Inverse association between *trans* isomeric and long-chain polyunsaturated fatty acids in cord blood lipids of full-term infants^{1–3}

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ABSTRACT

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Background: Previous studies showed significant inverse correlations between values of *trans* isomeric and long-chain polyunsaturated fatty acids in plasma lipids of preterm infants and healthy children aged 1–15 y.

Objective: We sought to evaluate the same correlations in full-term infants at birth.

Design: We studied healthy full-term infants (n = 42) born after normal pregnancies and deliveries. All infants had a family history of atopy (both parents or one of the parents and a sibling had atopic symptoms). The fatty acid composition of venous cord blood lipids was determined by high-resolution capillary gasliquid chromatography.

Results: The mean (±SEM) sum of *trans* fatty acids was $0.49 \pm 0.02\%$ by wt in phospholipids, $2.47 \pm 0.20\%$ by wt in cholesterol esters, $1.73 \pm 0.09\%$ by wt in triacylglycerols, and $1.59 \pm 0.07\%$ by wt in nonesterified fatty acids. Linear correlation analysis showed significant inverse correlations between the sum of *trans* fatty acids and both arachidonic acid and docosahexaenoic acid in phospholipids (r = -0.56, P < 0.001, and r = -0.48, P = 0.01, respectively), cholesterol esters (r = -0.52, P < 0.001, and r = -0.41, P = 0.007, and r = -0.41, P = 0.007, and r = -0.41, P = 0.006, respectively).

Conclusion: Because *trans* fatty acids in the fetal circulation must originate from the maternal diet, our results indicate that maternal exposure to *trans* fatty acids may represent a previously neglected variable that inversely influences long-chain polyunsaturated fatty acid status in full-term infants at birth. *Am J Clin Nutr* 2001;74:364–8.

KEY WORDS Arachidonic acid, cord blood, docosahexaenoic acid, full-term infant, long-chain polyunsaturated fatty acid, *trans* fatty acid, maternal diet, maternal nutrition, prenatal nutrition, pregnancy

INTRODUCTION

In human diets, *trans* fatty acids are found in the meat and milk of ruminant animals, but the major sources of *trans* fatty acids are partially hydrogenated vegetable and fish oils. In North America, *trans* fatty acid intakes of pregnant and breast-feeding women were estimated to be 6.4 g/d in the United States (1) and 6.9 g/d in Canada (2). In the United Kingdom, *trans* fatty acid intakes were reported to be ≈ 5.6 g/d in men and 4.0 g/d in women (3).

However, many studies consistently reported wide interindividual variation in dietary exposure to *trans* fatty acids (1–5).

The potential of *trans* fatty acids to induce atherogenic changes in plasma cholesterol profiles was shown convincingly (6, 7). In addition, the ability of *trans* fatty acids to interfere with essential fatty acid metabolism (8, 9) may be of practical importance in pediatrics. Evidence of this interference in humans includes significant inverse correlations between plasma values of *trans* isomeric and long-chain polyunsaturated fatty acids, both in preterm infants (10) and in healthy children aged 1–15 y (11).

Because long-chain polyunsaturated fatty acids play an important role in early human development (12–15), interference with their availability caused by exposure to *trans* fatty acids is of special concern during the perinatal period. Indeed, an expert panel on *trans* fatty acids and early development said that "Additional research is needed to determine...whether the conversion in infants of linoleic acid and α -linolenic acid to arachidonic acid and docosahexaenoic acid, respectively, is influenced by concentrations of *trans* fatty acids in tissue lipids or diet..." (1). In the present article, we report data on the associations between *trans* isomeric and long-chain polyunsaturated fatty acids in the cord blood lipids of full-term infants with an atopic trait.

