



## Fermentation Quality of Ensiled Water Hyacinth (*Eichhornia crassipes*) as Affected by Additives

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**ABSTRACT:** A lab-scale ensiling study was carried out to investigate the fermentation quality of water hyacinth (WH) supplemented with molasses, rice bran, as an absorbent, and an inoculant in the form of fermented vegetable juice and their combinations. After wilting the water hyacinths for 7 h to a dry matter (DM) content of 240 to 250 g/kg, the following treatments were applied: i) Control (C), WH only; ii) WH with sugarcane molasses at 40 g/kg WH (CM); iii) WH inoculated with fermented vegetable juice at 10 ml/kg WH (CI); iv) CM and CI (CMI) combined; v) WH with 150 g rice bran/kg WH (CA); vi) CA and CI combined (CAI); vii) CA and CM combined (CAM); and viii) CA, CM and CI combined (CAMI). After application of additives, the differently treated forages were mixed and ensiled in triplicates in 1,500-ml polyethylene jars. After ensiling for 3 d, pH values in all treatments, except C and CI, had decreased to approximately 4.0 and remained low till 14 d. After 56 d, pH had increased between 0.4 to 0.9 pH-units compared to those at 14 d. The ammonia nitrogen (NH<sub>3</sub>-N) concentration ranged from an acceptable level in treatment CM (8 g/kg N) to a high NH<sub>3</sub>-N value in treatment CMI (16 g/kg N). Lactic acid formation was higher in CI than in all other treatments. Butyric acid contents, which indicate badly fermented silages, were low in all silages (<2 g/kg DM). There were two-way interactions (p-values from <0.001 to 0.045) for almost all fermentation end-products and pH, except for the molasses×inoculant interaction on NH<sub>3</sub>-N (p = 0.26). Significant 3-way interactions were found on all observed variables except for weight losses of silages. It is concluded that conserving wilted WH as silage for ruminants may be improved by the addition of molasses or rice bran. (**Key Words:** Inoculant, Molasses, Rice Bran, Silage, Water Hyacinth)

### INTRODUCTION

Ruminant production plays an important role in the predominantly agricultural economy of Vietnam (Trach, 1998). Due to the extreme scarcity of land for grazing and grass cultivation as well as adequate feeds, particularly in the dry season, the possibility of using fibrous feeds such as crop residues and water hyacinth has become more interesting for the improvement of ruminant production.

Water hyacinth (*Eichhornia crassipes*) (WH) has become the major floating water weed of tropical and subtropical regions worldwide because of its rapid growth (De Groote et al., 2003). The capacity for vegetative

reproduction allows this aquatic plant to quickly occupy all available space (Gutiérrez et al., 1996). Water hyacinth is available all year round and is widespread on freshwater wetlands of the Mekong Delta, especially in standing water (MWBP/RSCP, 2006).

Water hyacinth has received much attention in recent years due to its potential benefits as animal fodder, aquafeed, water purification, fertilizer, biogas production, etc. Numerous studies have evaluated WH as a feed source to ruminants (Baldwin et al., 1975; Abdalla et al., 1987; Agarwala, 1988; Islam et al., 2009; Sophal, 2010), fish (Okoye et al., 2002; Abdel-Fattah, 2003) and other animals (Dey et al., 1983; Men et al., 2002, 2006). Water hyacinth control methods fall into three main categories: physical, chemical and biological (Julien et al., 2001). Its use as a livestock feed is considered as an effective physical control method.

Ensiling aquatic weeds, especially WH, could become important in humid tropical regions where complete drying into hay is difficult (National Academy of Sciences, 1976). Effects of additives on the quality of WH silage have been

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evaluated by some researchers (Byron et al., 1978; Lowilai et al., 1994; 1995; Manh et al., 2002). Effluent should always be avoided because it represents a loss of water-soluble nutrients which may reach 10% in very wet crops (Wilkins, 2005). To reduce effluents, McDonald et al. (1991) suggested wilting of forage crops to achieve a dry matter (DM) content of 260 to 300 g/kg. Development of undesirable microorganisms may be reduced, a desirable fermentation will be promoted and livestock intake can be increased by wilting (Graci, 1999).

