## Effect of UV-B radiation treatments on growth, physiology and antioxidant systems of cucumber seedlings in artificial climate chamber

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**Abstract:** Ultraviolet radiation (UV-B) radiation is a key environmental signal for plant growth and development. An excess or lack of UV-B can affect plant resistance, yield and quality. However, the appropriate dose of UV-B for cucumber seedlings growth in plant factories is not well understood. In this study, the effect of different doses of UV-B radiation on the growth, physiology and antioxidant systems of cucumber seedlings in an artificial climate chamber was studied. The results showed that UV-B radiation effectively inhibited the elongation of cucumber seedlings by 4.2%-32.0% and decreased soluble protein content in cucumber leaves by 14.2%-28.2%. 3.33  $\mu$ mol/(m<sup>2</sup>·s) UV-B promoted stem diameter growth, soluble sugar content, total ascorbic acid and the superoxide dismutase, peroxidase and catalase activities in cucumber leaves by 13.6%-22.3%, 22.7%-56.7%, 16.9%-23.2%, 23.8%-25.9%, 34.1%-50.4% and 27.4%-36.4%, respectively. However, this UV-B dose had no influence on the net photosynthetic rate of cucumber leaves. Therefore, we conclude that 3.33  $\mu$ mol/(m<sup>2</sup>·s) UV-B is beneficial for growth and increases the resistance of cucumber seedlings in an artificial climate chamber. This study is hoped to provides a theoretical basis for cucumber and other seedling growth under UV-B treatments.

Keywords: photosynthesis; ultraviolet radiation; growth; cucumber seedlings; H<sub>2</sub>O<sub>2</sub>; antioxidant enzyme system; ascorbic acid; artificial climate chamber

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### **0** Introduction

Light is necessary for plant life, serving as both an energy source for photosynthesis and a signal for development and growth. Furthermore, light quality is a crucial variable for the growth and development of plants, and among the different light wavelengths, UV-B (280 315 nm) is a particularly important environmental factor.

Many studies have reported that excess UV-B has a negative effect on the growth and physiological metabolism of crops, including corn, wheat, rice, and soybeans<sup>[1-3]</sup>. However, due to the covering materials (e.g., plastic film or glass) used in different light shields and filters (photosynthetic active radiation transmittance is 80-90% and

UV-B transmittance is 15-30%), standard light components are not sufficient for greenhouse vegetable cultivation<sup>[4-5]</sup>. Under such circumstances, vegetable can display excessive growth<sup>[6]</sup>, weak growth, poor resistance<sup>[7]</sup>, low yield<sup>[8]</sup> and poor quality<sup>[8-10]</sup>.

Recently, several studies have examined the effects of supplemental UV-B on vegetable quality under greenhouse conditions. These studies showed that UV-B can induce secondary metabolites, which are beneficial for both plants and humans. Wang et al. reported that 0.22  $\mu$ mol/(m<sup>2</sup>·s) of supplemental UV-B radiation could improve fruit quality in tomatoes in winter plastic greenhouses<sup>[4]</sup>. Luthriaand Krizek reported that phenolic acid content in tomatoes was approximately 20% higher under +UV conditions compared with UV conditions<sup>[11]</sup>. Chen et al. reported that 6.91-10.37  $\mu$ mol/(m<sup>2</sup>·s) supplemental UV-B radiation could increase ascorbic acid (AsA) content in pakchoi leaves<sup>[8]</sup>. Hou et al. reported the effect of  $4.15 \,\mu \text{mol}/(\text{m}^2 \cdot \text{s})$  UV-B on photosynthesis and antioxidant enzyme activity in cucumber seedlings<sup>[12-13]</sup>. Sun et al. reported the effect of 0.86 and 4.15  $\mu$ mol/(m<sup>2</sup>·s) UV-B on the growth and photosynthesis of cucumber seedlings, but the results were not consistent with those of previous reports<sup>[12,14]</sup>.

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Plant factories are believed to solve the problems of cultivated land limitation and enable vegetable to grow all year. Plant factories can be established by controlling temperature, nutrient supply, light quality and other factors. At present, red light and blue light are widely used in plant factories<sup>[15-16]</sup>. However, the effect of different doses of UV-B radiation on the growth and antioxidant systems of cucumber seedlings in plant factories has been poorly described.