SUBJECTS AND METHODS

Subjects

We studied 42 singleton full-term infants with a mean (\pm SEM) birth weight of 3461 \pm 53 g and a mean birth length of 51.8 \pm 0.3 cm. The infants were enrolled in an Austrian nutritional trial investigating the effects of different feeding regimens on the prevention of allergy in high-risk infants. All the infants had a positive family history of atopy; both parents or one of the parents and a sibling had atopic symptoms. Maternal age ranged from 22 to 35 y (mean: 28.2 \pm 0.5 y). None of the mothers

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Concentrations of selected fatty acids in cord blood lipids of 42 full-term infants¹

	Phospholipids	Cholesterol esters	Triacylglycerols	Nonesterified fatty acids		
	% by wt					
Sum of saturated fatty acids	52.70 ± 0.58	28.79 ± 0.96	41.40 ± 0.81	58.76 ± 0.97		
Sum of cis monounsaturated fatty acids	14.34 ± 0.27	39.42 ± 0.69	37.97 ± 0.84	21.92 ± 0.68		
Sum of <i>trans</i> fatty acids	0.49 ± 0.02	2.47 ± 0.20	1.73 ± 0.09	1.59 ± 0.07		
n-6 PUFAs						
18:2n-6	6.06 ± 0.20	16.10 ± 0.74	11.32 ± 0.48	9.14 ± 0.43		
20:4n-6	13.82 ± 0.36	9.47 ± 0.54	2.68 ± 0.20	4.44 ± 0.30		
Ratio of 20:4n-6 to 18:2n-6	2.37 ± 0.10	0.60 ± 0.03	0.25 ± 0.02	0.53 ± 0.05		
Sum of n-6 long-chain PUFAs	20.88 ± 0.50	11.26 ± 0.59	5.43 ± 0.31	6.61 ± 0.39		
n-3 PUFAs						
18:3n-3	0.03 ± 0.01	0.16 ± 0.03	0.24 ± 0.03	0.27 ± 0.02		
22:6n-3	4.42 ± 0.22	0.68 ± 0.05	0.80 ± 0.08	1.12 ± 0.10		
Ratio of 22:6n-3 to 18:3n-3	302 ± 67	8.65 ± 0.85	13.54 ± 8.20	5.88 ± 0.69		
Sum of n-3 long-chain PUFAs	5.03 ± 0.23	0.97 ± 0.07	1.22 ± 0.15	1.31 ± 0.10		

 ${}^{1}\overline{x} \pm$ SEM. Sum of saturated fatty acids: 12:0 + 14:0 + 16:0 + 17:0 + 18:0 + 20:0 + 22:0 + 24:0; sum of *cis* monounsaturated fatty acids: 14:1n-5 + 16:1n-7 + 18:1n-9 + 18:1n-7 + 20:1n-9 + 22:1n-9 + 24:1n-9; sum of *trans* fatty acids: *t*-16:1 + *t*-18:1 + *ct*, *tc*, *tt*-18:2; sum of n-6 long-chain polyunsaturated fatty acids (PUFAs): 20:2n-6 + 20:3n-6 + 20:4n-6 + 22:4n-6 + 22:5n-6; sum of n-3 long-chain PUFAs: 20:3n-3 + 20:5n-3 + 22:5n-3 + 22:5n-3.

followed any self-restrictive diet. The pregnancy and delivery were normal in all cases. About two-thirds of the mothers (27 of 42) decided to breast-feed.

Methods

Venous cord blood was collected into tubes containing EDTA (1 g/L) and was centrifuged immediately (1500 \times g for 5 min at room temperature). Blood plasma was stored at -20 °C until analyzed. Blood samples were transported frozen to the laboratory in Hungary; all samples were thawed only once.