Additional fermentable carbohydrates are necessary to promote fermentation of most tropical plants and to achieve acceptable silage quality (Bagnall et al., 1974). Sugarcane molasses is used widely for silage making in Vietnam due to its high level of water soluble carbohydrates. This additive has been shown to decrease pH and ammonia levels in silages (McDonald et al., 2011). As WH has a low DM content, wilting is an option to reduce silage losses. However, weather conditions are not always favourable for adequate wilting, and the use of a suitable absorbent can be considered as an alternative to reduce effluents (McDonald et al., 1991). Rice bran is one of the main by-products of the rice milling industry. It has been used as an absorbent in silage (Lowilai et al., 1995; Yokota et al., 1998) and is readily available in large quantities in the Mekong Delta of Vietnam. Homofermentative lactic acid bacteria (LAB) alone or in combination with heterofermentative LAB are commonly used as silage additives to improve silage fermentation (McDonald et al., 1991). Inoculation rates of  $10^5$  to  $10^6$  colony forming units (cfu)/g herbage are often recommended (Henderson, 1993). The effect of adding bacterial inoculants to wilted silage may increase lactic acid production and result in a faster decrease of silage pH if the forage contains at least 20 g water soluble carbohydrates (WSC)/kg fresh matter (Pettersson and Lindgren, 1990). Ensiling studies with WH, using additives such as rice bran, cassava root, ground corn, wheat bran (El-Serafy et al., 1989; Lowilai et al., 1993), organic acids (Byron et al., 1975) or urea and rice straw (Wanapat et al., 1985; Chkraborty et al., 1991), have been promising but have not yet been investigated in Vietnam.

The aim of this study was to evaluate the influence of molasses, rice bran and fermented vegetable juice and their combined effects on the fermentation quality of ensiled WH.

## MATERIALS AND METHODS

### Study site and experimental design

The experiment was conducted at Department of Animal Sciences, College of Agriculture and Applied Biology, Can Tho University, Vietnam between March and May 2011. A completely randomized experimental design

was used, with eight treatments and three silo replicates for each treatment. Treatments were: i) Control (C), WH only; ii) WH and sugarcane molasses, at 40 g/kg wilted WH (CM); iii) WH inoculated with fermented vegetable juice at 10 ml/kg WH (CI); iv) CM and CI (CMI) combined; v) WH and 150 g rice bran/kg WH (CA); vi) CA and CI combined (CAI); vii) CA and CM (CAM) combined; and viii) CA, CM and CI combined (CAMI).

*Plant material:* Water hyacinths at 6 to 7 wks of age were collected along Tien River in Vinh Long province. The petioles were cut 4 to 5 cm from the base of the plant. Both petiole and leaf fractions were cut by hand into 4 to 5 cm pieces, spread in a 4 to 5 cm thick layer on plastic sheeting corresponding to a weight of 4 kg/m<sup>2</sup>. The initial dry matter (DM) content was approximately 90 to 110 g/kg WH and the material was sun-dried for approximately 7 h to a DM content of 240 to 250 g/kg. The plant material was turned every 30 min during wilting. Temperature and humidity were recorded every 30 min by a digital thermo-hygrometer (NJ-2099-TH, Nakata, Japan) and ranged from 30 to 42°C and 34 to 74%, respectively. During wilting, two representative samples were collected at two occasions to monitor approximate DM changes using a microwave oven (Undersander et al., 1993). The wilted material was subsequently transferred to Can Tho University for ensiling.

*Preparation of the inoculants suspension:* A local cabbage type vegetable *cải bẹ* or *cải tía xài* (*Brassica campestris* L.), which is a very popular food item in Vietnam, was acquired from a local market and fermented in the following way. Yellow or rotten leaves were removed and the plants were thoroughly washed and sun-dried for approximately 2 h. The vegetable was first blanched in boiling water for 2 min to minimize enzyme activity. After blanching, it was drained and immersed in cool sterile water in a capped bucket. Fermentation was done for a period of one week after which one litre of cooled sterile water and 5 g salt was added to one kg vegetable. The juice was then extracted by squeezing and filtering.