In the present study, we determined the effects of different doses of UV-B radiation on the growth, physiology and antioxidant systems of cucumber seedlings, and we aimed to determine the dose of UV-B that is beneficial for cucumber seedling growth in plant factories.

#### **1** Materials and methods

#### 1.1 Plant materials and growth conditions

Cucumber seeds (Cucumis sativus L. cv. Chinese long 9930) were germinated on September 28, 2015. The germinated seeds were transferred to a greenhouse at the Institute of Vegetables and Flowers at the Chinese Academy of Agricultural Sciences (Beijing, China, 39.9°N, 116.5°E). Uniform seedlings were transplanted into 7 L pots filled with commercial substrate (Shandong, China) when the third leaf had expanded. When the sixth leaf had expanded, the plants were moved to a controlled chamber with a 14 h photoperiod, 28/20 °C, 60% relative humidity and 120  $\mu$ mol/(m<sup>2</sup>·s) photon flux density (400-700 nm) supplemented with high-pressure sodium lamps from 6:00 AM to 8:00 PM. The experiment was a completely randomized block design with three replicates. Except for the treatment content, the same local management practices were applied in all treatments.

#### 1.2 Experimental design

After the cucumber seedlings were moved to a controlled chamber under 14 h photoperiod, 28/20 °C, 60% relative humidity and 120  $\mu$ mol/(m<sup>2</sup>·s) photon flux density (400-700 nm) supplemented with high-pressure sodium lamps from 6:00 AM to 8:00 PM for three days (on October 26, 2015), they were exposed to biologically effective UV-B irradiance for 4 h (11:00 AM-2:00 PM). The experiments included 5 different doses: 0(CK), 1.67(T1), 3.33(T2), 5.01(T3) and  $6.67(T4) \mu mol/(m^2 \cdot s)$  (Table 1). The supplemental UV-B was applied using Philips TL20 W/01 RS tubes (311-313 nm spectrum peak, Philips) (Fig.1). The tubes were suspended at different distances above the plant canopy so that the doses of UV-B radiation could be adjusted every 3 days. Samples were collected on 0 d (October 26, 2015), 7 d (November 2, 2015), 14 d (November 9, 2015), 21 d (November 16, 2015) and 28 d (November 23, 2015) after the treatment.

### 1.3 Growth parameters

The growth parameters of 4 independent cucumber seedlings for each treatment were determined on 0, 7, 14, 21 and 28 d after the start of the treatment<sup>[17]</sup>. Plant height was measured from the base of the stem to the tip of the stem. Stem diameter was measured at the base of the stem using a

caliper. The second, third and fourth leaves counted from bottom to top were selected to determine leaf photosynthesis parameters using a LI-6400 portable photosynthesis system (Li-Cor 6400XT, Lincoln, NE, USA). Measurements were taken in a controlled chamber within a 2 h interval (08:00-10:00), and red and blue LEDs were selected as the light source. The set values of photosynthetic photon flux density, CO<sub>2</sub> concentration (Ca), air temperature, relative humidity (RH) and air flow rate inside sample chamber were 800  $\mu$ mol/(m<sup>2</sup>·s), 400  $\mu$ mol/s, 28 °C, 60% and 400  $\mu$ mol/s, respectively.

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Treatment	Supplemental UV-B dose/ $(\mu \text{mol} \cdot (\text{m}^{-2} \cdot \text{s}^{-1}))$	Supplemental UV-B time (hour per day)					
СК	0	4 (11:00 AM-2:00 PM)					
T1	1.67	4 (11:00 AM-2:00 PM)					
T2	3.33	4 (11:00 AM-2:00 PM)					
Т3	5.01	4 (11:00 AM-2:00 PM)					
T4	6.67	4 (11:00 AM-2:00 PM)					
output/(a.u.)	50 275 300 325	350 375 400					
2	Wavelength/	mm					

Fig.1 Spectrum of the Philips TL20 W/01 RS tubes

The dry weight of 3 independent cucumber seedlings for each treatment was determined 28 d after the start of the treatment<sup>[18]</sup>. Plant samples were dried in an oven at 105 °C for at least 30 min and then stored at 80 °C for at least 4 days before being weighed.