Plasma lipids were extracted from 0.25 mL plasma with chloroform and methanol (16) after addition of internal standards (phosphatidylcholine-, cholesterol-, and triacylglycerol-derived and free pentadecanoic acids, 15:0). Lipid classes were separated on silica gel plates (no. 5721; Merck, Darmstadt, Germany) with one 8-cm run of hexane, diethylether, chloroform, and acetic acid (70:20:10:1, by vol). Fatty acids in plasma phospholipids, cholesterol esters, triacylglycerols, and nonesterified fatty acids were transesterified with methanol and hydrochloric acid (17). Fatty acid methyl esters were determined by high-resolution capillary gas-liquid chromatography with a Finnigan 9001 chromatograph (Finnigan/Tremetrics Inc, Austin, TX) with split injection (1:15) and a flame ionization detector. A 40-m cyanopropyl column (DB-23; J & W Scientific, Folsom, CA) was used. The temperature program was as follows: an initial temperature of 100°C for 0.1 min, followed by a temperature increase of 40°C/min up to 180°C, a 1-min isotherm period, a temperature increase of 2°C/min up to 200°C, a 1-min isotherm period, a temperature increase of 10°C/min up to 240°C, and a 9.9-min isotherm period. The constant linear velocity was 0.3 m/s (referred to 100°C).

Identification of fatty acids was confirmed by comparison with authentic standards (NuChek Prep, Elysian, MN). Fatty acid results were expressed as % by wt of 31 fatty acids detected with chain lengths between 12 and 24 carbon atoms (**Table 1**). Because *trans* hexadecenoic acid (*t*-16:1), *trans* octadecenoic acid (*t*-18:1), and *trans* octadecadienoic acid (*ct*, *tc*, *tt*-18:2) were detected in all samples, the value calculated by summing these

trans fatty acids — termed for the purposes of this paper as sum of *trans* fatty acids — was entered into the correlation analysis. A typical chromatogram with identification marks for *t*-16:1, *t*-18:1, and *ct*, *tc*, *tt*-18:2 is shown in **Figure 1**. We used the STATISTI-CAL PACKAGE FOR SOCIAL SCIENCES FOR WINDOWS, release 7.5 (SPSS Inc, Chicago) for the statistical analyses. Linear correlation coefficients were considered statistically significant at P < 0.05.

RESULTS

Values of selected fatty acids in plasma lipid classes are shown in Table 1. Values of n-6 long-chain polyunsaturated fatty acids were highest in phospholipids and lowest in triacyl-glycerols, whereas values of n-3 long-chain polyunsaturated fatty acids were highest in phospholipids and lowest in cholesterol esters. The percentage of *trans* fatty acids was highest in cholesterol esters and lowest in phospholipids.

The linear correlation coefficients between the sum of trans fatty acids and the major polyunsaturated fatty acids and product-to-substrate ratios of long-chain polyunsaturated fatty acid biosynthesis (arachidonic acid:linoleic acid and docosahexaenoic acid: α -linolenic acid) are shown in Table 2. The sum of trans fatty acids was significantly and inversely associated with values of both arachidonic and docosahexaenoic acids in phospholipids, cholesterol esters, and nonesterified fatty acids. In cholesterol esters, the sum of trans fatty acids was significantly and inversely associated with the parent n-6 polyunsaturated fatty acid, linoleic acid. The sum of trans fatty acids was significantly and inversely related to the ratio of arachidonic acid to linoleic acid in phospholipids and to the ratio of docosahexaenoic acid to α -linolenic acid in cholesterol esters (Table 2). There was a significant inverse association between the sum of trans fatty acids and the sum of n-3 plus n-6 long-chain polyunsaturated fatty acids in phospholipids, cholesterol esters, and nonesterified fatty acids (Figure 2). There were no correlations between trans fatty acid values and infant birth weight or birth length.



FIGURE 1. A typical chromatogram of cord blood plasma phospholipids. The numbers over the peaks identify selected fatty acids as follows: 1, 16:0; 2, t-16:1; 3, 16:1n-7; 4, 18:0; 5, t-18:1; 6, 18:1n-9; 7, 18:1n-7; and 8, ct, tc, tt-18:2. The insets show with greater magnification the *trans* fatty acids included when calculating the sum of *trans* fatty acids (t-16:1 + t-18:1 + ct, tc, tt-18:2).

DISCUSSION

The most notable findings of the present study are the significant inverse associations between *trans* isomeric and long-chain polyunsaturated fatty acids in cord blood lipids at the end of apparently normal human pregnancies. Because humans do not synthesize *trans* fatty acids, the *trans* fatty acids detected in cord blood lipids must have originated from the maternal diet. Moreover, it is reasonable to assume that high *trans* fatty acid concentrations in cord blood indicate high maternal dietary intakes of *trans* fatty acids.