*Mini-silo preparation:* The wilted WH was used in the present experiment with or without additives as outlined above. Molasses was collected from a local sugarcane factory. Depending on treatments, the order of addition was: vegetable juice, followed by rice bran and molasses. The wilted material was spread on a plastic sheeting, sprayed with the vegetable juice extract (10 ml/kg WH), or “dressed” with rice bran (150 g/kg WH) followed by the addition of molasses (40 g/kg WH). All ingredients were then mixed thoroughly.

A total of 850 g material was filled into 1500-ml gastight polyethylene jars (Duytan Corp., Ho Chi Minh city, Vietnam). The mixtures were compressed by hand to remove as much air as possible. Lids were sealed with

silicone caulk. Fermentation gases could escape via water-filled plastic siphons, which were fitted on the silo lids.

Three mini-silos were prepared for each treatment, giving a total of 24 silos. They were stored indoors at a temperature of 27 to 31°C and a relative humidity of 65 to 75%. To follow the pH changes over time, a second set of silos were made of the same plant material. Two silos per treatment were opened after 3, 7, and 14 d of storage.

### Measurements and laboratory analyses

Before ensiling, all samples of the mixed material were collected for the analysis of pH, DM and chemical composition. Weight losses during storage were determined by weighting the silos. Weights of the filled mini-silos were recorded at 0, 3, 7 and 14 d after filling at 07:30 h and then at weekly intervals until opening and sampling after 56 d. Mouldy silage found on the surface was discarded before sampling. The contents from each mini-silo were transferred into plastic bags, mixed thoroughly and sampled for chemical analyses.

The silages were analyzed for pH, chemical composition, including fermentation end-products. Concentration of LAB in the fermented vegetable juice and the wilted WH was determined based on the technique described by International Organization for Standardization (1998) (ISO 15214:1998).

Dry matter content of the wilted and ensiled WH was determined by oven-drying at 105°C for 12 h. Dry matter (DM) (967.03), crude protein (CP) (984.13), ash (942.05), acid detergent fibre (ADFom) 973.18), ammonia nitrogen (NH<sub>3</sub>-N) (941.04) were analyzed as described by AOAC (1990). Neutral detergent fibre (NDFom) was analyzed according to Van Soest et al. (1991). Amylase and sodium sulfite was not used for NDFom determination and both ADFom and NDFom were expressed exclusive of residual ash. Water soluble carbohydrates were analysed following the protocol of Udén (2006). Ethanol, acetic, propionic, butyric and lactic acids were measured by HPLC (Model 10A, Shimadzu Corp., Tokyo, Japan). After opening the mini-silos, 10 g of silage from each replicate were homogenized in 20 ml distilled water, and pH of the filtrate was determined using a pH meter (Mi150, Martini, Romania) with a glass electrode.

### Statistical analysis

Data for each analysis consisted of 2 (levels of molasses)×2 (levels of rice bran)×2 (levels of inoculants)×3 (replicated silos) = 24. All observations were used in the analysis of variance of chemical composition, fermentation quality and weight loss of silage DM. The GLM procedure of Minitab software, version 16.1 (Minitab Inc., 2010) was used and with the following model of fixed factors and their

interactions:

$$Y_{ijkl} = \mu + M_i + A_j + I_k + MA_{ij} + MI_{ik} + AI_{jk} + MAI_{ijk} + e_{ijkl}$$

where  $Y_{ijkl}$  = the dependent variable,  $\mu$  = the overall mean,  $M_i$  = effect of sugarcane molasses,  $A_j$  = effect of rice bran as absorbent,  $I_k$  = effect of inoculants and  $e_{ijkl}$  = the random error.

Pearson correlation coefficients among all dependent variables were also calculated using the same software.

## RESULTS

### Chemical composition of silage ingredients

The chemical composition of the wilted WH, rice bran and sugarcane molasses is shown in Table 1. Rice bran and molasses had a DM content of 920 and 814 g/kg, respectively, and wilted WH before ensiling had a DM content of 244 g/kg. The initial content of WSC was 4.3 g/kg DM in wilted WH. Concentrations of LAB in the fermented vegetable juice and the wilted WH were  $2.1 \times 10^7$  cfu/ml and  $1.2 \times 10^7$  cfu/g wilted WH, respectively.