### 1.4 Physiological parameter measurements

For measurement of physiological responses, three fully expanded leaves were collected from each plant, and the leaves of three plants were mixed as one replicate. The collected leaves were measured immediately and stored at 4 °C for no more than 24 h. Soluble protein concentration was determined using a Coomassie brilliant blue staining protein assay kit (Suzhou, China). Soluble sugar, hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and malondialdehyde (MDA) content were measured according to the methods of Wang<sup>[6]</sup>.

## 1.5 Antioxidant system measurements

For analysis of antioxidant system responses, the sampling method is consistent with 2.4. The collected leaves were immediately frozen in liquid nitrogen and stored at -80 °C until analysis. Superoxide dismutase (SOD) activity was assayed using the nitro blue tetrazolium (NBT) inhibition protocol designed by Wang<sup>[6]</sup>. Peroxidase (POD) activity was determined using a POD assay kit (Suzhou, China). Catalase (CAT) activity was determined using a CAT assay kit (Suzhou, China). Total AsA content was assayed using the spectrophotometric method described by Wang<sup>[6]</sup>.

#### 1.6 Statistical analysis

The data were analyzed using the statistical analysis software program SPSS. Statistically significant differences among means were determined using Duncan's multiple range test at a significance level of P<0.05.

### 2 Results

# 2.1 Effects of different doses of UV-B radiation on the growth characteristics of cucumber seedlings

The growth characteristics of the cucumber seedling under different doses of UV-B radiation are shown in Fig 2. plant height decreased as UV-B radiation dose and treatment time increased. Specifically, plant height decreased by 4.2%-16.0%, 12.2%-22.6%, 18.9%-24.0% and 27.0%-32.0% for T1, T2, T3 and T4, respectively (Fig. 2a). Stem diameter in the T2 group was significantly increased from days7 to 28 and was significantly increased on day 7 for T1 and T3 (Fig.2b).

We found that the net photosynthesis in cucumber leaves under low and medium doses of UV-B radiation (1.67 or 3.33  $\mu$ mol/(m<sup>2</sup>·s)) did not change significantly between days 14 and 28, whereas photosynthesis was significantly reduced under high doses of UV-B radiation (5.01 or 6.67  $\mu$ mol/(m<sup>2</sup>·s)) between days 7 and 28 (Fig.2c). However, the effects of different doses of UV-B radiation on the dry weight of the cucumber seedlings were minimal (Fig.2d).



Note: Mean values with the same letters are not significantly different using Duncan's multiple range test at P<0.05, the same below.

Fig.2 Effects of different UV-B radiation doses on plant height, stem diameter, net photosynthetic rate and dry weight

# **2.2** Effects of different doses of UV-B radiation on the physiological characteristics of cucumber seedlings

The physiological characteristics of the cucumber seedlings under different doses of UV-B radiation are shown in Figure 3. The results demonstrated that the soluble protein content of cucumber seedling leaves exposed to UV-B radiation decreased (Fig.3a). The level of soluble sugar in the T2 treatment group was higher than the control from day 7 to 28, whereas the level of soluble sugar for the T4 treatment group was lower than the control from day 7 to 28. In addition, the soluble sugar content of the T3 treatment group was higher than the control from day 7 to 14, although it was lower than the control on day 28. Overall, these

results indicate that 3.33  $\mu$ mol/(m<sup>2</sup>·s) UV-B radiation is beneficial for the accumulation of soluble sugars in cucumber seedling leaves.

The levels of  $H_2O_2$  and MDA showed similar trends (Fig.3c, d). The content of  $H_2O_2$  in T1, T3 and T4 was significantly higher than that in the control during the whole treatment period, and the content of T2  $H_2O_2$  was not significantly different from that of CK. In addition, the content of  $H_2O_2$  in T4 was significantly higher than that in other treatments. In plants, MDA is the most abundant aldehydic lipid breakdown product. The level of MDA in T3 and T4 was higher than that in the control on day 7. The level of MDA in T1, T2, T3 and T4 was significantly higher than that in the control on day 7. The level of MDA in T1, T2, T3 and T4 was significantly higher than that in the control on day 7. The level of MDA in T1, T2, T3 and T4 was significantly higher than that in the control, and the control, and the control on day 14. The level of MDA in T3 and T4 was significantly higher than that in the control, and the control, and the control of MDA in T2 was not significantly different from that in the control from day 21 to 28 (Fig.3d).



