). Despite of higher APOE- ε 2 allele frequency in AD cases compared with control subjects, this difference was not statistically significant (*p*=0.243 and OR=0.67, 95%CI=0.34-1.32) (Table 2).

Table 3 shows APOE genotype and allele frequencies distributed by sex groups. $\epsilon 2\epsilon 3$ genotype frequency in control subjects was higher than AD subjects in men and women group (p>0.05). The genotype frequency of $\epsilon 3\epsilon 4$ in AD cases was higher than control subjects in both male and female groups but it was significant just in female group (p=0.001). The frequency of APOE- $\epsilon 4$ allele in patients was significantly higher than control subjects in both males and females with different OR [males: p=0.002, OR=8.3 (1.86-37); females: p=0.001, OR=5.59 (2.07-15.05)]. The genotypes and alleles of ABCA1 gene was compared in two groups of AD cases and controls. As it has been summarized in Table 4, the GG genotype frequency was not different between AD cases and controls (Reference Group). Comparing GA and AA genotype frequencies, there was no significant difference between AD cases and control subjects (GA: p=0.451 and AA: p=0.696). No significant difference was observed between allele frequencies in cases and controls (p=0.592). Examining data stratified by gender, neither female AD cases nor male AD cases showed significant genotype and allele frequencies compared with female and male controls.

Furthermore, stratification of data by $\varepsilon 4$ allele of APOE, which had been genotyped for the AD cases and control subjects in the previous study, did not change the results. ABCA1 genotypes and alleles of AD and control subjects were compared between $\varepsilon 4$ carriers and $\varepsilon 4$ non-carriers (Table 5). Distribution of GG, GA and AA genotypes were not significantly different between AD cases and control subjects of $\varepsilon 4$ carriers and non- $\varepsilon 4$ carriers.

Discussion

According to this study, APOE-ɛ4 allele is a risk

 Table 3: APOE genotype and allele frequencies distributed by sex.

Genotype/Allele	Female	Male	
ApoE genotypes			
ε3/ε3	Rf [*]	Rf [*]	
ε2/ε3	<i>P</i> =0.522, OR=0.663 (0.22-1.75)	<i>P</i> =0.104, OR=0.23 (0.05-1.13)	
ε3/ε4	P=0.001, OR=7.86 (2.58-23.9)	P=0.080, OR=4.7 (0.96-22.8)	
ε4/ε4	No data	P=0.319, OR= undefined	
APOE alleles			
ε3	Rf [*]	Rf [*]	
ε4	<i>P</i> =0.001, OR=5.59 (2.07-15.05)	<i>P</i> =0.002, OR=8.3 (1.86-37)	
ε2	<i>P</i> =0.157, OR=0.46 (0.17-1.19)	P=0.878, OR=0.8 (0.29-2.24)	

*Reference Group

		Total			Female			Male	
	AD	Control	P value	AD	Control	P value	AD	Control	P value
Genotype	(No.=154)	(No.=162)		(No.=91)	(No.=99)		(No.=63)	(No.=63)	
GG	33.1%	37%	Rf	9.6%	39.4%	Rf	23.8%	33.3%	Rf
GA	48.7%	45.1%	0.451	0.4%	40.4%	0.803	55.6%	52.4%	0.341
AA	18.2%	17.9%	0.696	16.4%	20.2%	0.487	0.6%	14.3%	0.197
Allele									
G	57.5%	59.6%	0.592	59.6%	61.5%	0.699	51.6%	59.5%	0.205
Α	42.5%	40.4%		40.4%	38.5%		48.4%	40.5%	

*Reference Group

		APOE ε4 carrier	s	APOE ε4 non-carriers		
Genotype Frequency	Controls (No.=7)	Patients (No.=36)	P value	Controls (No.=155)	Patients (No.=118)	<i>P</i> value
GG	42.9%	25%	Rf	36.8%	35.6%	Rf
GA	42.9%	58.3%	0.618**	45.2%	45.8%	0.866
AA	14.2%	16.7%	1**	18%	18.6%	0.854
Allele Frequency						
G	64.3%	54.2%	0 495	64.3%	54.2%	0.926
А	35.7%	45.8%	0.400	35.7%	45.8%	0.030

Table 5: ABCA1	genotypes and alleles distri	bution in APOE £4 carriers	s and APOE £4 non-carriers.