### The fermentation quality of the silages

No free liquid or effluent was found in any of the mini-silos. On visual inspection (d 56), spoilage moulds were found on the top of C and CA silos. Mould development was virtually absent in CAM silage, while the remaining silages of CM, CI, CMI, CAI and CAMI had little or no mould growth. Silage colour was greenish yellow and changed to a brownish yellow colour after 56 d of storage. Addition of rice bran caused increase in DM content from 242 to 301 g/kg and by adding molasses, it was increased further to 334 g/kg (Table 2). The results show differences among treatments for all variables ( $p < 0.001$ ). Both rice bran and molasses increased WSC levels, each by approximately 5 g/kg DM. Rice bran predictably decreased CP and NDFom contents ( $p < 0.001$ ).

There were overall differences ( $p < 0.001$ ) among treatments in pH, fermentation end-products and weight

**Table 1.** Dry matter and chemical composition (g/kg DM) of the wilted water hyacinth, rice bran and sugarcane molasses (n = 3)

Item <sup>a</sup>	Water hyacinth		Rice bran		Sugarcane molasses	
	Mean	SD	Mean	SD	Mean	SD
Dry matter	244	0.83	920	1.26	814	4.22
Crude protein	155	0.61	82	0.52	33	0.91
Ash	165	1.45	117	0.27	64	0.47
ADFom	364	1.66	337	1.20	nd	nd
NDFom	622	3.81	479	22.1	nd	nd
WSC	4.3	-	nd	nd	nd	nd

<sup>a</sup> ADFom = Acid detergent fibre; NDFom = Neutral detergent fibre; WSC = Water soluble carbohydrates; nd = Not determined.

**Table 2.** Dry matter (DM) and concentrations (g/kg DM) of water soluble carbohydrates (WSC), crude protein (CP), neutral detergent fibre (NDFom) and ash of water hyacinth (WH) silages after ensiling for 56 d (n = 3)

	Treatments*								SEM	p
	C	CM	CI	CMI	CA	CAI	CAM	CAMI		
DM	242 <sup>g</sup>	258 <sup>e</sup>	234 <sup>h</sup>	252 <sup>f</sup>	301 <sup>d</sup>	306 <sup>c</sup>	334 <sup>a</sup>	318 <sup>b</sup>	0.57	<0.001
WSC	0.7 <sup>c</sup>	1.1 <sup>c</sup>	0.4 <sup>c</sup>	1.1 <sup>c</sup>	5.4 <sup>b</sup>	6.9 <sup>b</sup>	12.1 <sup>a</sup>	11.4 <sup>a</sup>	0.52	<0.001
CP	157 <sup>a</sup>	164 <sup>a</sup>	156 <sup>a</sup>	156 <sup>a</sup>	139 <sup>b</sup>	136 <sup>b</sup>	135 <sup>b</sup>	139 <sup>b</sup>	1.82	<0.001
NDFom	587 <sup>a</sup>	500 <sup>d</sup>	584 <sup>a</sup>	520 <sup>c</sup>	557 <sup>b</sup>	549 <sup>b</sup>	529 <sup>c</sup>	522 <sup>c</sup>	3.04	<0.001
Ash	180 <sup>a</sup>	164 <sup>bc</sup>	178 <sup>a</sup>	174 <sup>ab</sup>	153 <sup>cd</sup>	160 <sup>cd</sup>	152 <sup>d</sup>	152 <sup>d</sup>	2.36	<0.001

\* C = WH only (Control); CM = C+40 g sugarcane molasses/kg WH; CI = C+10 ml inoculant/kg WH; CMI = CM+CI; CA = C+150 g rice bran/kg WH; CAI = CA+CI; CAM = CA+CM; CAMI = CA+CM+CI.

SEM = Standard error of means. <sup>a,b,c,d,e,f,g,h</sup> Means within rows with different letters differ (p<0.05).