Therefore, the data obtained in the present study raise the possibility that maternal exposure to *trans* fatty acids may be inversely related to long-chain polyunsaturated fatty acid status in full-term infants at birth. Because long-chain polyunsaturated fatty acids are important for early postnatal visual and cognitive development (12–15), factors that reduce the availability of longchain polyunsaturated fatty acids at birth are of serious concern. The potential importance of the inverse association between *trans* fatty acids and long-chain polyunsaturated fatty acid status at birth is supported further by the recent observation that for docosahexaenoic acid and arachidonic acid, status at birth is one of the major determinants of postnatal changes in these fatty acids (18).

The cord blood *trans* fatty acid values measured in the present study were well within the range reported in the literature (19–22). For example, in 1994 one of us reported the total *trans* fatty acid concentrations in cord blood phospholipids (0.69%), cholesterol esters (2.02%), and triacylglycerols (1.91%) of German neonates, and these values were similar to those measured in Austrian newborns in the present study. However, correlations between *trans* isomeric and long-chain polyunsaturated fatty

TABLE 2

Linear correlation coefficients (*r*) between percentage contributions (% by wt) of the sum of *trans* fatty acids and the major polyunsaturated fatty acids and product-to-substrate ratios for 20:4n-6 and 22:6n-3 biosynthesis in cord blood lipids of 42 full-term infants¹

	Phospholipids	Cholesterol esters	Triacylglycerols	Nonesterified fatty acids
n-6 PUFAs				
18:2n-6	-0.02	-0.61^{2}	-0.25	-0.27
20:4n-6	-0.56^{2}	-0.52^{2}	-0.01	-0.41^{3}
Sum of n-6 long-chain PUFAs	-0.48^{3}	-0.49^{3}	0.17	-0.33^{4}
Ratio of 20:4n-6 to 18:2n-6	-0.37^{4}	-0.15	0.14	-0.19
n-3 PUFAs				
18:3n-3	0.24	0.22	0.03	-0.20
22:6n-3	-0.48^{3}	-0.36^{4}	-0.23	-0.41^{3}
Sum of n-3 long-chain PUFAs	-0.47^{3}	-0.03	-0.01	-0.41^{3}
Ratio of 22:6n-3 to 18:3n-3	-0.28	-0.39^{4}	0.07	0.03

¹Sum of n – 6 long-chain polyunsaturated fatty acids (PUFAs): 20:2n-6 + 20:3n-6 + 20:4n-6 + 22:4n-6 + 22:5n-6; sum of n – 3 long-chain PUFAs: 20:3n-3 + 22:5n-3 + 22:5n-3 + 22:5n-3 + 22:5n-3.

 $^{2}P < 0.001.$

 ${}^{3}P < 0.01.$

 $^{4}P < 0.05.$

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FIGURE 2. Relations between the sum of *trans* fatty acids (*t*-16:1 + *t*-18:1 + *ct*, *tc*, *tt*-18:2) and the sum of n-3 plus n-6 long-chain polyunsaturated fatty acids (LCP; 20:3n-3 + 20:5n-3 + 22:5n-3 + 22:6n-3 + 20:2n-6 + 20:3n-6 + 20:4n-6 + 22:4n-6 + 22:5n-6) in nonesterified fatty acids (top), cholesterol esters (middle), and phospholipids (bottom) in venous cord blood of 42 full-term infants.

acids were not reported in the aforementioned studies describing cord blood *trans* fatty acids (19–22).