*Reference Group, **Fischer exact test p value

factor for developing late onset AD in Iranian population like many other populations.²⁴⁻²⁹ Although $\varepsilon 2\varepsilon 3$ genotype seems to play a protective role against AD but the protective role of APOE- $\varepsilon 2$ allele has not demonstrated in this study and it may be proved by a larger sample size.

The risk of developing AD in individuals with $\epsilon 2\epsilon 3$ genotype is about 0.53 (95%CI=0.23-1.21) compared with individuals without this genotype so ε2ε3 genotype seems to be protective against AD whereas protective role of $\varepsilon 2$ allele has not demonstrated in Iranian population yet. APOE-E4 allele carriers develops AD, 6.5 times more than non-carriers (6.52, 95%CI=2.63-16.17). This allele's risk seems different in males and females. Different OR for E4 allele in men and women indicates that risk of AD in male APOE-ɛ4 allele carriers (OR=8.421, CI=1.894-37.44) is higher than female carriers (OR=5.846, CI=2.173-15.73), so it seems that despite the agedependant and dosage-dependent manner of this allele which were investigated in Iranian population by Raygani et al.,²² it may act in a sex-dependent way as well. As three patients were observed with £4£4 genotype, it was not possible to assess the dosagedependent action of $\varepsilon 4$ allele in this study.

As the study groups were similar based on potential confounders (age, sex, genetic background, job and education), it can be assumed that the results are mainly unbiased. There was no reliable history or evidence for the time of AD onset, so we couldn't evaluate the effect of different genotypes or alleles on the age of onset in the AD subjects. In an autopsybased study, the frequency of ε 4 allele and ε 4 ε 4, ε 3 ε 4 genotypes were 40%, 16.5% and 43.2% in AD patients and 16%, 2.2 and 20.9% in the control group.³⁰ In a group of African Americans AD patients, a significantly increased risk of AD was associated with two ε 4 alleles or one ε 4 allele when compared to ε 3 ε 3 genotype.³¹ In our study, the frequencies of $\varepsilon 4$ allele and $\varepsilon 4 \varepsilon 4$, $\varepsilon 3 \varepsilon 4$ genotypes were lower than results of Raygani *et al.*,²² but the proportion of them was the same and the results of two studies in Iranian population are consistent. No significant association was found between $\varepsilon 2$ allele or related genotypes and AD but it sounds to work as protective factor for AD; however this finding, should be confirmed in further studies with a larger sample size .

In initial studies, it had been shown that ABCA1 expression can affect the level of APOE expression. Increased amyloid deposition in ABCA1 null mice and position of this gene on chromosome 9, which was linked to AD, were another evidence to confirm ABCA1 gene as a target gene for AD association studies.^{3,5} In 2003, Wollmer reported that A allele of ABCA1 causes 1.7 years delay at onset of AD and A allele carriers had 33% lower cholesterol in cerebrospinal fluid, than non-carriers of this allele.¹⁵ Significant increased frequency of A allele in control subjects compared with AD cases in Chinese population was reported by Wang et al. in 2007, while Sunder et al. reported that AD incidence is 1.5 times more frequent in A allele female carriers compared with female non-carriers in white American population^{8,13}

In this study, ABCA1 and its association with AD was studied in Iranian population for the first time. No association was observed between genotypes and alleles of ABCA1and AD. In 2006, Shibata *et al.* reported that ABCA1-A allele frequency was significantly higher in APOE- ε 4 carriers compared with ε 4 non-carriers.⁷ Therefore it was decided to investigate about this finding, but stratification by gender and APOE- ε 4 allele did not change the result and there was no association between genotypes and alleles of ABCA1 and the risk of AD among APOE- ε 4 carriers and non-carriers.