Fig.3 Effects of different UV-B radiation doses on soluble content, soluble sugar content, hydrogen peroxide content and malondialdehyde content

## 2.3 Effects of different doses of UV-B radiation on the antioxidant systems of cucumber seedlings

We found that UV-B radiation could change superoxide dismutase (SOD), peroxidase (POD) and catalase (CAT) activities in cucumber seedling leaves (Fig.4).



Fig.4 Effects of different UV-B radiation doses on activity of SOD, POD, CAT and total AsA content

SOD, POD and CAT activities in the T1, T2 and T3 treatment groups were markedly increased compared with the control on day 7, whereas SOD and POD activities in theT4 treatment group were significantly decreased on day 7(Fig.4a, b). By contrast, SOD, POD and CAT activities in seedlings exposed to the low and medium doses of UV-B radiation (T1 and T2) increased, while the 5.01  $\mu$ mol/(m<sup>2</sup>·s) UV-B treatment (T3) resulted in no significant changes. The 6.67  $\mu$ mol/(m<sup>2</sup>·s) UV-B radiation treatment decreased SOD and POD activities in leaves compared with the control. Furthermore, total AsA content in the cucumber seedling leaves was significantly higher in the T1, T2 and T3 treatment groups compared with the control. However, at the highest dose of UV-B radiation, the total AsA content decreased significantly.

## **3** Discussion

Several studies have shown that UV-B radiation inhibits plant elongation<sup>[19]</sup>. As expected, the elongation of cucumber seedlings was effectively inhibited by UV-B radiation in our study. The effects of UV-B radiation on plant stem diameters<sup>[20]</sup>, net photosynthesis<sup>[21]</sup> and dry weight<sup>[22]</sup> have also been reported. We also found that medium intensity UV-B (3.33  $\mu$ mol/(m<sup>2</sup>·s)) favored stem diameter growth, whereas high intensity UV-B (5.54 or 6.67  $\mu$ mol/(m<sup>2</sup>·s)) led to decrease net photosynthesis significantly. By contrast, low and medium intensity UV-B (1.67 or 3.33  $\mu$ mol/(m<sup>2</sup>·s)) did not affect net photosynthesis. We found low, medium and high intensity UV-B radiation did not affect the dry weight of the plants, we speculate that there was no significant difference due to too short treatment time.

UV-B radiation can damage biological macromolecules such as proteins and DNA<sup>[23-25]</sup>. In our study, the results showed that UV-B radiation reduced soluble protein content in cucumber leaves. Previous studies have also indicated that UV-B radiation can reduce soluble sugar content in plant tissues. We found that soluble sugar content increased under low and medium intensity UV-B radiation (1.67 or  $3.33 \,\mu$ mol/(m<sup>2</sup>·s)) and decreased under the higher intensity UV-B treatments (particularly the 6.67  $\mu$ mol/(m<sup>2</sup>·s) UV-B treatment).

Jenkins reported that UV-B promoted the synthesis of  $H_2O_2$  in plants<sup>[26]</sup>. Our results showed that three doses of UV-B radiation induced  $H_2O_2$ , synthesis, except for the medium intensity UV-B (3.33  $\mu$ mol/(m<sup>2</sup>·s)). We hypothesized that UV-B could increase  $H_2O_2$  content. However, 3.33  $\mu$ mol/(m<sup>2</sup>·s) activated antioxidant systems and increased SOD, POD, CAT activities, and thus,  $H_2O_2$  content under 3.33  $\mu$ mol/(m<sup>2</sup>·s) UV-B showed no differences compared to the control. In addition, we found that the SOD, CAT and POD enzyme activities increased significantly under 3.33  $\mu$ mol/(m<sup>2</sup>·s) UV-B. MDA levels are an indicator of the extent of membrane lipid peroxidation in plants. In general, MDA content in most UV-B radiation-treated seedlings was higher than in the control group, with the exception of the T2 group (3.33  $\mu$ mol/(m<sup>2</sup>·s) UV-B

treatment), consistent with the changes in  $H_2O_2$  content. These results indicated that membrane damage occurred in cucumber seedlings under 1.67, 5.01 and 6.67  $\mu$ mol/(m<sup>2</sup>·s) UV-B.