**Table 3.** Fermentation end-product concentrations, pH and weight losses in water hyacinth (WH) silages after 56 days of ensiling (g/kg DM or as otherwise stated) after addition of molasses (M), absorbent (A), inoculants (I) or combinations

Item	Treatments*								SEM	p	P						
	C	CM	CI	CMI	CA	CAI	CAM	CAMI			M	A	I	M×A	M×I	A×I	M×A×I
Lactic acid	10.9 <sup>b</sup>	21.1 <sup>f</sup>	95.4 <sup>a</sup>	11.7 <sup>g</sup>	60.4 <sup>c</sup>	69.2 <sup>b</sup>	44.9 <sup>d</sup>	40.2 <sup>e</sup>	0.06	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001
Acetic acid	1.5 <sup>b</sup>	2.0 <sup>a</sup>	1.0 <sup>c</sup>	1.6 <sup>b</sup>	1.0 <sup>c</sup>	0.7 <sup>d</sup>	1.0 <sup>c</sup>	0.9 <sup>c</sup>	0.03	<0.001	<0.001	<0.001	<0.001	<0.001	0.005	<0.001	0.653
Propionic acid	0.4 <sup>f</sup>	1.5 <sup>a</sup>	0.2 <sup>g</sup>	1.0 <sup>c</sup>	0.7 <sup>d</sup>	0.5 <sup>e</sup>	1.3 <sup>b</sup>	1.0 <sup>c</sup>	0.02	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	0.015	<0.001
Butyric acid	0.3 <sup>g</sup>	1.4 <sup>d</sup>	1.6 <sup>bc</sup>	1.7 <sup>b</sup>	1.5 <sup>c</sup>	0.4 <sup>f</sup>	1.9 <sup>a</sup>	0.9 <sup>e</sup>	0.02	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001
Ethanol	0.5 <sup>a</sup>	0.2 <sup>c</sup>	0.3 <sup>b</sup>	0.2 <sup>cd</sup>	0.2 <sup>c</sup>	0.1 <sup>d</sup>	0.2 <sup>b</sup>	0.2 <sup>c</sup>	0.01	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	0.006	<0.001
pH	5.4 <sup>a</sup>	4.9 <sup>cd</sup>	5.2 <sup>b</sup>	4.6 <sup>f</sup>	4.9 <sup>c</sup>	4.7 <sup>e</sup>	4.8 <sup>de</sup>	4.8 <sup>de</sup>	0.02	<0.001	<0.001	<0.001	0.001	<0.001	0.045	0.010	0.002
NH <sub>3</sub> -N (g/kg N)	15.2 <sup>a</sup>	8.4 <sup>c</sup>	14.4 <sup>a</sup>	15.6 <sup>a</sup>	11.2 <sup>b</sup>	13.4 <sup>ab</sup>	14.7 <sup>a</sup>	10.9 <sup>bc</sup>	0.55	<0.001	0.01	0.05	0.007	0.001	0.259	<0.001	<0.001
Weight loss	39.8 <sup>a</sup>	35.0 <sup>ab</sup>	42.0 <sup>a</sup>	45.2 <sup>a</sup>	26.1 <sup>bc</sup>	22.4 <sup>c</sup>	18.2 <sup>c</sup>	21.0 <sup>c</sup>	2.45	<0.001	0.135	<0.001	0.115	0.281	0.054	0.075	0.84

\* C = WH only (control); CM = C+40 g sugarcane molasses/kg WH; CI = C+10 ml inoculant/kg WH; CMI = CM+CI; CA = C+150 g rice bran/kg WH; CAI = CA+CI; CAM = CA+CM; CAMI = CA+CM+CI.

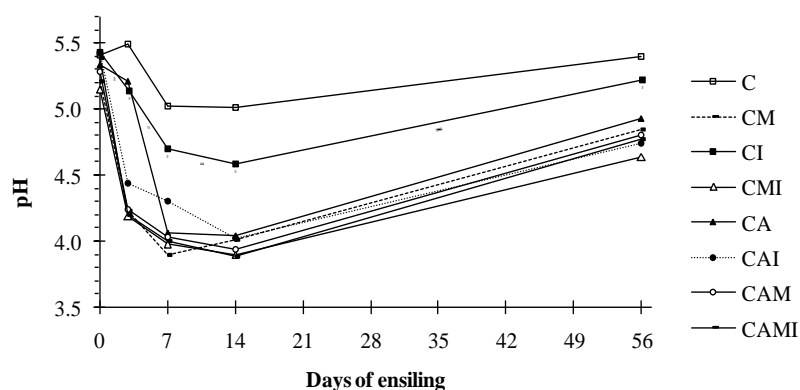
SEM = Standard error of means. <sup>a, b, c, d, e, f, g, h</sup> Means within rows with different letters differ (p<0.05).