Acknowledgment

We express our sincere thanks and gratitude to all Alzheimer's and control persons or their families for their

References

- St George-Hyslop PH, Petit A. Molecular biology and genetics of Alzheimer's disease. *C R Biol* 2005; 328:119-30. [15770998] [doi:10.10 16/j.crvi.2004.10.013]
- 2 Avella AMB, Oostra BA, Heutink P. Chasing genes in Alzheimer's and Parkinson's disease. *Hum Genet* 2004;**114**:413-38. [14999561] [doi: 10.1007/s00439-004-1097-7]
- 3 Hirsch-Reinshagen V, Zhou S, Burgess BL, Bernier L, McIsaac SA, Chan JY, Tansley JH, Cohn JS, Hayden MR, Wellington CL. Deficiency of ABCA1 impairs Apolipoprotein E metabolism in brain. J Biol Chem 2004;279:41197-207. [15269 218] [doi:10.1074/jbc.M407962200]
- 4 Hooijmans CR, Kiliaan AJ. Fatty acids, lipid metabolism and Alzheimer pathology. *Eur J Pharmacol* 2008;**585**:176-96. [18378224] [doi: 10.1016/j.ejphar.2007.11.081]
- 5 Wahrle SE, Jiang H, Parsadanian M, Legleiter J, Han- X, Fryer JD, Kowalewski T, Holtzman DM. ABCA1 is required for normal central nervous system ApoE levels and for lipidation of astrocyte-secreted apoE. *J Biol Chem* 2004;279:40987-93. [15269217] [doi:10.1074/jbc. M407963200]
- 6 Puglilli L, Tanzi RE, Kovacs DM. Alzheimer's disease: Cholesterol connection. Nat Neurosci 2003;6: 345-51. [12658281] [doi:10.1038/ nn0403-345]
- 7 Shibata N, Kawarai T, Lee JH, Lee H-S, Shibata E, Sato C, Liang Y, Duara R, Mayeux RP, St George-Hyslop PH, Rogaeva E. Association studies of cholesterol metabolism genes (CH25H, ABCA1 and CH24H) in Alzheimer's disease. *Neurosci Lett* 2006;**391**:142-6. [16157450] [doi:10. 1016/j.neulet.2005.08.048]
- 8 Sundar PD, Feingold E, Minster RL, DeKosky ST, Kamboh MI. Genderspecific association of ATP-binding cassette transporter 1 (ABCA1) polymorphisms with the risk of lateonset Alzheimer's disease. *Neurobiol Aging* 2007;**28**:856-62. [16725 228] [doi:10.1016/j.neurobiolaging. 2006.04.005]
- 9 Fassbender K, Simons M, Bergmann C, Stroick M, Lutjohann D, Keller P, Runz H, Kuhl S, Bertsch T, von Bergmann K, Hennerici M,

Beyreuther K, Hartmann T. Simvastatin strongly reduces levels of Alzheimer's disease betaamyloid peptides Abeta 42 and Abeta 40 in vitro and in vivo. *Proc Natl Acad Sci U S A* 2001;51;**98**:5856-61. [11296263] [doi:10.1073/pnas.081620098]