Reactive oxygen species (ROS), such as H<sub>2</sub>O<sub>2</sub>, are important signal molecules that induce multiple responses, both biotic and abiotic, in plants following environmental stress<sup>[27-28]</sup>. Following an increase in the ROS concentration, the antioxidant system will be activated in plants to eliminate excessive ROS. However, when the ROS levels are too high, these molecules will cause irreversible damage to plants<sup>[29-30]</sup>. In the present study, the activities of the SOD, CAT and POD enzymes, which are involved in the antioxidant defense system, increased significantly under 1.67 and 3.33  $\mu$ mol/(m<sup>2</sup>·s) UV-B radiation. Similarly, AsA, a non-enzymatic antioxidant molecule, also showed increased levels under 1.67, 3.33 and 5.01  $\mu$ mol/(m<sup>2</sup>·s) UV-B radiation. By contrast, the activities of the antioxidant enzymes and AsA levels decreased under 6.67  $\mu$ mol/(m<sup>2</sup>·s) UV-B, this perhaps indicating that the antioxidant system had broken down. In summary, our results suggest that cucumber seedlings are not injured by 3.33  $\mu$ mol/(m<sup>2</sup>·s) UV-B and in fact show increased resistance based on the induction of secondary metabolites and activation of the antioxidant system. In addition, 1.67  $\mu$ mol/(m<sup>2</sup>·s) UV-B caused repairable damage to cucumber seedlings. By contrast, we observed clear evidence of damage in plants treated with high intensity UV-B.

In general, we found that low and medium intensity UV-B treatments are beneficial to plant growth due to enhanced resistance, whereas high intensity UV-B can damage macromolecules such as proteins and nucleic acids. Therefore, it is necessary to provide a suitable dose of UV-B radiation for vegetable production under greenhouse conditions.

In this report, the effects of different doses UV-B on the physiological characteristics of cucumber seedlings were studied. A further study could assess the effects of long-term UV-B on the yield and quality of cucumbers and determine the appropriate UV-B intensity for different cucumber growing periods. These findings are of important practical value. And how does UV-B activate the antioxidant system (such as ascorbic acid metabolism) in leaves, whether it is related to reactive oxygen species, and we are doing further research.

#### 4 Conclusions

UV-B is a key environmental signal for plant growth and development. An excess or lack of UV-B can affect plant resistance. However, the appropriate dose of UV-B for cucumber seedlings growth in plant factories is not well understood. The major aim of the current study was to determine the suitable UV-B doses for cucumber growth in a plant factory. This study showed that the growth of cucumber seedlings is promoted by  $3.33 \ \mu mol/(m^2 \cdot s)$  UV-B, and  $3.33 \ \mu mol/(m^2 \cdot s)$  UV-B increased the antioxidase activities and AsA content of cucumber seedlings. By contrast, these characteristics were inhibited by 6.67  $\mu$ mol/(m<sup>2</sup>·s) UV-B. Furthermore, 3.33  $\mu$ mol/(m<sup>2</sup>·s) UV-B had no significant influence on H<sub>2</sub>O<sub>2</sub> and MDA content in cucumber leaves, whereas 5.01 or 6.67  $\mu$ mol/(m<sup>2</sup>·s) UV-B led to an increase in MDA and H<sub>2</sub>O<sub>2</sub> levels. The results of this study indicate that 3.33  $\mu$ mol/(m<sup>2</sup>·s) UV-B promoted the growth and improved the stress resistance of cucumber plants in a plant factory.