losses (Table 3). All additives influenced the formation of end-products and pH (p<0.001). There were also two-way interactions (p-values from <0.001 to 0.045) for almost all fermentation end-products and pH except for the molasses × inoculant interaction on NH<sub>3</sub>-N (p = 0.26). Significant 3-way interactions were found on all observed variables except for weight losses of silage.

Ammonia nitrogen levels in silages were significantly decreased by the addition of molasses (M) and rice bran (A) but not by inoculants (I), while weight losses were decreased only by rice bran. Lactic acid formation was higher (p<0.001) in CI (95.4 g/kg DM) than in all other

treatments. Butyric acid contents were relatively low, with the highest value in CAM (1.9 g/kg DM) and the lowest value in C (0.3 g/kg DM). Ammonia nitrogen differed among treatments (p<0.001), with treatment CM giving the lowest value (8.4 g/kg N), and treatment CMI giving the highest value (15.6 g/kg N).

After ensiling for 7 d, pH values in all treatments, except C and CI, had decreased to approximately 4.0 and remained on that level until at least d 14 (Figure 1). After 56 d, pH had increased to approximately 4.8. Both C and CI silages decreased only to pH 5.0 and 4.5, respectively, at d 7 and then increased between 14 and 56 d to approximately

**Figure 1.** Changes in pH during ensilage of water hyacinth (WH). C = WH only (control); CM = C+40 g sugarcane molasses/kg WH; CI = C+10 ml inoculant/kg WH; CMI = CM+CI; CA = C+150 g rice bran/kg WH; CAI = CA+CI; CAM = CA+CM; CAMI = CA+CM+CI.

5.3 (Figure 1).

## DISCUSSION

Because of its rapid spoilage and high moisture content, it is recommended to wilt WH in the shade for 2 d (Göhl, 1981). Prolonged wilting of crops has been shown to result in lower WSC of the crop (Uchida et al., 1989; Pettersson and Lindgren, 1990), and to reduce acetic acid and  $\text{NH}_3\text{-N}$  production (Liu et al., 2011).

As shown in Figure 1, the pH of treatment CI gradually decreased from d 3 until d 14. Applying fermented juice made from fresh alfalfa and glucose fermented for 2 d was shown to improve alfalfa silage quality (Ohshima et al., 1997). In our study, inoculation with LAB from fermented vegetable juice did not initiate a desirable fermentation of WH with an inherently low WSC content (4 g/kg DM). It is well known that WSC content is a crucial and limiting factor of silage fermentation (Liu et al., 2011). Haigh and Parker (1985) regarded 25 to 35 g/kg in fresh herbage as minimal WSC concentrations for successful ensiling of unwilted herbage. The WSC contents of wilted WH in the present study and molasses according to McDonald et al. (2011), was 4 and 700 g/kg DM, respectively. Based on values above, an addition of 40 g molasses to the wilted WH would result in approximately 24 g WSC/kg in the CM mixture (1 g from WH and 23 g from molasses). The addition of 40 g molasses per kg WH should therefore supply enough WSC to avoid any shortage of substrate during the fermentation process. The pH course over time in Figure 1 confirms that since only treatments C and CI appeared to lack WSC, which made it difficult for the LAB to reduce silage pH to the same level in the other treatments. Figure 1 indicates further that the addition of rice bran (A) had a similar effect on the WSC level of the WH silage as the addition of molasses. Manh et al. (2002) found that WH (15% DM) was successfully preserved by adding 4.5% of molasses (fresh matter basis). After ensiling for 3 d, the pH value had decreased from 5.1 to 4.0 and remained stable for 30 d. Rice bran contains only 53 g WSC/kg DM compared to 700 g WSC/kg DM in molasses (Lowilai et al., 1994; McDonald et al., 2011). Even when the amount of applied rice bran was 4 to 5 times higher than molasses, the WSC contribution from rice bran was only approximately 7 g/kg forage. Molasses will therefore promote fermentation more than rice bran (Ngoan et al., 2000). Silage pH is a standard indicator for the course of fermentation. However, it is heavily influenced by the DM content which means that it is only applicable as an indicator at lower DM contents (Weissbach, 1996). At higher DM contents, silages stabilise at a higher pH values (Leibeinsperger and Pitt, 1988). If the pH fall is insufficient to prevent development of coliform and clostridial bacteria, it may rise again (Wilkinson, 2005).