- 10 Kojro E, Gimpl G, Lammich S, Marz W, Fahrenholz F. Low cholesterol stimulates the nonamyloidogenic pathway by its effect on the alpha secretase ADAM 10. Proc Natl Acad Sci U S A 2001;98:5815-20. [11309 494] [doi:10.1073/pnas.081612998]
- 11 Richard F, Amouyel P. Genetic susceptibility factors for Alzheimer's disease. *Eur J Pharmacol* 2001; 412:1-12. [11166730] [doi:10.1016/ S0014-2999(00)00903-1]
- 12 Tanzi RE, Bertram L. New Frontiers in Alzheimer's disease Genetics. *Neuron* 2001;**32**:181-4. [11683989] [doi: 10.1016/S0896-6273(01)00476-7]
- 13 Wang F, Jia J. Polymorphisms of cholesterol metabolism genes CYP46 and ABCA1 and the risk of sporadic Alzheimer's disease in Chinese. *Brain Res* 2007;1147:34-8. [17335784] [doi:10.1016/j.brainres. 2007.02.005]
- 14 Wolozin B, Kellman W, Ruosseau P, Celesia GG, Siegel G. Decreased prevalence of Alzheimer disease associated with 3-hydroxy-3methyglutaryl coenzyme A reductase inhibitors. Arch Neurol 2000; 57:1439-43. [11030795] [doi:10.100 1/archneur.57.10.1439]
- Wollmer MA, Streffer JR, Lütjohann D, Tsolaki M, Iakovidou V, Hegi T, Pasch T, Jung HH, Bergmann K, Nitsch RM, Hock C, Papassotiropoulos A. ABCA1 modulates CSF cholesterol levels and influences the age at onset of Alzheimer's disease. *Neurobiol Aging* 2003;24;421-6. [12600718] [doi:10.1016/S0197-45 80(02)00094-5]
- 16 Selkoe DJ. Alzheimer's Disease: Genes, Proteins, and Therapy. *Physiol Rev* 2001;81:741-66. [11 274343]
- Wavrant-De Vrièze F, Compton D, Womick M, Arepalli S, Adighibe O, Li L, Pérez-Tur J, Hardy J. ABCA1 polymorphisms and Alzheimer's disease. *Neurosci Lett* 2007;**416**:180-3. [17324514] [doi:10.1016/j.neulet. 2007.02.010]

kindly participation in this study. We also thank Iran Alzheimer Association for their sincere collaborations.

Conflict of interest: None declared.

- 18 Blacker D, Bertram L, Saunders AJ, Moscarillo TJ, Albert MS, Wiener H, Perry RT, Collins JS, Harrell LE, Go RC, Mahoney A, Beaty T, Fallin MD, Avramopoulos D, Chase GA, Folstein MF, McInnis MG, Bassett SS, Doheny KJ, Pugh EW, Tanzi RE; NIMH Genetics Initiative Alzheimer's Disease Study Group. Results of a high-resolution genome screen of 437 Alzheimer's Disease families. *Hum Mol Genet* 2003;12:23-32. [12 490529] [doi:10.1093/hmg/ddg007]
- 19 Liang Y, Lin S, Beyer TP, Zhang Y, Wu X, Bales KR, DeMattos RB, May PC, Li SD, Jiang XC, Eacho PI, Cao G, Paul SM. A liver X receptor and retinoid X receptor heterodimer mediates apolipoprotein E expression, secretion and cholesterol homeostasis in astrocytes. J Neurochem 2004;88:623-34. [14720212] [doi: 10.1111/j.1471-4159.2004.02183.x]
- 20 Katzov H, Chalmers K, Palmgren J, Andreasen N, Johansson B, Cairns NJ, Gatz M, Wilcock GK, Love S, Pedersen NL, Brookes AJ, Blennow K, Kehoe PG, Prince JA. Genetic variants of *ABCA1* modify Alzheimer disease risk and quantitative traits related to beta-amyloid metabolism. *Hum Mutat* 2004;**23**:358-67. [1502 4730] [doi:10.1002/humu.20012]
- 21 Li Y, Tacey K, Doil L, van Luchene R, Garcia V, Rowland C, Schrodi S, Leong D, Lau K, Catanese J, Sninsky J, Nowotny P, Holmans P, Hardy J, Powell J, Lovestone S, Thal L, Owen M, Williams J, Goate A, Grupe A. Association of *ABCA1* with late-onset Alzheimer's disease is not observed in a case-control study. *Neurosci Lett* 2004;**366**:268-71. [1528843] [doi:10.1016/j.neulet. 2004.05.047]
- 22 Raygani AV, Zahrai M, Raygani AV, Doosti M, Javadi E, Rezaei M, Pourmotabbed T. Association between apolipoprotein E polymorphism and Alzheimer disease in Tehran, Iran. *Neurosci Lett* 2005; 375:1-6. [15664112] [doi:10.1016/ j.neulet.2004.10.073]
- Wenham PR, Price WH, Blandell G. Apolipoprotein E genotyping by onestage PCR. *Lancet* 1991;337:1158-9. [1674030] [doi:10.1016/0140-6736(91)92823-K]
- 24 Corder ÉH, Saunders AM, Risch NJ,