Three previous reports indicated interference of *trans* fatty acids with the availability of long-chain polyunsaturated fatty acids in humans. First, there were significant inverse correlations between total trans fatty acids and both arachidonic and docosahexaenoic acids, but not linoleic and α -linolenic acids, in plasma cholesterol esters and triacylglycerols of 29 premature infants on day 4 of life (10). trans Fatty acids were also significantly and inversely correlated with birth weight but not with gestational age in that study (10). Second, trans fatty acids were significantly and inversely correlated with arachidonic acid but not with linoleic acid in plasma phospholipids of 53 healthy children aged 1-15 y (11). Third, a significant inverse relation was found between plasma triacylglycerol t-18:1 and plasma phospholipid docosahexaenoic acid values in cord blood of 50 full-term infants in a study that was reported in abstract form (23). In the same study, a strong association was found between maternal trans fatty acid intake and both maternal and infant trans fatty acid status at delivery (24).

The inhibiting effect of trans fatty acids on the enzymatic conversion of linoleic acid to arachidonic acid was documented in numerous studies conducted in animal models and in human fibroblasts in vitro (reviewed in 25). More recently, in pregnant rats, high dietary intakes of trans fatty acids were associated with diminished \triangle^6 -fatty acid desaturase activity in liver microsomes and decreased contribution of n-3 long-chain polyunsaturated fatty acids to fetal plasma lipids in the offspring (26). In the present study, trans fatty acids were inversely related not only to long-chain polyunsaturated fatty acids and the product-to-substrate ratios of their biosynthesis but also to the parent n-6 essential fatty acid, linoleic acid, in cholesterol esters. Therefore, the present study did not determine whether the observed inverse association between trans isomeric and long-chain polyunsaturated fatty acids resulted from an interaction between trans fatty acid intake and essential fatty acid intake, or an inhibitory effect of trans isomers on long-chain polyunsaturated fatty acid biosynthesis, or a combination of these factors.

Did the atopic trait of the infants studied influence the inverse association between *trans* isomeric and long-chain polyunsaturated fatty acids? Higher prevalences of childhood asthma and allergy were associated with higher *trans* fatty acid intakes in an extensive epidemiologic study that compared 55 populations (27). In a different study of umbilical cord serum phospholipids, significantly higher concentrations of long-chain polyunsaturated fatty acids were found in infants whose mothers had allergies than in infants whose mothers did not (28). These data do not seem to suggest an atopy-related inverse association between *trans* isomeric and long-chain polyunsaturated fatty acids, and we are not aware of any studies that measured these 2 groups of fatty acids simultaneously in atopy.

In summary, we report a significant inverse correlation between concentrations of *trans* isomeric fatty acids and longchain polyunsaturated fatty acids in venous cord blood lipids of full-term infants. Although the available data do not allow us to reach any firm conclusion on the nutritional safety of *trans* fatty acids in the diets of pregnant women, the results reported here suggest that maternal *trans* fatty acid intake may be inversely associated with infant long-chain polyunsaturated fatty acid status at birth. Further prospective studies and reevaluation of existing databases on the fatty acid composition of cord blood lipids should contribute to a better understanding of the nutritional role of *trans* fatty acids during pregnancy.

REFERENCES

- Carlson SE, Clandinin MT, Cook HW, Emken EA, Filer LJ Jr. *trans* Fatty acids: infant and fetal development. Am J Clin Nutr 1997; 66(suppl):715S–36S.
- Innis SM, King DJ. *trans* Fatty acids in human milk are inversely associated with concentrations of essential *all-cis* n-6 and n-3 fatty acids and determine *trans*, but not n-6 and n-3, fatty acids in plasma lipids of breast-fed infants. Am J Clin Nutr 1999;70:383–90.
- 3. The British Nutrition Foundation. Unsaturated fatty acids. Nutritional and physiological significance. The Report of the British Nutrition Foundation's Task Force. London: Chapman & Hall, 1995:13.
- 4. Stender S, Dyerberg J, Holmer G, Ovesen L, Sandström B. The influence of *trans* fatty acids on health: a report from The Danish Nutrition Council. Clin Sci 1995;88:375–92.
- Position paper on *trans* fatty acids. ASCN/AIN Task Force on *trans* Fatty Acids. American Society for Clinical Nutrition and American Institute of Nutrition. Am J Clin Nutr 1996;63:663–70.
- Mensink RPM, Katan MB. Effect of dietary *trans* fatty acids on high-density and low-density lipoprotein cholesterol levels in healthy subjects. N Engl J Med 1990;323:439–45.
- trans Fatty acids and coronary heart disease risk. Report of the Expert Panel on trans Fatty Acids and Coronary Heart Disease. Am J Clin Nutr 1995;62(suppl):655S–708S.
- Cook HW. The influence of *trans*-acids on desaturation and elongation of fatty acids in developing brain. Lipids 1981;16:920–6.
- Cook HW, Emken EA. Geometric and positional fatty acid isomers interact differently with desaturation and elongation of linoleic and linolenic acids in cultured glioma cells. Biochem Cell Biol 1990; 68:653–60.
- Koletzko B. *trans* Fatty acids may impair biosynthesis of long-chain polyunsaturates and growth in man. Acta Paediatr Scand 1992;81: 302–6.
- Decsi T, Koletzko B. Do *trans* fatty acids impair linoleic acid metabolism in children? Ann Nutr Metab 1995;39:36–41.
- 12. Innis SM. Essential fatty acids in human growth and development. Prog Lipid Res 1991;30:39–103.
- Decsi T, Koletzko B. Polyunsaturated fatty acids in infant nutrition. Acta Paediatr Suppl 1994;395:31–7.