#### [Reference]

- Teramura A H, Sullivan J H, Lydon, J. Effects of UV B radiation on soybean yield and seed quality: A 6-year field study [J]. Physiologia Plantarum, 1990, 80(1): 5-11.
- [2] Correia C M, Torres-Pereira M S, Torres-Pereira J M G. Growth, photosynthesis and UV-B absorbing compounds of Portuguese Barbela wheat exposed to Ultraviolet-B radiation[J]. Environmental Pollution, 1999, 104(3): 383-388.
- [3] Feng H, Li S, Xue L, et al. The interactive effects of enhanced UV-B radiation and soil drought on spring wheat[J]. South African Journal of Botany, 2007, 73(3): 429-434.
- [4] Wang Y L, Wang X L, Yue M. Effects of supplementary radiation of UV-B and red light on fruit quality of tomato in winter plastic greenhouse[J]. Acta Botanica Boreali-Occidentalia Sinica, 1999, 20(4): 590-595. (in Chinese with English abstract)
- [5] Wang S H, Kong Y, Chen Q J, et al. The effects of different light qualities on cucumber fruit quality and yield in greenhouse[J]. Chinese Journal of Eco-Agriculture, 2006, 14(04): 119–121. (in Chinese with English abstract)
- [6] Xu J J, Li J, Feng X H, et al. Effects of enhanced UV-radiation on morphological and physiological characters of welsh onion[J]. Chinese Agricultural Science Bulletin, 2008, 24(11): 307-309. (in Chinese with English abstract)
- [7] Wargent J J, Jordan B R. From ozone depletion to agriculture: Understanding the role of UV radiation in sustainable crop production[J]. New Phytologist, 2013, 197(4): 1058–1076.
- [8] Chen L, Wu Z, Jiang F L, et al. Effects of Ultraviolet-B radiation on growth, yield and quality of pakchoi (*Brassica campestris* ssp. chinensis)[J]. Journal of Plant Resources and Environment, 2008, 17(1): 43-47.
- [9] Lee M J, Son J E, Oh M M. Growth and phenolic compounds of Lactuca sativa L. grown in a closed-type plant production system with UV-A, -B, or -C lamp[J]. Journal of the Science of Food and Agriculture, 2014, 94(2): 197–204.
- [10] Yao X Q, Chu J Z, He X L, et al. The changes in quality ingredients of Qi chrysanthemum flowers treated with elevated UV-B radiation at different growth stages[J]. Journal of Photochemistry and Photobiology B: Biology, 2015, 146: 18-23.
- [11] Luthria D L, Mukhopadhyay S, Krizek D T. Content of total phenolics and phenolic acids in tomato (*Lycopersicon esculentum* Mill.) fruits as influenced by cultivar and solar UV radiation[J]. Journal of Food Composition and Analysis, 2006, 19(8): 771-777.

- Hou F J, Ben G Y. Effect of UV-B radiation on physiological properties in soybean and cucumber seedlings[J]. Chinese Journal of Applied and Environmental Biology, 1999, 5(5): 455-458. (in Chinese with English abstract)
- [13] Hou F J, Li G, Ben G Y. Effects of enhanced Ultraviolet-B radiation on growth, photosynthesis and respiration of cucumis sativus leaves on different positions[J]. Chinese Journal of Applied and Environmental Biology, 2001, 7(4): 321-326. (in Chinese with English abstract)
- [14] Sun L H, Li Z H, Duan L S, et al. Effect of UV-B radiation on the growth and photosynthesis of cucumber (*Cucumis* sativus L.) seedlings[J]. Acta Agriculturae Boreali-Sinica, 2006, 21(6): 79-82. (in Chinese with English abstract)
- [15] Li K, Li Z P, Yang Q C. Improving light distribution by zoom lens for electricity savings in a plant factory with light-emitting diodes[J]. Frontiers in Plant Science, 2016, 7(28): Article92
- [16] Lin K H, Huang M Y, Huang W D, et al. The effects of red, blue, and white light-emitting diodes on the growth, development, and edible quality of hydroponically grown lettuce (*Lactuca sativa* L. var. capitata)[J]. Scientia Horticulturae, 2013, 150(2): 86–91.
- [17] Rybus-Zając M. Kubiś J. Effect of UV-B radiation on antioxidative enzyme activity in cucumber cotyledons[J]. Acta Biologica Cracoviensia Series Botanica, 2011, 52(2): 97-102.
- [18] Xu W D, Liu X Y, Jiao X L, et al. Different ratio of red and blue LED regulation growth and development of cucumber seedlings[J]. Plant Physiology Journal, 2015, 51(8): 1273-1279. (in Chinese with English abstract)
- [19] Randriamanana T R, Lavola A, Julkunen-Tiitto R. Interactive effects of supplemental UV-B and temperature in European aspen seedlings: Implications for growth, leaf traits, phenolic defense and associated organisms[J]. Plant Physiology and Biochemistry, 2015, 93(3): 84–93.
- [20] Liu B, Liu X B, Li Y S, et al. Effects of enhanced UV-B radiation on seed growth characteristics and yield components in soybean[J]. Field Crops Research, 2013, 154(3): 158-163.
- [21] Yu G H, Li W, Yuan Z Y, et al. The effects of enhanced UV-B radiation on photosynthetic and biochemical activities in super-high-yield hybrid rice Liangyoupeijiu at the reproductive stage[J]. Photosynthetica, 2013, 51(1): 33-44.
- [22] Comont D, Winters A, Gomez L D, et al. Latitudinal variation in ambient UV-B radiation is an important determinant of Loliumperenne forage production, quality, and digestibility[J]. Journal of Experimental Botany, 2013, 64(8): 2193-2204.
- [23] Fujimori N, Suzuki N, Nakajima Y, et al. Plant DNA-damage repair/toleration 100 protein repairs UV-B-induced DNA damage[J]. Dna Repair, 2014, 21(3): 171–176.