However, the lack of butyrate in our silages excludes clostridia as the causative agents. Mould growth in the top layer of some of our silos suggests that yeasts and moulds might have been stimulated by the ingress of small amounts of air (Elferink et al., 1999). These fungi are able to metabolize lactate in the presence of oxygen, which eventually will lead to increased pH values in silages. The  $\text{NH}_3\text{-N}$  concentration in silage reflects the degree of amino acid degradation (Driehuis et al., 2001) if ammonia formation from nitrate and nitrite in the forage is considered. An acceptable level of  $\text{NH}_3\text{-N}$  in silage should be below 110 g/kg N (Church, 1991) which only CM and CAMI silages achieved. Slightly increased ammonia levels should however not be harmful to cattle, if the animals' production level is not very high.

Although commercial inoculants have proved effective in improving silage fermentation (Weinberg and Muck, 1996; Arriola et al., 2011), successful control of fermentation depends on several factors, primarily the inoculation rate (at least  $10^5$  cfu/g fresh crop), and the presence of an adequate level of WSC (Islam et al., 2011; McDonald et al., 2011).

There are only a limited number of reports on feeding WH to animals. Pigs showed an immediate acceptance of WH without drying (Men et al., 2006) while Islam et al. (2011) claimed that a major constraint for its use as a ruminant feed is the high water content. Cattle preferred WH silage containing the highest level of additives, 4 kg dried citrus pulp and 1 kg sugarcane molasses/100 kg pressed WH residue (Baldwin et al., 1974). However, the higher butyric acid contents of the aquatic plant silages may have a depressing effect on DM intake (Linn et al., 1975). In the present study, the CAM silage had the highest level of butyric acid (1.9 g/kg DM), which was below the acceptable limit of 2 g/kg DM silage stated by Castle and Watson (1985).

During feed scarcity period in the dry season, WH could be ensiled for ruminant feeding. An additional carbohydrate source such as molasses, 4% as shown in this research, is necessary to produce acceptable silage. Baldwin et al. (1975) failed to get a good fermentation quality with WH due to excessive losses of soluble material in the effluents. The addition of suitable absorbents to reduce moisture content and to minimise WSC losses with the effluent are therefore needed. Absorbents have the additional advantage that the wilting process can be shortened or omitted and that the silage can be supplemented with suitable nutrients. Yokota et al. (1998) found that a combination of molasses and rice bran improved fermentation quality of ensiled napier grass (*Pennisetum purpureum* Schum.) more than each additive alone. Although the pH values of the silages were not as low as expected even in the best treatments, the silages were preserved well as illustrated by low acetic and

butyric acid as well as  $\text{NH}_3\text{-N}$  concentrations. Water hyacinths were successfully ensiled with either 4% of sugarcane molasses, 15% of rice bran (fresh weight basis) or with a combination of the two additives. The best fermentation quality in the present study was achieved with silage treatments CM, CA and CAMI. Both rice bran and molasses are readily available all year round, but rice bran is more expensive than molasses. If the choice stands between molasses or rice bran, then molasses should be the first choice with regard to improving silage fermentation.

## CONCLUSIONS

The present results suggest that there are marked influences on the formation of end-products and on pH in WH silages by addition of molasses or rice bran. The good results obtained in lab-scale silos suggest that the ensiling of WH on a larger scale could be a viable alternative to farmers and ensiled WH could serve as a source of roughage for ruminants in the tropics. Focus should be put on the maintenance of anaerobic conditions in silos because that is a vital precondition for good fermentation quality. Under practical conditions, the advantages of these additives must be balanced against their cost and efficacy.

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