- Carlson SE, Neuringer M. Polyunsaturated fatty acid status and neurodevelopment: a summary and critical analysis of the literature. Lipids 1999;34:171–8.
- Decsi T, Koletzko B. Role of long-chain polyunsaturated fatty acids in early human neurodevelopment. Nutr Neurosci 2000;3:293–306.
- Folch J, Lees M, Sloane-Stanley GH. A simple method for the isolation and purification of total lipids from animal tissues. J Biol Chem 1957;226:497–509.
- 17. Stoffel W, Chu F, Ahrens EH. Analysis of long-chain fatty acids by gas-liquid chromatography. Anal Chem 1959;31:307–8.
- Guesnet P, Pugo-Gunsam P, Maurage C, et al. Blood lipid concentrations of docosahexaenoic and arachidonic acids at birth determine their relative postnatal changes in term infants fed breast milk or formula. Am J Clin Nutr 1999;70:292–8.
- Laryea M, Cieslicki E, Dickmann E, Wendel U. Age-dependent fatty acid composition of erythrocyte membrane phospholipids in healthy children. Z Ernahrungswiss 1990;29:284–94.
- Koletzko B, Müller L. *cis* And *trans*-isomeric fatty acids in plasma lipids of newborn infants and their mothers. Biol Neonat 1990;57: 172–8.
- Decsi T, Koletzko B. Fatty acid composition of plasma lipid classes in healthy subjects from birth to young adulthood. Eur J Pediatr 1994;153:520–5.
- Berghaus T, Demmelmair H, Koletzko B. Fatty acid composition of lipid classes in maternal and cord plasma at birth. Eur J Pediatr 1998;157:763–8.
- Ayagari A, Peeples JM, Carlson SE. Relationship of isomeric fatty acids in human cord blood to n–3 and n–6 status. Pediatr Res 1996; 39:304A (abstr).
- Ayagari A, Peeples JM, Carlson SE. Maternal *trans* fatty acid intake and *trans* fatty acids in plasma of mother-infant pairs. FASEB J 1996;10:A190 (abstr).
- Koletzko B, Decsi T. Metabolic aspects of *trans* fatty acids. Clin Nutr 1997;16:229–37.
- Larqué E, Pérez-Llamas F, Puerta V, et al. Dietary *trans* fatty acids affect docosahexaenoic acid concentrations in plasma and liver but not brain of pregnant and fetal rats. Pediatr Res 2000;47: 278–83.
- Weiland SK, Mutius E, Hüsing A, Asher MI. Intake of *trans* fatty acids and prevalence of asthma and allergies in Europe. Lancet 1999;353:2040–1.
- Yu G, Kjellman N-IM, Björkstén B. Phospholipid fatty acids in cord blood: family history and development of allergy. Acta Paediatr 1996;85:679–83.

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