- [24] Singh J, Dwivedi A, Mujtaba S F, et al. Ambient UV-B exposure reduces the binding of ofloxacin with bacterial DNA gyrase and induces DNA damage mediated apoptosis[J]. International Journal of Biochemistry & Cell Biology, 2016, 73: 111-126.
- [25] Spetea C, Éva H, Vass I. The quinone electron acceptors are not the main sensitizers of UV-B induced protein damage in isolated photosystem II reaction centre and core complexes[J]. Plant Science, 1996, 115(2): 207–215.
- [26] Jenkins G I. Signal transduction in responses to UV-B radiation[J]. Annual Review of Plant Biology, 2009, 60(1): 407-431.
- [27] Foyer C H, Noctor G. Redox sensing and signaling

associated with reactive oxygen in chloroplasts, peroxisomes and mitochondria[J]. Physiologia Plantarum, 2003, 119(3): 355–364.

- [28] Foyer C H, Noctor G. Oxidant and antioxidant signaling in plants: A re-evaluation of the concept of oxidative stress in a physiological context[J]. Plant, Cell & Environment, 2005, 28(8): 1056-1071.
- [29] Blokhina O, Virolainen E, Fagerstedt K V. Antioxidants, oxidative damage and oxygen deprivation stress: A review[J]. Annals of Botany, 2003, 91(2): 179-194.
- [30] Neill S, Desikan R, Hancock, J. Hydrogen peroxide signaling[J]. Current Opinion in Plant Biology, 2002, 5(5): 388-395.

## UV-B 对人工气候室内黄瓜苗期生长、生理及 抗氧化系统的影响

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**摘 要:**紫外线(UV-B)是植物生长发育的关键信号因子。过量或缺少UV-B都会影响作物的抗性、产量和品质。 然而,目前植物工厂中适宜黄瓜生长的UV-B强度尚不明确。以黄瓜(*Cucumis sativus* L.)苗期植株为材料,研究不同强度UV-B对人工气候室内黄瓜苗期植株生长、生理和抗氧化系统的影响。结果表明:与对照相比,UV-B处理黄瓜植株高度降低4.2%~32.0%,叶片中可溶性蛋白含量降低14.2%~28.2%。3.33 μmol/(m<sup>2</sup>·s) UV-B处理植株茎粗增加13.6%~22.3%,叶片中可溶性糖的含量增加22.7%~56.7%,同时激活抗氧化系统,超氧化物歧化酶(SOD)、过氧化物酶(POD)、过氧化氢酶(CAT)活性分别提高16.9%~23.2%,23.8%~25.9%,34.1%~50.4%,抗坏血酸含量增加27.4%~36.4%。由此可知,3.33 μmol/(m<sup>2</sup>·s) UV-B 有利于人工气候室中黄瓜苗期植株的生长发育、抗氧化酶活性提高及抗氧化物质生成。

关键词:光合作用;紫外光;生长;黄瓜植株;过氧化氢;抗氧化酶系统;抗坏血酸;人工气候室