Strittmatter WJ, Schmechel DE, Gaskell PC Jr, Rimmler JB, Locke PA, Conneally PM, Schmader KE, et al. Protective effect of apolipoprotein E type 2 allele for late onset Alzheimer disease. *Nat Genet* 1994;**7**:180-4. [7920638] [doi:10.10 38/ng0694-180]

- 25 Scott WK, Saunders AM, Gaskell PC, Locke PA, Growdon JH, Farrer LA, Auerbach SA, Roses AD, Haines JL, Pericak-Vance MA. Apolipoprotein E ɛ2 does not increase risk of early-onset sporadic Alzheimer's Disease. *Ann Neurol* 1997; 42:376-8. [9307262] [doi:10.1002/ana.410420317]
- 26 Pericak-Vance MA, Grubber J, Bailey LR, Hedges D, West S, Santoro L, Kemmerer B, Hall JL, Saunders AM, Roses AD, Small GW, Scott WK, Conneally PM, Vance JM, Haines JL. Identification of novel genes in late-onset Alzheimer's

Disease. *Exp Gerontol* 2000; **35**:1343-52. [11113612] [doi: 10.1016/S0531-5565(00)00196-0]

- 27 Holtzman DM, Bales KR, Tenkova T, Fagan AM, Parsadanian M, Sartorius LJ, Mackey B, Olney J, McKeel D, Wozniak D, Paul SM. Apolipoprotein E isoform-dependent amyloid deposition and neuritic degeneration in a mouse model of Alzheimer's disease. *Proc Natl Acad Sci U S A* 2000;97:2892-7. [1069 457] [doi:10.1073/pnas.050004797]
- 28 Berr C, Hauw JJ, Delaere P, Duyckaerts C, Amouyel P. Apolipoprotein E allele epsilon 4 is linked to increased deposition of the amyloid beta-peptide A-beta in cases with or without Alzheimer's disease. *Neurosci Lett* 1994;178:221-4. [7824200] [doi:10.1016/0304-3940 (94)90763-3]
- (94)90763-3] 29 Gomez-Isla T, West HL, Rebeck GW, Harr SD, Growdon JH, Locascio

JJ, Perls TT, Lipsitz LA, Hyman BT. Clinical and pathological correlates of apolipoprotein E epsilon 4 in Alzheimer's disease. *Ann Neurol* 1996;**39**:62-70. [8572669] [doi:10. 1002/ana.410390110]

- 30 Rose DA. Alzheimer Diseases: A model of gene mutations and susceptibility polymorphisms for complex psychiatric diseases. Am J Med Genet 1998;81:49-57. [9514588] [doi:10.1002/(SICI)1096-8628(1998 0207)81:1<49::AID-AJMG10>3.0.C O;2-W]
- 31 Perry RT, Collins JS, Harrell, Acton RT, Go RC. Investigation of association of polymorphisms in eight genes in southeastern African American Alzheimer disease patients as compared to age-matched controls. Am J Med Genet 2001; 105:332-42. [11378846] [doi:10.10 02/ajmg.1371]